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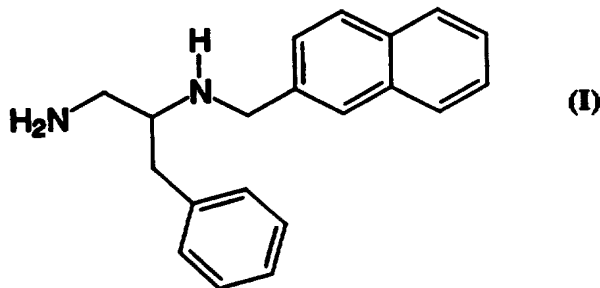
## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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**08/349,025**      **2 December 1994 (02.12.94)**      **US**(71) Applicant: **SYNAPTIC PHARMACEUTICAL CORPORATION [US/US]; 215 College Road, Paramus, NJ 07652 (US).**(72) Inventors: **GERALD, Christophe, P., G.; 204-B Union Street, Ridgewood, NJ 07450 (US). WALKER, Mary, W.; 9 Spruce Street, Elmwood Park, NJ 07407 (US). BRANCHEK, Theresa; 518 Standish Road, Teaneck, NJ 07666 (US). WEINSHANK, Richard, L.; 268 Vandelinda Avenue, Teaneck, NJ 07666 (US).**(74) Agent: **WHITE, John, P.; Cooper & Dunham L.L.P., 1185 Avenue of the Americas, New York, NY 10036 (US).**(81) Designated States: **AM, AT, AU, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LT, LU, LV, MD, MG, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TT, UA, UG, UZ, VN, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG), ARIPO patent (KE, LS, MW, SD, SZ, UG).****Published***With international search report.**Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.*(54) Title: **METHODS OF MODIFYING FEEDING BEHAVIOR, COMPOUNDS USEFUL IN SUCH METHODS, AND DNA ENCODING A HYPOTHALAMIC ATYPICAL NEUROPEPTIDE Y/PEPTIDE YY RECEPTOR (Y5)**

## (57) Abstract

This invention provides methods of modifying feeding behavior, including increasing or decreasing food consumption, e.g., in connection with treating obesity, bulimia or anorexia. These methods involve administration of selective agonists or antagonists or the Y5 receptor. One such antagonist has structure (I). In addition, this invention provides an isolated nucleic acid molecule encoding a Y5 receptor, an isolated Y5 receptor protein, vectors comprising an isolated nucleic acid molecule encoding a Y5 receptor, cells comprising such vectors, antibodies directed to the Y5 receptor, nucleic acid probes useful for detecting nucleic acid encoding Y5 receptors, antisense oligonucleotides complementary to any unique sequences of a nucleic acid molecule which encodes a Y5 receptor, and nonhuman transgenic animals which express DNA a normal or a mutant Y5 receptor.



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- 1 -

5 METHODS OF MODIFYING FEEDING BEHAVIOR, COMPOUNDS USEFUL  
IN SUCH METHODS, AND DNA ENCODING A HYPOTHALAMIC ATYPICAL  
NEUROPEPTIDE Y/PEPTIDE YY RECEPTOR (Y5)

10 This application is a continuation-in-part of U.S. Serial  
No. 08/349,025, filed December 2, 1994, the contents of  
which are hereby incorporated by reference into the  
subject application.

Background of the Invention

15 Throughout this application, various references are  
referred to within parentheses. Disclosures of these  
publications in their entireties are hereby incorporated  
by reference into this application to more fully describe  
the state of the art to which this invention pertains.  
Full bibliographic citation for these references may be  
found at the end of this application, preceding the  
20 sequence listing and the claims.

Neuropeptide Y (NPY) is a member of the pancreatic  
polypeptide family with widespread distribution  
throughout the mammalian nervous system. NPY and its  
25 relatives (peptide YY or PYY, and pancreatic polypeptide  
or PP) elicit a broad range of physiological effects  
through activation of at least five G protein-coupled  
receptor subtypes known as Y1, Y2, Y3, Y4 (or PP), and  
the "atypical Y1". The role of NPY as the most powerful  
30 stimulant of feeding behavior yet described is thought to  
occur primarily through activation of the hypothalamic  
"atypical Y1" receptor. This receptor is unique in that  
its classification was based solely on feeding behavior  
data, rather than radioligand binding data, unlike the  
35 Y1, Y2, Y3, and Y4 (or PP) receptors, each of which were  
described previously in both radioligand binding and  
functional assays. Applicants now report the use of a  
<sup>125</sup>I-PYY-based expression cloning technique to isolate a  
rat hypothalamic cDNA encoding an "atypical Y1" receptor  
40 referred to herein as the Y5 subtype. Applicants also

-2-

report the isolation and characterization of a Y5 homolog from human hippocampus. Protein sequence analysis reveals that the Y5 receptor belongs to the G protein-coupled receptor superfamily. Both the human and rat homolog display  $\leq 42\%$  identity in transmembrane domains with the previously cloned "Y-type" receptors. Rat brain localization studies using in situ hybridization techniques verified the existence of Y5 receptor mRNA in rat hypothalamus. Pharmacological evaluation revealed the following similarities between the Y5 and the "atypical Y1" receptor. 1) Peptides bound to the Y5 receptor with a rank order of potency identical to that described for the feeding response:  $\text{NPY} \geq \text{NPY}_{2-36} = \text{PYY} = [\text{Leu}^{31}, \text{Pro}^{34}]\text{NPY} \gg \text{NPY}_{13-36}$ . 2) The Y5 receptor was negatively coupled to cAMP accumulation, as had been proposed for the "atypical Y1" receptor. 3) Peptides activated the Y5 receptor with a rank order of potency identical to that described for the feeding response. 4) The reported feeding "modulator"  $[\text{D-Trp}^{32}]\text{NPY}$  bound selectively to the Y5 receptor and subsequently activated the receptor. 5) Both the Y5 and the "atypical Y1" receptors were sensitive to deletions or modifications in the midregion of NPY and related peptide ligands. These data support the identity of the Y5 receptor as the previously described "atypical Y1", and furthermore indicate a role for the Y5 receptor as a potential target in the treatment of obesity, metabolism, and appetite disorders.

The peptide neurotransmitter neuropeptide Y (NPY) is a 36 amino acid member of the pancreatic polypeptide family with widespread distribution throughout the mammalian nervous system. NPY is considered to be the most powerful stimulant of feeding behavior yet described (Clark et al., 1984; Levine and Morley, 1984; Stanley and Leibowitz, 1984). Direct injection into the hypothalamus of satiated rats, for example, can increase food intake

-3-

up to 10-fold over a 4-hour period (Stanley et al., 1992). The role of NPY in normal and abnormal eating behavior, and the ability to interfere with NPY-dependent pathways as a means to appetite and weight control, are areas of great interest in pharmacological and pharmaceutical research (Sahu and Kalra, 1993; Dryden et al., 1994). Any credible means of studying or controlling NPY-dependent feeding behavior, however, must necessarily be highly specific as NPY can act through at least 5 pharmacologically defined receptor subtypes to elicit a wide variety of physiological functions (Dumont et al., 1992). It is therefore vital that knowledge of the molecular biology and structural diversity of the individual receptor subtypes be understood as part of a rational drug design approach to develop subtype selective compounds. A brief review of NPY receptor pharmacology is summarized below and also in Table 1.

**TABLE 1: Pharmacologically defined receptors for NPY and related pancreatic polypeptides.**

Rank orders of affinity for key peptides (NPY, PYY, PP, [Leu<sup>31</sup>,Pro<sup>34</sup>]NPY, NPY<sub>2-36</sub>, and NPY<sub>13-36</sub>) are based on previously reported binding and functional data (Schwartz et al., 1990; Wahlestedt et al., 1991; Dumont et al., 1992; Wahlestedt and Reis, 1993). Data for the Y2 receptor were disclosed in U.S. patent application 08/192,288 filed on 2/3/94, currently pending, the foregoing contents of which are hereby incorporated by reference. Data for the Y4 receptor were disclosed in U.S. patent application 08/176,412 filed on 12/28/93, currently pending, the foregoing contents of which are hereby incorporated by reference. Missing peptides in the series reflect a lack of published information.

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TABLE 1

Receptor	Affinity (pK <sub>i</sub> or pEC <sub>50</sub> )					
	11 to 10	10 to 9	9 to 8	8 to 7	7 to 6	< 6
Y1	NPY PYY [Leu <sup>31</sup> ,Pro <sup>34</sup> ] NPY		NPY <sub>2-36</sub>	NPY <sub>13-36</sub>	PP	
Y2		PYY NPY NPY <sub>2-36</sub>	NPY <sub>13-36</sub>			[Leu <sup>31</sup> , Pro <sup>34</sup> ] NPY PP
Y3		NPY	[Pro <sup>34</sup> ] NPY	NPY <sub>13-36</sub> PP		PYY
Y4	PP	PYY [Leu <sup>31</sup> ,P ro <sup>34</sup> ] NPY	NPY NPY <sub>2-36</sub>	NPY <sub>13-36</sub>		
atypical Y1 (feeding)		PYY NPY NPY <sub>2-36</sub> [Leu <sup>31</sup> ,P ro <sup>34</sup> ] NPY		NPY <sub>13-36</sub>		

NPY Receptor Pharmacology

NPY receptor pharmacology has historically been based on structure/activity relationships within the pancreatic polypeptide family. The entire family includes the namesake pancreatic polypeptide (PP), synthesized primarily by endocrine cells in the pancreas; peptide YY (PYY), synthesized primarily by endocrine cells in the gut; and NPY, synthesized primarily in neurons (Michel, 1991; Dumont et al., 1992; Wahlestedt and Reis, 1993). All pancreatic polypeptide family members share a compact

-5-

structure involving a "PP-fold" and a conserved C-terminal hexapeptide ending in Tyr<sup>36</sup> (or Y<sup>36</sup> in the single letter code). The striking conservation of Y<sup>36</sup> has prompted the reference to the pancreatic polypeptides' receptors as "Y-type" receptors (Wahlestedt et al., 1987), all of which are proposed to function as seven transmembrane-spanning G protein-coupled receptors (Dumont et al., 1992).

10 The Y1 receptor recognizes NPY ≥ PYY >> PP (Grundemar et al., 1992). The receptor requires both the N- and the C-terminal regions of the peptides for optimal recognition. Exchange of Gln<sup>34</sup> in NPY or PYY with the analogous residue from PP (Pro<sup>34</sup>), however, is well-tolerated. The

15 Y1 receptor has been cloned from a variety of species including human, rat and mouse (Larhammar et al, 1992; Herzog et al, 1992; Eva et al, 1990; Eva et al, 1992). The Y2 receptor recognizes PYY ~ NPY >> PP and is relatively tolerant of N-terminal deletion (Grundemar et

20 al., 1992). The receptor has a strict requirement for structure in the C-terminus (Arg<sup>33</sup>-Gln<sup>34</sup>-Arg<sup>35</sup>-Tyr<sup>36</sup>-NH<sub>2</sub>); exchange of Gln<sup>34</sup> with Pro<sup>34</sup>, as in PP, is not well tolerated. The Y2 receptor has recently been cloned (disclosed in US patent application Serial No.

25 08/192,288, filed February 3, 1994). The Y3 receptor is characterized by a strong preference for NPY over PYY and PP (Wahlestedt et al., 1991). [Pro<sup>34</sup>]NPY is reasonably well tolerated even though PP, which also contains Pro<sup>34</sup>, does not bind well to the Y3 receptor. This receptor

30 (Y3) has not yet been cloned. The Y4 receptor (disclosed in U.S. patent application Serial No. 08/176,412, filed December 28, 1993) binds PP > PYY > NPY. Like the Y1, the Y4 requires both the N- and the C-terminal regions of the peptides for optimal recognition (Synaptic Y4 patent).

35 The "atypical Y1" or "feeding" receptor was defined exclusively by injection of several pancreatic polypeptide analogs into the paraventricular nucleus of

-6-

the rat hypothalamus which stimulated feeding behavior with the following rank order:  $\text{NPY}_{2-36} \geq \text{NPY} \sim \text{PYY} \sim [\text{Leu}^{31}, \text{Pro}^{34}]\text{NPY} > \text{NPY}_{13-36}$  (Kalra et al., 1991; Stanley et al., 1992). The profile is similar to that of a Y1-like receptor except for the anomalous ability of  $\text{NPY}_{2-36}$  to stimulate food intake with potency equivalent or better than that of NPY. A subsequent report in *J. Med. Chem.* by Balasubramaniam and co-workers (1994) showed that feeding can be regulated by  $[\text{D-Trp}^{32}]\text{NPY}$ . While this peptide was presented as an NPY antagonist, the published data at least in part support a stimulatory effect of  $[\text{D-Trp}^{32}]\text{NPY}$  on feeding.  $[\text{D-Trp}^{32}]\text{NPY}$  thereby represents another diagnostic tool for receptor identification. In contrast to other NPY receptor subtypes, the "feeding" receptor has never been characterized for peptide binding affinity in radioligand binding assays and the fact that a single receptor could be responsible for the feeding response has been impossible to validate in the absence of an isolated receptor protein; the possibility exists, for example, that the feeding response could be a composite profile of Y1 and Y2 subtypes.

Applicants now report the isolation by expression cloning of a novel Y-type receptor from a rat hypothalamic cDNA library, along with its pharmacological characterization, in situ localization, and human homologues. The data provided link this newly-cloned receptor subtype, from now on referred to as the Y5 subtype, to the "atypical Y1" feeding response. This discovery therefore provides a novel approach, through the use of heterologous expression systems, to develop a subtype selective antagonist for obesity and other indications.

Applicants further report the isolation of a canine Y5 receptor. In addition, applicants report the discovery of chemical compounds which bind selectively to the Y5 receptor of the present invention and which act as



-7-

antagonists of the Y5 receptor. Several of the compounds were further shown to inhibit food intake in rats.

5 The treatment of disorders or diseases associated with the inhibition of the Y5 receptor subtype, especially diseases caused by eating disorders such as obesity, bulimia nervosa, diabetes, and dislipidimia may be effected by administration of compounds which bind  
10 selectively to the Y5 receptor and inhibit the activation of the Y5 receptor. Furthermore, any disease states in which the Y5 receptor subtype is involved, for example, memory loss, epileptic seizures, migraine, sleep disturbance, and pain may also be treated using compounds which bind selectively to the Y5 receptor.

Summary of the Invention

5 This invention provides a method of modifying feeding behavior of a subject which comprises administering to the subject an amount of a compound which is a Y5 receptor agonist or antagonist effective to increase or decrease consumption of food by the subject so as to thereby modify feeding behavior of the subject.

10 This invention provides a method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a non-peptidyl compound which is a Y5 receptor antagonist effective to inhibit the activity of the subject's Y5 receptor, wherein the  
15 binding of the compound to the human receptor is characterized by a  $K_i$  less than 100 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.

20 This invention provides a method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a peptidyl compound which is a Y5 receptor antagonist effective to inhibit the activity of the subject's Y5 receptor, wherein the compound's  
25 binding to the human Y5 receptor is characterized by a  $K_i$  less than 10 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.

30 This invention provides a method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a non-peptidyl compound which is a Y5 receptor agonist effective to increase the activity of the subject's Y5 receptor, wherein (a) the binding of the compound to the human Y5 receptor is characterized by a  $K_i$  less than 100 nanomolar when measured in the presence  
35 of  $^{125}\text{I}$ -PYY; and (b) the binding of the compound to any other human Y-type receptor is characterized by a  $K_i$  greater than 1000 nanomolar when measured in the presence

of  $^{125}\text{I}$ -PYY.

5 This invention provides a method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a non-peptidyl compound which is a Y5 receptor agonist effective to increase the activity of the subject's Y5 receptor, wherein (a) the binding of the compound to the human Y5 receptor is characterized by a  $K_i$  less than 1 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY; and (b) the compound's binding to any other human Y-type receptor is characterized by a  $K_i$  greater than 100 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.

15 This invention provides a method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a peptidyl compound which is a Y5 receptor agonist effective to increase the activity of the subject's Y5 receptor, wherein (a) the binding of the compound to the human Y5 receptor is characterized by a  $K_i$  less than 1 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY; and (b) the binding of the compound to any other human Y-type receptor is characterized by a  $K_i$  greater than 25 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.

25 This invention provides a method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a peptidyl compound which is a Y5 receptor agonist effective to increase the activity of the subject's Y5 receptor, wherein (a) the binding of the compound to the human Y5 receptor is characterized by a  $K_i$  less than 0.1 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY; and (b) the binding of the compound to any other human Y-type receptor is characterized by a  $K_i$  greater than 1 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.

-10-

5 This invention provides a method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a peptidyl compound which is a Y5 receptor agonist effective to increase the activity of the subject's Y5 receptor, wherein (a) the binding of the compound to the human Y5 receptor is characterized by a  $K_i$  less than 0.01 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY; and (b) the binding of the compound to any other human Y-type receptor is characterized by a  $K_i$  greater than 1 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.

10 This invention provides an isolated nucleic acid encoding a Y5 receptor. This invention also provides an isolated Y5 receptor protein. This invention provides a vector comprising the above-described nucleic acid.

15 This invention provides a plasmid which comprises the regulatory elements necessary for expression of DNA in a mammalian cell operatively linked to the DNA encoding the human Y5 receptor as to permit expression thereof designated pcEXV-hY5 (ATCC Accession No. 75943).

20 This invention provides a plasmid which comprises the regulatory elements necessary for expression of DNA in a mammalian cell operatively linked to the DNA encoding the rat Y5 receptor as to permit expression thereof designated pcEXV-rY5 (ATCC Accession No. 75944).

25 This invention provides a mammalian cell comprising the above-described plasmid or vector.

30 This invention provides a nucleic acid probe comprising a nucleic acid of at least 15 nucleotides capable of specifically hybridizing with a unique sequence included within the sequence of a nucleic acid encoding a Y5 receptor.

-11-

This invention provides an antisense oligonucleotide having a sequence capable of specifically hybridizing to mRNA encoding a Y5 receptor so as to prevent translation of the mRNA.

5

This invention provides an antibody directed to a Y5 receptor.

10

This invention provides a pharmaceutical composition comprising an amount of the oligonucleotide effective to reduce activity of a human Y5 receptor by passing through a cell membrane and binding specifically with mRNA encoding a human Y5 receptor in the cell so as to prevent its translation and a pharmaceutically acceptable carrier capable of passing through a cell membrane.

15

This invention provides a pharmaceutical composition comprising an amount of an antagonist effective to reduce the activity of a human Y5 receptor and a pharmaceutically acceptable carrier.

20

This invention provides a pharmaceutical composition comprising an amount of an agonist effective to increase activity of a Y5 receptor and a pharmaceutically acceptable carrier.

25

This invention provides the above-described pharmaceutical composition which comprises an amount of the antibody effective to block binding of a ligand to the Y5 receptor and a pharmaceutically acceptable carrier.

30

This invention provides a transgenic nonhuman mammal expressing DNA encoding a human Y5 receptor.

35

This invention also provides a method for determining whether a ligand can specifically bind to a Y5 receptor

-12-

5 which comprises contacting a cell transfected with and expressing DNA encoding the Y5 receptor with the ligand under conditions permitting binding of ligands to such receptor, detecting the presence of any such ligand specifically bound to the Y5 receptor, and thereby determining whether the ligand specifically binds to the Y5 receptor.

10 This invention provides a method for determining whether a ligand can specifically bind to a Y5 receptor which comprises preparing a cell extract from cells transfected with and expressing DNA encoding the Y5 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with the ligand under  
15 conditions permitting binding of ligands to such receptor, detecting the presence of the ligand specifically bound to the Y5 receptor, and thereby determining whether the ligand specifically binds to the Y5 receptor.

20 This invention provides a method for determining whether a ligand is a Y5 receptor agonist which comprises contacting a cell transfected with and expressing nucleic acid encoding a human Y5 receptor with the ligand under  
25 conditions permitting activation of the Y5 receptor, detecting an increase in Y5 receptor activity, and thereby determining whether the ligand is a human Y5 receptor agonist.

30 This invention provides a method for determining whether a ligand is a Y5 receptor antagonist which comprises contacting a cell transfected with and expressing DNA encoding a Y5 receptor with the ligand in the presence of a known Y5 receptor agonist, such as PYY or NPY, under  
35 conditions permitting the activation of the Y5 receptor, detecting a decrease in Y5 receptor activity, and thereby determining whether the ligand is a Y5 receptor

-13-

antagonist.

This invention provides a method of screening a plurality of chemical compounds not known to bind to a Y5 receptor to identify a compound which specifically binds to the Y5 receptor, which comprises (a) contacting a cell transfected with and expressing DNA encoding the Y5 receptor with a compound known to bind specifically to the Y5 receptor; (b) contacting the preparation of step (a) with the plurality of compounds not known to bind specifically to the Y5 receptor, under conditions permitting binding of compounds known to bind the Y5 receptor; (c) determining whether the binding of the compound known to bind to the Y5 receptor is reduced in the presence of the compounds, relative to the binding of the compound in the absence of the plurality of compounds; and if so (d) separately determining the binding to the Y5 receptor of each compound included in the plurality of compounds, so as to thereby identify the compound which specifically binds to the Y5 receptor.

This invention provides a method of screening a plurality of chemical compounds not known to bind to a Y5 receptor to identify a compound which specifically binds to the Y5 receptor, which comprises (a) preparing a cell extract from cells transfected with and expressing DNA encoding the Y5 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with a compound known to bind specifically to the Y5 receptor; (b) contacting preparation of step (a) with the plurality of compounds not known to bind specifically to the Y5 receptor, under conditions permitting binding of compounds known to bind the Y5 receptor; (c) determining whether the binding of the compound known to bind to the Y5 receptor is reduced in the presence of the compounds, relative to the binding of the compound in the absence of the plurality of compounds; and if so (d) separately

-14-

determining the binding to the Y5 receptor of each compound included in the plurality of compounds, so as to thereby identify the compound which specifically binds to the Y5 receptor.

5

This invention provides a method of screening a plurality of chemical compounds not known to activate a Y5 receptor to identify a compound which activates the Y5 receptor which comprises (a) contacting a cell transfected with and expressing the Y5 receptor with the plurality of compounds not known to bind specifically to the Y5 receptor, under conditions permitting activation of the Y5 receptor; (b) determining whether the activity of the Y5 receptor is increased in the presence of the compounds; and if so (c) separately determining whether the activation of the Y5 receptor is increased by each compound included in the plurality of compounds, so as to thereby identify the compound which activates the Y5 receptor.

20

This invention provides a method of screening a plurality of chemical compounds not known to activate a Y5 receptor to identify a compound which activates the Y5 receptor which comprises (a) preparing a cell extract from cells transfected with and expressing DNA encoding the Y5 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with the plurality of compounds not known to bind specifically to the Y5 receptor, under conditions permitting activation of the Y5 receptor; (b) determining whether the activity of the Y5 receptor is increased in the presence of the compounds; and if so (c) separately determining whether the activation of the Y5 receptor is increased by each compound included in the plurality of compounds, so as to thereby identify the compound which activates the Y5 receptor.



-15-

This invention provides a method of screening a plurality of chemical compounds not known to inhibit the activation of a Y5 receptor to identify a compound which inhibits the activation of the Y5 receptor, which comprises (a) contacting a cell transfected with and expressing the Y5 receptor with the plurality of compounds in the presence of a known Y5 receptor agonist, under conditions permitting activation of the Y5 receptor; (b) determining whether the activation of the Y5 receptor is reduced in the presence of the plurality of compounds, relative to the activation of the Y5 receptor in the absence of the plurality of compounds; and if so (c) separately determining the inhibition of activation of the Y5 receptor for each compound included in the plurality of compounds, so as to thereby identify the compound which inhibits the activation of the Y5 receptor.

This invention provides a method of screening a plurality of chemical compounds not known to inhibit the activation of a Y5 receptor to identify a compound which inhibits the activation of the Y5 receptor, which comprises (a) preparing a cell extract from cells transfected with and expressing DNA encoding the Y5 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with the plurality of compounds in the presence of a known Y5 receptor agonist, under conditions permitting activation of the Y5 receptor; (b) determining whether the activation of the Y5 receptor is reduced in the presence of the plurality of compounds, relative to the activation of the Y5 receptor in the absence of the plurality of compounds; and if so (c) separately determining the inhibition of activation of the Y5 receptor for each compound included in the plurality of compounds, so as to thereby identify the compound which inhibits the activation of the Y5 receptor.

This invention provides a method of screening drugs to

-16-

5 identify drugs which specifically bind to a Y5 receptor on the surface of a cell which comprises contacting a cell transfected with and expressing DNA encoding a Y5 receptor with a plurality of drugs under conditions permitting binding of drugs to the Y5 receptor, determining those drugs which specifically bind to the transfected cell, and thereby identifying drugs which specifically bind to the Y5 receptor.

10 This invention provides a method of screening drugs to identify drugs which act as agonists of a Y5 receptor which comprises contacting a cell transfected with and expressing DNA encoding a Y5 receptor with a plurality of  
15 drugs under conditions permitting the activation of a functional Y5 receptor response, determining those drugs which activate such receptor in the cell, and thereby identify drugs which act as Y5 receptor agonists.

20 This invention provides a method of screening drugs to identify drugs which act as Y5 receptor antagonists which comprises contacting cells transfected with and expressing DNA encoding a Y5 receptor with a plurality of drugs in the presence of a known Y5 receptor agonist, such as PYY or NPY, under conditions permitting the  
25 activation of a functional Y5 receptor response, determining those drugs which inhibit the activation of the receptor in the mammalian cell, and thereby identifying drugs which act as Y5 receptor antagonists.

30 This invention provides a method of treating an abnormality in a subject, wherein the abnormality is alleviated by the inhibition of a Y5 receptor which comprises administering to a subject an effective amount of Y5 receptor antagonist.

35 This invention provides a method of treating an abnormality in a subject wherein the abnormality is

-17-

alleviated by the activation of a Y5 receptor which comprises administering to a subject an effective amount of a Y5 receptor agonist.

5 This invention provides a method for diagnosing a predisposition to a disorder associated with the activity of a specific human Y5 receptor allele which comprises:

10 a. obtaining DNA of subjects suffering from the disorder; performing a restriction digest of the DNA with a panel of restriction enzymes; c. electrophoretically separating the resulting DNA fragments on a sizing gel; d. contacting the resulting gel with a nucleic acid probe capable of specifically hybridizing to DNA encoding a human Y5 receptor and labelled with a detectable marker;

15 e. detecting labelled bands which have hybridized to the DNA encoding a human Y5 receptor labelled with a detectable marker to create a unique band pattern specific to the DNA of subjects suffering from the disorder; f. preparing DNA obtained for diagnosis by

20 steps a-e; and g. comparing the unique band pattern specific to the DNA of subjects suffering from the disorder from step e and the DNA obtained for diagnosis from step f to determine whether the patterns are the same or different and to diagnose thereby predisposition

25 to the disorder if the patterns are the same.

This invention provides a method of preparing the isolated Y5 receptor which comprises: a. inserting nucleic acid encoding Y5 receptor in a suitable vector

30 which comprises the regulatory elements necessary for expression of the nucleic acid operatively linked to the nucleic acid encoding a Y5 receptor; b. inserting the resulting vector in a suitable host cell so as to obtain a cell which produces the Y5 receptor; c. recovering the

35 receptor produced by the resulting cell; and d. purifying the receptor so recovered.

-18-

**Brief Description of the Figures**

**Figure 1** Competitive displacement of  $^{125}\text{I}$ -PYV on membranes from rat hypothalamus. Membranes were incubated with  $^{125}\text{I}$ -PYV and increasing concentrations of peptide competitors.  $\text{IC}_{50}$  values corresponding to 50% displacement were determined by nonlinear regression analysis. Data are representative of at least two independent experiments.  $\text{IC}_{50}$  values for these compounds are listed separately in Table 2.

**Figure 2** Competitive displacement of  $^{125}\text{I}$ -PYV<sub>3-36</sub> on membranes from rat hypothalamus. Membranes were incubated with  $^{125}\text{I}$ -PYV<sub>3-36</sub> and increasing concentrations of peptide competitors.  $\text{IC}_{50}$  values corresponding to 50% displacement were determined by nonlinear regression analysis. Data are representative of at least two independent experiments.  $\text{IC}_{50}$  values for these compounds are listed separately in Table 2.

**Figure 3** Nucleotide sequence of the rat hypothalamic Y5 cDNA clone (Seq. I.D. No 1). Initiation and stop codons are underlined. Only partial 5' and 3' untranslated sequences are shown.

**Figure 4** Corresponding amino acid sequence of the rat hypothalamic Y5 cDNA clone (Seq. I.D. No. 2).

**Figure 5** Nucleotide sequence of the human hippocampal Y5 cDNA clone (Seq. I.D. No. 3). Initiation and stop codons are underlined. Only partial 5' and 3' untranslated sequences are shown.

**Figure 6** Corresponding amino acid sequence of the human hippocampal Y5 cDNA clone (Seq. I.D. No. 4).

**Figure 7** A-E. Comparison of coding nucleotide sequences

-19-

between rat hypothalamic Y5 (top row) and human hippocampal Y5 (bottom row) cDNA clones (84.1% nucleotide identity). F-G. Comparison of deduced amino acid sequence between rat hypothalamic Y5 (top row) and human  
5 hippocampal Y5 (Bottom row) cDNA clones (87.2% overall and 98.8% transmembrane domain identities).

Figure 8 Comparison of the human Y5 receptor deduced amino acid sequence with those of the human Y1, Y2, Y4  
10 sequences. Solid bars, the seven putative membrane-spanning domains (TM I-VII). Shading, identities between receptor sequences.

Figure 9 Equilibrium binding of  $^{125}\text{I}$ -PYV to membranes from  
15 COS-7 cells transiently expressing rat Y5 receptors. Membranes were incubated with  $^{125}\text{I}$ -PYV for the times indicated, in the presence or absence of 300 nM human NPY. Specific binding, B, was plotted against time, t, to obtain the maximum number of equilibrium binding  
20 sites,  $B_{\text{max}}$ , and observed association rate,  $K_{\text{obs}}$ , according to the equation,  $B = B_{\text{max}} * (1 - e^{-(k_{\text{obs}} * t)})$ . Binding is shown as the percentage of total equilibrium binding,  $B_{\text{max}}$ , determined by nonlinear regression analysis. Each point represents a triplicate determination.

25 Figure 10 Saturable equilibrium binding of  $^{125}\text{I}$ -PYV to membranes from COS-7 cells transiently expressing rat Y5 receptors. Membranes were incubated with  $^{125}\text{I}$ -PYV ranging in concentration from 0.4 pM to 2.7 nM, in the presence  
30 or absence of 300 nM human NPY. Specific binding, B, was plotted against the free  $^{125}\text{I}$ -PYV concentration, [L], to obtain the maximum number of saturable binding sites,  $B_{\text{max}}$ , and the  $^{125}\text{I}$ -PYV equilibrium dissociation constant,  $K_d$ , according to the binding isotherm,  $B = B_{\text{max}}[L]/([L] + K_d)$ .  
35 Specific binding is shown. Data are representative of three independent experiments, with each point measured in triplicate.

-20-

Figure 11 Competitive displacement of  $^{125}\text{I}$ -PYV from COS-7 cells transiently expressing rat Y5 receptors. Membranes were incubated with  $^{125}\text{I}$ -PYV and increasing concentrations of peptide competitors.  $\text{IC}_{50}$  values corresponding to 50% displacement were determined by nonlinear regression analysis and converted to  $K_i$  values according to the equation,  $K_i = \text{IC}_{50} / (1 + [\text{L}] / K_d)$ , where  $[\text{L}]$  is the  $^{125}\text{I}$ -PYV concentration and  $K_d$  is the equilibrium dissociation constant of  $^{125}\text{I}$ -PYV. Data are representative of at least two independent experiments. Rank orders of affinity for these and other compounds are listed separately in Table 4.

Figure 12 Inhibition of forskolin-stimulated cAMP accumulation in intact 293 cells stably expressing rat Y5 receptors. Functional data were derived from radioimmunoassay of cAMP in 293 cells stimulated with  $10\ \mu\text{M}$  forskolin over a 5 minute period. Rat/human NPY was tested for agonist activity at concentrations ranging from  $0.03\ \text{pM}$  to  $0.3\ \mu\text{M}$  over the same period. The  $\text{EC}_{50}$  value corresponding to 50% maximal activity was determined by nonlinear regression analysis. The data shown are representative of three independent experiments.

Figure 13 Schematic diagrams of coronal sections through the rat brain, illustrating the distribution of NPY Y5 receptor mRNA, as visualized microscopically in sections dipped in liquid emulsion. The sections are arranged from rostral (A) to caudal (H). Differences in silver grain density over individual neurons in a given area are indicated by the hatching gradient. The full definitions for the abbreviations are as follows:

Aco = anterior cortical amygdaloid nucleus;  
AD = anterodorsal thalamic nucleus;  
APT = anterior pretectal nucleus;  
Arc = arcuate hypothalamic nucleus;

-21-

BLA = basolateral amygdaloid nucleus anterior;  
CA3 = field CA3 of Ammon's horn, hippocampus;  
CeA = central amygdaloid nucleus;  
Cg = cingulate cortex;  
5 CL = centrolateral thalamic nucleus;  
CM = central medial thalamic nucleus  
DG = dentate gyrus, hippocampus;  
DMH = dorsomedial hypothalamic nucleus;  
DR = dorsal raphe;  
10 GiA = gigantocellular reticular nucleus, alpha;  
HDB = nucleus horizontal limb diagonal band;  
InG = intermediate gray layer superior  
colliculus;  
LC = locus coeruleus;  
15 LH = lateral hypothalamic area;  
MePV = medial amygdaloid nucleus,  
posteroventral;  
MVe = medial vestibular nucleus;  
MHb = medial habenular nucleus;  
20 MPN = medial preoptic nucleus;  
PAG = periaqueductal gray;  
PaS = parasubiculum;  
PC = paracentral thalamic nucleus;  
PCRtA = parvocellular reticular nucleus, alpha;  
25 Pe = periventricular hypothalamic nucleus;  
PrS = presubiculum;  
PN = pontine nuclei;  
PVH = paraventricular hypothalamic nucleus;  
PVHmp = paraventricular hypothalamic nucleus,  
30 medial parvicellular part  
PVT = paraventricular thalamic nucleus;  
Re = reunions thalamic nucleus;  
RLi = rostral linear nucleus raphe;  
RSG = retrosplenial cortex;  
35 SCN = suprachiasmatic nucleus;  
SNC = substantia nigra, pars compacta; and  
SON = supraoptic nucleus.

-22-

Figure 14 Partial Nucleotide sequence of the canine Y5 cDNA clone beginning immediately upstream of TM III to the stop codon (underlined), (Seq. I.D. No 5). Only partial 3' untranslated sequence is shown.

5

Figure 15 Corresponding amino acid sequence of the canine Y5 cDNA clone (Seq. I.D. No. 6).

10 Figure 16 A. Northern blot analysis of various rat tissues. B. Northern blot analysis of various human brain areas: amygdala, caudate nucleus, corpus callosum, hippocampus, whole brain, substantia nigra, subthalamic nucleus, and thalamus. C. Northern blot analysis of  
15 various additional human brain areas: cerebellum, cerebral cortex, medula, spinal cord, occipital lobe, frontal lobe, temporal lobe, and putamen. Hybridization was done under conditions of high stringency, as described in Experimental Details.

20 Figure 17 Southern blot analysis of human or rat genomic DNA encoding the Y5 receptor subtype. Hybridization was done under conditions of high stringency, as described in Experimental Details.

25 Figure 18 Time course for equilibrium binding of  $^{125}\text{I}$ -Leu<sup>31</sup>,Pro<sup>34</sup>-PYY to the rat Y5 receptor. Membranes were incubated with 0.08 nM radioligand at room temperature for the length of time indicated in binding buffer containing either 10 mM Na<sup>+</sup> or 138 mM Na<sup>+</sup>.

30

Figure 19 Guanine Nucleotide Modulation of Y5 Peptide Binding. Human or rat Y5 receptors transiently expressed in COS-7 cell membranes, or human Y5 receptors stably expressed in LM(tk-) cell membranes, were incubated with  
35 0.08 nM  $^{125}\text{I}$ -PYY and increasing concentrations of Gpp(NH)p as indicated under standard binding assay conditions. Radioligand binding is reported as cpm, efficiency = 0.8.



-23-

For the human Y5 in LM(tk-) (0.007 mg membrane protein/sample), the maximum  $\Delta$  cpm = -2343. Given a specific activity of 2200 Ci/mmol, the change in radioligand binding is therefore calculated to be -0.6  
5 fmol/0.007 mg protein = -85 fmol/mg membrane protein.

Figure 20 NPY-Dependent Inhibition of Forskolin Stimulated cAMP Accumulation by Cloned Y5 Receptors. Intact cells stably transfected with human or rat Y5  
10 receptors were incubated with forskolin plus a range of human NPY concentrations as indicated. A representative experiment is shown for each receptor system ( $n \geq 2$ ).

Figure 21 Calcium Mobilization: Fura-2 Assay. Cloned  
15 human Y-type receptors in the host cells indicated were screened for intracellular calcium mobilization in response to NPY and related peptides. Representative calcium transients are shown for each receptor system.

20 Figure 22 Illustrates the structure of a compound which binds selectively to the human and rat Y5 receptors.

-24-

Detailed Description of the Invention

Throughout this application, the following standard abbreviations are used to indicate specific nucleotide  
5 bases:

C=cytosine	A=adenine
T=thymine	G=guanine

Furthermore, the term "agonist" is used throughout this  
10 application to indicate any peptide or non-peptidyl  
compound which increases the activity of any of the  
receptors of the subject invention. The term  
"antagonist" is used throughout this application to  
15 indicate any peptide or non-peptidyl compound which  
decreases the activity of any of the receptors of the  
subject invention.

The activity of a G-protein coupled receptor such as a Y5  
20 receptor may be measured using any of a variety of  
appropriate functional assays in which activation of the  
receptor in question results in an observable change in  
the level of some second messenger system, including but  
not limited to adenylate cyclase, calcium mobilization,  
25 inositol phospholipid hydrolysis or guanylyl cyclase.

This invention provides a method of modifying feeding  
behavior of a subject which comprises administering to  
the subject an amount of a compound which is a Y5  
30 receptor agonist or antagonist effective to increase or  
decrease consumption of food by the subject so as to  
thereby modify feeding behavior of the subject. In one  
embodiment, the compound is a Y5 receptor antagonist and  
the amount is effective to decrease the consumption of  
food by the subject. In another embodiment, the compound  
35 is administered in combination with food. In a further  
embodiment, the compound is a Y5 receptor agonist and  
the amount is effective to increase the consumption of

-25-

food by the subject. In another embodiment, the compound is administered with food. The subject may be a vertebrate, a mammal, a human or a canine subject.

5 This invention provides a method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a non-peptidyl compound which is a Y5 receptor antagonist effective to inhibit the activity of the subject's Y5 receptor, wherein the  
10 binding of the compound to the human receptor is characterized by a  $K_i$  less than 100 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY. In one embodiment, the compound has a  $K_i$  less than 5 nanomolar. In another embodiment, the compound has a  $K_i$  less than 1 nanomolar.  
15 In a further embodiment, the binding of the compound to any other human Y-type receptor is characterized by a  $K_i$  greater than 10 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY. In a further embodiment, the binding of the compound to each of the human Y1, human Y2,, and human Y4  
20 receptors is characterized by a  $K_i$  greater than 10 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY. In another embodiment, the binding of the compound to any other human Y-type receptor is characterized by a  $K_i$  greater than 50 nanomolar. In another embodiment, the  
25 binding of the compound to any other human Y-type receptor is characterized by a  $K_i$  greater than 100 nanomolar. In one embodiment, the compound binds to the human Y5 receptor with an affinity greater than ten-fold higher than the affinity with which the compound binds to  
30 any other human Y-type receptor. In another embodiment, the compound binds to the human Y5 receptor with an affinity greater than ten-fold higher than the affinity with which the compound binds to each of the human Y1, human Y2, and human Y4 receptors.

35

In a further embodiment, the feeding disorder is bulimia. The subject may be a vertebrate, a mammal, a human or a

-26-

canine subject.

5 This invention provides a method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a peptidyl compound which is a Y5 receptor antagonist effective to inhibit the activity of the subject's Y5 receptor, wherein the compound's binding to the human Y5 receptor is characterized by a  $K_i$  less than 10 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY. In one embodiment, the compound's binding is characterized by a  $K_i$  less than 1 nanomolar. In another embodiment, the compound's binding to any other human Y-type receptor is characterized by a  $K_i$  greater than 10 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY. In a further embodiment, the compound's binding to each of the human Y1, human Y2, and human Y4 receptors is characterized by a  $K_i$  greater than 10 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY. In another embodiment, the compound's binding to any other human Y-type receptor is characterized by a  $K_i$  greater than 50 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY. In another embodiment, the compound's binding to any other human Y-type receptor is characterized by a  $K_i$  greater than 100 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY. In another embodiment, the compound binds to the human Y5 receptor with an affinity greater than ten-fold higher than the affinity with which the compound binds to any other human Y-type receptor. In one embodiment, the compound binds to the human Y5 receptor with an affinity greater than ten-fold higher than the affinity with which the compound binds to each of the human Y1, human Y2, and human Y4 receptors.

35 In one embodiment of the above-described methods, the feeding disorder is obesity. In another embodiment, the feeding disorder is bulimia. The subject may be a vertebrate, a mammal, a human, or a canine subject.

-27-

This invention provides a method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a non-peptidyl compound which is a Y5 receptor agonist effective to increase the activity of the subject's Y5 receptor, wherein (a) the binding of the compound to the human Y5 receptor is characterized by a  $K_i$  less than 100 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY; and (b) the binding of the compound to any other human Y-type receptor is characterized by a  $K_i$  greater than 1000 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.

In one embodiment, the binding of the compound to the human Y5 receptor is characterized by a  $K_i$  less than 10 nanomolar.

This invention provides a method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a non-peptidyl compound which is a Y5 receptor agonist effective to increase the activity of the subject's Y5 receptor, wherein (a) the binding of the compound to the human Y5 receptor is characterized by a  $K_i$  less than 1 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY; and (b) the compound's binding to any other human Y-type receptor is characterized by a  $K_i$  greater than 100 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.

In one embodiment, the compound binds to the human Y5 receptor with an affinity greater than ten-fold higher than the affinity with which the compound binds to any other human Y-type receptor. In another embodiment, the compound binds to the human Y5 receptor with an affinity greater than ten-fold higher than the affinity with which the compound binds to each of the human Y1, human Y2 and human Y4 receptors.

-28-

In one embodiment, the feeding disorder is anorexia. The subject may be a vertebrate, a mammal, a human, or a canine subject.

5 This invention provides a method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a peptidyl compound which is a Y5 receptor agonist effective to increase the activity of the subject's Y5 receptor, wherein (a) the binding of the  
10 compound to the human Y5 receptor is characterized by a  $K_i$  less than 1 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY; and (b) the binding of the compound to any other human Y-type receptor is characterized by a  $K_i$  greater than 25 nanomolar when measured in the presence of  $^{125}\text{I}$ -  
15 PYY.

This invention provides a method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a peptidyl compound which is a  
20 Y5 receptor agonist effective to increase the activity of the subject's Y5 receptor, wherein (a) the binding of the compound to the human Y5 receptor is characterized by a  $K_i$  less than 0.1 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY; and (b) the binding of the compound to any  
25 other human Y-type receptor is characterized by a  $K_i$  greater than 1 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.

In one embodiment, the binding of the agonist to any  
30 other human Y-type receptor is characterized by a  $K_i$  greater than 10 nanomolar.

This invention provides a method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a peptidyl compound which is a  
35 Y5 receptor agonist effective to increase the activity of the subject's Y5 receptor, wherein (a) the binding of the

-29-

compound to the human Y5 receptor is characterized by a  $K_i$  less than 0.01 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY; and (b) the binding of the compound to any other human Y-type receptor is characterized by a  $K_i$  greater than 1 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.

In one embodiment, the compound binds to the human Y5 receptor with an affinity greater than ten-fold higher than the affinity with which the compound binds to any other human Y-type receptor. In another embodiment, the compound binds to the human Y5 receptor with an affinity greater than ten-fold higher than the affinity with which the compound binds to each of the human Y1, human Y2, and human Y4 receptors.

In one embodiment, the feeding disorder is anorexia. The subject may be a vertebrate, a mammal, a human or a canine subject.

This invention provides for the use of the compounds described herein for the preparation of a pharmaceutical composition, medicament, or drug for modifying feeding behavior of a subject.

This invention provides for the use of the compounds described herein for the preparation of a pharmaceutical composition, medicament, or drug for treating a disorder in which antagonism of the Y5 receptor may be useful, in particular, for treating a feeding disorder such as obesity or bulimia.

This invention provides for the use of the compounds described herein for the preparation of a pharmaceutical composition, medicament, or drug for treating a disorder in which agonism of the Y5 receptor may be useful, in particular, for treating a feeding disorder such as

-30-

anorexia.

5 This invention provides an isolated nucleic acid encoding a Y5 receptor. In an embodiment, the Y5 receptor is a vertebrate or a mammalian Y5 receptor. In an embodiment, the isolated nucleic acid encodes a receptor being characterized by an amino acid sequence in the transmembrane region, which amino acid sequence has 60% homology or higher to the amino acid sequence in the transmembrane region of the human Y5 receptor shown in Figure 6. In another embodiment, the Y5 receptor has substantially the same amino acid sequence as described in Figure 4. In another embodiment, the Y5 receptor has substantially the same amino acid sequence as described in Figure 6.

20 This invention provides the above-described isolated nucleic acid, wherein the nucleic acid is DNA. In an embodiment, the DNA is cDNA. In another embodiment, the DNA is genomic DNA. In still another embodiment, the nucleic acid is RNA. In a separate embodiment, the nucleic acid encodes a human Y5 receptor. In an embodiment, the human Y5 receptor has the amino acid sequence as described in Figure 6.

25 This invention further provides DNA which is degenerate with any of the DNA shown in Figures 3, 5 and 14, which DNA encode Y5 receptors having the amino acid sequences shown in Figures 4, 6, and 15, respectively.

30 This invention also encompasses DNAs and cDNAs which encode amino acid sequences which differ from those of Y5 receptor, but which should not produce phenotypic changes. Alternatively, this invention also encompasses DNAs and cDNAs which hybridize to the DNA and cDNA of the subject invention. Hybridization methods are well known to those of skill in the art.



-31-

The DNA molecules of the subject invention also include DNA coding for polypeptide analogs, fragments or derivatives of antigenic polypeptides which differ from naturally-occurring forms in terms of the identity or location of one or more amino acid residues (deletion analogs containing less than all of the residues specified for the protein, substitution analogs wherein one or more residues specified are replaced by other residues and addition analogs where in one or more amino acid residues is added to a terminal or medial portion of the polypeptides) and which share some or all properties of naturally-occurring forms. These nucleic acids include: the incorporation of codons "preferred" for expression by selected non-mammalian hosts; the provision of sites for cleavage by restriction endonuclease enzymes; and the provision of additional initial, terminal or intermediate DNA sequences that facilitate construction of readily expressed vectors.

The nucleic acids described and claimed herein are useful for the information which they provide concerning the amino acid sequence of the polypeptide and as products for the large scale synthesis of the polypeptide by a variety of recombinant techniques. The nucleic acid is useful for generating new cloning and expression vectors, transformed and transfected prokaryotic and eukaryotic host cells, and new and useful methods for cultured growth of such host cells capable of expression of the polypeptide and related products.

In a separate embodiment, the nucleic acid encodes a rat Y5 receptor. In another embodiment, the rat Y5 receptor has the amino acid sequence shown in Figure 4. In another embodiment, the nucleic acid encodes a canine Y5 receptor. In a further embodiment, the canine Y5 receptor has the amino acid sequence as shown in Figure 15.

-32-

This invention also provides an isolated Y5 receptor protein. In separate embodiments, the Y5 protein may be a human, a rat, or a canine protein.

- 5 This invention provides a vector comprising the above-described nucleic acid.

10 Vectors which comprise the isolated nucleic acid described hereinabove also are provided. Suitable vectors comprise, but are not limited to, a plasmid or a virus. These vectors may be transformed into a suitable host cell to form a host cell vector system for the production of a polypeptide having the biological activity of a Y5 receptor.

15 This invention provides the above-described vector adapted for expression in a bacterial cell which further comprises the regulatory elements necessary for expression of the nucleic acid in the bacterial cell  
20 operatively linked to the nucleic acid encoding the Y5 receptor as to permit expression thereof.

25 This invention provides the above-described vector adapted for expression in a yeast cell which comprises the regulatory elements necessary for expression of the nucleic acid in the yeast cell operatively linked to the nucleic acid encoding the Y5 receptor as to permit expression thereof.

30 This invention provides the above-described vector adapted for expression in an insect cell which comprises the regulatory elements necessary for expression of the nucleic acid in the insect cell operatively linked to the nucleic acid encoding the Y5 receptor as to permit  
35 expression thereof.

In an embodiment, the vector is adapted for expression in

-33-

a mammalian cell which comprises the regulatory elements necessary for expression of the DNA in the mammalian cell operatively linked to the DNA encoding the mammalian Y5 receptor as to permit expression thereof.

5

In a further embodiment, the vector is adapted for expression in a mammalian cell which comprises the regulatory elements necessary for expression of the DNA in the mammalian cell operatively linked to the DNA encoding the human Y5 receptor as to permit expression thereof.

10

In a still further embodiment, the plasmid is adapted for expression in a mammalian cell which comprises the regulatory elements necessary for expression of the DNA in the mammalian cell operatively linked to the DNA encoding the rat Y5 receptor as to permit expression thereof.

15

This invention provides the above-described plasmid adapted for expression in a mammalian cell which comprises the regulatory elements necessary for expression of DNA in a mammalian cell operatively linked to the DNA encoding the mammalian Y5 receptor as to permit expression thereof.

20

25

This invention provides a plasmid which comprises the regulatory elements necessary for expression of DNA in a mammalian cell operatively linked to the DNA encoding the human Y5 receptor as to permit expression thereof designated pcEXV-hY5 (ATCC Accession No. 75943).

30

This plasmid (pcEXV-hY5) was deposited on November 4, 1994 with the American Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, Maryland 20852, U.S.A. under the provisions of the Budapest Treaty for the International Recognition of the Deposit of

35

-34-

Microorganisms for the Purposes of Patent Procedure and was accorded ATCC Accession No. 75943.

5 This invention provides a plasmid which comprises the regulatory elements necessary for expression of DNA in a mammalian cell operatively linked to the DNA encoding the rat Y5 receptor as to permit expression thereof designated pcEXV-rY5 (ATCC Accession No. 75944).

10 This plasmid (pcEXV-rY5) was deposited on November 4, 1994 with the American Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, Maryland 20852, U.S.A. under the provisions of the Budapest Treaty for the International Recognition of the Deposit of  
15 Microorganisms for the Purposes of Patent Procedure and was accorded ATCC Accession No. CRL 75944. This invention provides a plasmid designated Y5-bd-5 (ATCC Accession No. \_\_\_\_\_). This invention also provides a plasmid designated Y5-bd-8 (ATCC Accession No. \_\_\_\_\_).  
20 \_\_\_\_\_). These plasmids were deposited on December 1, 1995 with the American Type Culture Collection (ATCC) 12301 Parklawn Drive, Rockville, Maryland 20852, U.S.A. under the provisions of the Budapest Treaty for the International Recognition of the Deposit of  
25 Microorganisms for the Purposes of Patent Procedure and was accorded ATCC Accession Nos. \_\_\_\_\_ and \_\_\_\_\_, respectively

30 This invention provides a baculovirus designated hY5-BB3 (ATCC Accession No. \_\_\_\_\_) This baculovirus was deposited on November 15, 1995 with the American Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, Maryland 20852, U.S.A. under the provisions of the Budapest Treaty for the International Recognition of  
35 the Deposit of Microorganisms for the Purposes of Patent Procedure and was accorded ATCC Accession No. \_\_\_\_\_.

-35-

This invention provides a mammalian cell comprising the above-described plasmid or vector. In an embodiment, the mammalian cell is a COS-7 cell.

- 5 In another embodiment, the mammalian cell is a 293 human embryonic kidney cell designated 293-rY5-14 (ATCC Accession No. CRL 11757).

10 This cell (293-rY5-14) was deposited on November 4, 1994 with the American Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, Maryland 20852, U.S.A. under the provisions of the Budapest Treaty for the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure and  
15 was accorded ATCC Accession No. CRL 11757.

In a further embodiment, the mammalian cell is a mouse fibroblast (tk-) cell, containing the plasmid pcEXV-hY5 and designated L-hY5-7 (ATCC Accesssion No. CRL-11995).

- 20 In another embodiment, the mammalian cell is a mouse embryonic NIH-3T3 cell containing the plasmid pcEXV-hY5 and designated N-hY5-8 (ATCC Accession No. CRL-11994). These cells were deposited on November 15, 1995 with the American Type Culture Collection (ATCC) 12301 Parklawn  
25 Drive, Rocville, Maryland 20852, U.S.A. under the provisions of the Budapest Treaty for the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure, and were accorded ATCC Accession Nos. CRL-11995 and CRL-11994, respectively.

30 This invention provides a nucleic acid probe comprising a nucleic acid of at least 15 nucleotides capable of specifically hybridizing with a unique sequence included within the sequence of a nucleic acid encoding a Y5  
35 receptor. In an embodiment, the nucleic acid is DNA.

This nucleic acid produced can either be DNA or RNA. As

-36-

used herein, the phrase "specifically hybridizing" means the ability of a nucleic acid to recognize a nucleic acid sequence complementary to its own and to form double-helical segments through hydrogen bonding between  
5 complementary base pairs.

This nucleic acid of at least 15 nucleotides capable of specifically hybridizing with a sequence of a nucleic acid encoding the human Y5 receptors can be used as a  
10 probe. Nucleic acid probe technology is well known to those skilled in the art who will readily appreciate that such probes may vary greatly in length and may be labeled with a detectable label, such as a radioisotope or  
15 fluorescent dye, to facilitate detection of the probe. DNA probes may be produced by insertion of a DNA which encodes the Y5 receptor into suitable vectors, such as plasmids or bacteriophages, followed by transforming into  
20 suitable bacterial host cells, replication in the transformed bacterial host cells and harvesting of the DNA probes, using methods well known in the art. Alternatively, probes may be generated chemically from  
DNA synthesizers.

RNA probes may be generated by inserting the DNA which  
25 encodes the Y5 receptor downstream of a bacteriophage promoter such as T3, T7 or SP6. Large amounts of RNA probe may be produced by incubating the labeled nucleotides with the linearized fragment where it contains an upstream promoter in the presence of the  
30 appropriate RNA polymerase.

This invention also provides a nucleic acid of at least 15 nucleotides capable of specifically hybridizing with a sequence of a nucleic acid which is complementary to  
35 the mammalian nucleic acid encoding a Y5 receptor. This nucleic acid may either be DNA or RNA.

-37-

This invention provides an antisense oligonucleotide having a sequence capable of specifically hybridizing to mRNA encoding a Y5 receptor so as to prevent translation of the mRNA.

5

This invention provides an antisense oligonucleotide having a sequence capable of specifically hybridizing to the genomic DNA of a Y5 receptor.

10 This invention provides an antisense oligonucleotide of Y5 receptor comprising chemical analogues of nucleotides.

This invention provides an antibody directed to a Y5 receptor. This invention also provides an antibody  
15 directed to a human Y5 receptor.

This invention provides a monoclonal antibody directed to an epitope of a human Y5 receptor present on the surface of a Y5 receptor expressing cell.

20

This invention provides a pharmaceutical composition comprising an amount of the oligonucleotide effective to reduce activity of a human Y5 receptor by passing through a cell membrane and binding specifically with mRNA  
25 encoding a human Y5 receptor in the cell so as to prevent its translation and a pharmaceutically acceptable carrier capable of passing through a cell membrane. In an embodiment, the oligonucleotide is coupled to a substance which inactivates mRNA. In another embodiment, the  
30 substance which inactivates mRNA is a ribozyme.

This invention provides the above-described pharmaceutical composition, wherein the pharmaceutically acceptable carrier capable of passing through a cell  
35 membrane comprises a structure which binds to a receptor specific for a selected cell type and is thereby taken up by cells of the selected cell type.

-38-

This invention provides a pharmaceutical composition comprising an amount of an antagonist effective to reduce the activity of a human Y5 receptor and a pharmaceutically acceptable carrier.

5

This invention provides a pharmaceutical composition comprising an amount of an agonist effective to increase activity of a Y5 receptor and a pharmaceutically acceptable carrier.

10

This invention provides the above-described pharmaceutical composition which comprises an amount of the antibody effective to block binding of a ligand to the Y5 receptor and a pharmaceutically acceptable carrier.

15

As used herein, "pharmaceutically acceptable carriers" means any of the standard pharmaceutically acceptable carriers. Examples include, but are not limited to, phosphate buffered saline, physiological saline, water and emulsions, such as oil/water emulsions.

20

This invention provides a transgenic nonhuman mammal expressing DNA encoding a human Y5 receptor.

25

This invention provides a transgenic nonhuman mammal comprising a homologous recombination knockout of the native Y5 receptor.

30

This invention provides a transgenic nonhuman mammal whose genome comprises antisense DNA complementary to DNA encoding a human Y5 receptor so placed as to be transcribed into antisense mRNA which is complementary to mRNA encoding a Y5 receptor and which hybridizes to mRNA encoding a Y5 receptor thereby reducing its translation.

35

This invention provides the above-described transgenic



-39-

nonhuman mammal, wherein the DNA encoding a human Y5 receptor additionally comprises an inducible promoter.

5 This invention provides the transgenic nonhuman mammal, wherein the DNA encoding a human Y5 receptor additionally comprises tissue specific regulatory elements.

In an embodiment, the transgenic nonhuman mammal is a mouse.

10

Animal model systems which elucidate the physiological and behavioral roles of Y5 receptor are produced by creating transgenic animals in which the activity of the Y5 receptor is either increased or decreased, or the amino acid sequence of the expressed Y5 receptor is altered, by a variety of techniques. Examples of these techniques include, but are not limited to: 1) Insertion of normal or mutant versions of DNA encoding a Y5 receptor, by microinjection, electroporation, retroviral transfection or other means well known to those skilled in the art, into appropriate fertilized embryos in order to produce a transgenic animal or 2) Homologous recombination of mutant or normal, human or animal versions of these genes with the native gene locus in transgenic animals to alter the regulation of expression or the structure of these Y5 receptor sequences. The technique of homologous recombination is well known in the art. It replaces the native gene with the inserted gene and so is useful for producing an animal that cannot express native Y5 receptors but does express, for example, an inserted mutant Y5 receptor, which has replaced the native Y5 receptor in the animal's genome by recombination, resulting in underexpression of the transporter. Microinjection adds genes to the genome, but does not remove them, and so is useful for producing an animal which expresses its own and added Y5 receptors, resulting in overexpression of the Y5 receptors.

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One means available for producing a transgenic animal, with a mouse as an example, is as follows: Female mice are mated, and the resulting fertilized eggs are dissected out of their oviducts. The eggs are stored in an appropriate medium such as M2 medium. DNA or cDNA encoding a Y5 receptor is purified from a vector by methods well known in the art. Inducible promoters may be fused with the coding region of the DNA to provide an experimental means to regulate expression of the trans-gene. Alternatively or in addition, tissue specific regulatory elements may be fused with the coding region to permit tissue-specific expression of the trans-gene. The DNA, in an appropriately buffered solution, is put into a microinjection needle (which may be made from capillary tubing using a pipet puller) and the egg to be injected is put in a depression slide. The needle is inserted into the pronucleus of the egg, and the DNA solution is injected. The injected egg is then transferred into the oviduct of a pseudopregnant mouse (a mouse stimulated by the appropriate hormones to maintain pregnancy but which is not actually pregnant), where it proceeds to the uterus, implants, and develops to term. As noted above, microinjection is not the only method for inserting DNA into the egg cell, and is used here only for exemplary purposes.

This invention also provides a method for determining whether a ligand can specifically bind to a Y5 receptor which comprises contacting a cell transfected with and expressing DNA encoding the Y5 receptor with the ligand under conditions permitting binding of ligands to such receptor, detecting the presence of any such ligand specifically bound to the Y5 receptor, and thereby determining whether the ligand specifically binds to the Y5 receptor.

This invention provides a method for determining whether

-41-

a ligand can specifically bind to a human Y5 receptor which comprises contacting a cell transfected with and expressing DNA encoding the human Y5 receptor with the ligand under conditions permitting binding of ligands to such receptor, detecting the presence of any such ligand specifically bound to the human Y5 receptor, and thereby determining whether the ligand specifically binds to the human Y5 receptor.

10 This invention provides a method for determining whether a ligand can specifically bind to a Y5 receptor which comprises contacting a cell transfected with and expressing DNA encoding the Y5 receptor with the ligand under conditions permitting binding of ligands to such  
15 receptor, detecting the presence of any such ligand specifically bound to the Y5 receptor, and thereby determining whether the ligand specifically binds to the human Y5 receptor, such Y5 receptor having substantially the same amino acid sequence shown in Figure 6.

20 This invention provides a method for determining whether a ligand can specifically bind to a Y5 receptor which comprises contacting a cell transfected with and expressing DNA encoding the Y5 receptor with the ligand  
25 under conditions permitting binding of ligands to such receptor, detecting the presence of any such ligand specifically bound to the Y5 receptor, and thereby determining whether the ligand specifically binds to the Y5 receptor, such Y5 receptor being characterized by an  
30 amino acid sequence in the transmembrane region having 60% homology or higher to the amino acid sequence in the transmembrane region of the Y5 receptor shown in Figure 6.

35 This invention provides a method for determining whether a ligand can specifically bind to a Y5 receptor which comprises preparing a cell extract from cells transfected

-42-

with and expressing DNA encoding the Y5 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with the ligand under conditions permitting binding of ligands to such  
5 receptor, detecting the presence of the ligand specifically bound to the Y5 receptor, and thereby determining whether the ligand specifically binds to the Y5 receptor.

10 In separate embodiments of the above-described methods, the Y5 receptor may be a human Y5 receptor, a rat Y5 receptor, or a canine Y5 receptor.

This invention provides a method for determining whether  
15 a ligand can specifically bind to a Y5 receptor which comprises preparing a cell extract from cells transfected with and expressing DNA encoding the Y5 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with the ligand under  
20 conditions permitting binding of ligands to the human Y5 receptor, detecting the presence of the ligand specifically bound to the Y5 receptor, and thereby determining whether the ligand can specifically bind to the Y5 receptor.

25 This invention provides a method for determining whether a ligand can specifically bind to a Y5 receptor which comprises preparing a cell extract from cells transfected with and expressing DNA encoding the Y5 receptor,  
30 isolating a membrane fraction from the cell extract, contacting the membrane fraction with the ligand under conditions permitting binding of ligands to the Y5 receptor, detecting the presence of the ligand specifically bound to the Y5 receptor, and thereby  
35 determining whether the ligand can specifically bind to the Y5 receptor, such Y5 receptor having substantially the same amino acid sequence shown in Figure 6.

-43-

This invention provides a method for determining whether a ligand can specifically bind to a Y5 receptor which comprises preparing a cell extract from cells transfected with and expressing DNA encoding the Y5 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with the ligand under conditions permitting binding of ligands to the Y5 receptor, detecting the presence of the ligand specifically bound to the Y5 receptor, and thereby determining whether the ligand can specifically bind to the Y5 receptor, such Y5 receptor being characterized by an amino acid sequence in the transmembrane region having 60% homology or higher to the amino acid sequence in the transmembrane region of the Y5 receptor shown in Figure 6.

In separate embodiments of the above-described methods, the Y5 receptor may be a human Y5 receptor, a rat Y5 receptor, or a canine Y5 receptor. In one embodiment of the above-described methods, the ligand is not previously known.

This invention further provides a ligand identified by any one of the above-described methods.

This invention provides a method for determining whether a ligand is a Y5 receptor agonist which comprises contacting a cell transfected with and expressing nucleic acid encoding a Y5 receptor with the ligand under conditions permitting activation of a functional Y5 receptor response, detecting a functional increase in Y5 receptor activity, and thereby determining whether the ligand is a Y5 receptor agonist.

This invention provides a method for determining whether a ligand is a Y5 receptor agonist which comprises contacting a cell transfected with and expressing nucleic

-44-

acid encoding a human Y5 receptor with the ligand under conditions permitting activation of the Y5 receptor, detecting an increase in Y5 receptor activity, and thereby determining whether the ligand is a human Y5  
5 receptor agonist.

This invention provides a method for determining whether a ligand is a Y5 receptor agonist which comprises preparing a cell extract from cells transfected with and  
10 expressing DNA encoding the Y5 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with the ligand under conditions permitting the activation of the Y5 receptor, and detecting an increase in Y5 receptor activity, so as to  
15 thereby determine whether the ligand is a Y5 receptor agonist.

This invention provides a method for determining whether a ligand is a Y5 receptor antagonist which comprises  
20 contacting a cell transfected with and expressing DNA encoding a Y5 receptor with the ligand in the presence of a known Y5 receptor agonist, such as PYY or NPY, under conditions permitting the activation of a functional Y5 receptor response, detecting a decrease in Y5 receptor  
25 activity, and thereby determining whether the ligand is a Y5 receptor antagonist.

This invention provides a method for determining whether a ligand is a Y5 receptor antagonist which comprises  
30 contacting a cell transfected with and expressing DNA encoding a Y5 receptor with the ligand in the presence of a known Y5 receptor agonist, such as PYY or NPY, under conditions permitting the activation of the Y5 receptor, detecting a decrease in Y5 receptor activity, and thereby  
35 determining whether the ligand is a Y5 receptor antagonist.

-45-

This invention provides a method for determining whether a ligand is a Y5 receptor antagonist which comprises preparing a cell extract from cells transfected with and expressing DNA encoding the Y5 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with the ligand in the presence of a known Y5 receptor agonist, such as PYY, under conditions permitting the activation of the Y5 receptor, and detecting a decrease in Y5 receptor activity, so as to thereby determine whether the ligand is a Y5 receptor antagonist.

In separate embodiments of the above-described methods the Y5 receptor is a human Y5 receptor, a rat Y5 receptor, or a canine Y5 receptor.

In an embodiment of the above-described methods, the cell is non-neuronal in origin. In a further embodiment, the non-neuronal cell is a COS-7 cell, 293 human embryonic kidney cell, NIH-3T3 cell or LM(tk-) cell.

In one embodiment of the above-described methods, the ligand is not previously known.

This invention provides a Y5 receptor agonist detected by the above-described method. This invention provides a Y5 receptor antagonist detected by the above-described method.

This invention provides a method of screening a plurality of chemical compounds not known to bind to a Y5 receptor to identify a compound which specifically binds to the Y5 receptor, which comprises (a) contacting a cell transfected with and expressing DNA encoding the Y5 receptor with a compound known to bind specifically to the Y5 receptor; (b) contacting the preparation of step (a) with the plurality of compounds not known to bind

-46-

specifically to the Y5 receptor, under conditions permitting binding of compounds known to bind the Y5 receptor; (c) determining whether the binding of the compound known to bind to the Y5 receptor is reduced in the presence of the compounds, relative to the binding of the compound in the absence of the plurality of compounds; and if so (d) separately determining the binding to the Y5 receptor of each compound included in the plurality of compounds, so as to thereby identify the compound which specifically binds to the Y5 receptor.

This invention provides a method of screening a plurality of chemical compounds not known to bind to a Y5 receptor to identify a compound which specifically binds to the Y5 receptor, which comprises (a) preparing a cell extract from cells transfected with and expressing DNA encoding the Y5 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with a compound known to bind specifically to the Y5 receptor; (b) contacting preparation of step (a) with the plurality of compounds not known to bind specifically to the Y5 receptor, under conditions permitting binding of compounds known to bind the Y5 receptor; (c) determining whether the binding of the compound known to bind to the Y5 receptor is reduced in the presence of the compounds, relative to the binding of the compound in the absence of the plurality of compounds; and if so (d) separately determining the binding to the Y5 receptor of each compound included in the plurality of compounds, so as to thereby identify the compound which specifically binds to the Y5 receptor.

This invention provides a method of screening a plurality of chemical compounds not known to activate a Y5 receptor to identify a compound which activates the Y5 receptor which comprises (a) contacting a cell transfected with and expressing the Y5 receptor with the plurality of



-47-

compounds not known to bind specifically to the Y5 receptor, under conditions permitting activation of the Y5 receptor; (b) determining whether the activity of the Y5 receptor is increased in the presence of the compounds; and if so (c) separately determining whether the activation of the Y5 receptor is increased by each compound included in the plurality of compounds, so as to thereby identify the compound which activates the Y5 receptor.

This invention provides a method of screening a plurality of chemical compounds not known to activate a Y5 receptor to identify a compound which activates the Y5 receptor which comprises (a) preparing a cell extract from cells transfected with and expressing DNA encoding the Y5 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with the plurality of compounds not known to bind specifically to the Y5 receptor, under conditions permitting activation of the Y5 receptor; (b) determining whether the activity of the Y5 receptor is increased in the presence of the compounds; and if so (c) separately determining whether the activation of the Y5 receptor is increased by each compound included in the plurality of compounds, so as to thereby identify the compound which activates the Y5 receptor.

This invention provides a method of screening a plurality of chemical compounds not known to inhibit the activation of a Y5 receptor to identify a compound which inhibits the activation of the Y5 receptor, which comprises (a) contacting a cell transfected with and expressing the Y5 receptor with the plurality of compounds in the presence of a known Y5 receptor agonist, under conditions permitting activation of the Y5 receptor; (b) determining whether the activation of the Y5 receptor is reduced in the presence of the plurality of compounds, relative to

-48-

the activation of the Y5 receptor in the absence of the plurality of compounds; and if so (c) separately determining the inhibition of activation of the Y5 receptor for each compound included in the plurality of compounds, so as to thereby identify the compound which inhibits the activation of the Y5 receptor.

This invention provides a method of screening a plurality of chemical compounds not known to inhibit the activation of a Y5 receptor to identify a compound which inhibits the activation of the Y5 receptor, which comprises (a) preparing a cell extract from cells transfected with and expressing DNA encoding the Y5 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with the plurality of compounds in the presence of a known Y5 receptor agonist, under conditions permitting activation of the Y5 receptor; (b) determining whether the activation of the Y5 receptor is reduced in the presence of the plurality of compounds, relative to the activation of the Y5 receptor in the absence of the plurality of compounds; and if so (c) separately determining the inhibition of activation of the Y5 receptor for each compound included in the plurality of compounds, so as to thereby identify the compound which inhibits the activation of the Y5 receptor.

In separate embodiments of the above-described methods, the Y5 receptor is a human Y5 receptor, a rat Y5 receptor, or a canine Y5 receptor. In an embodiment, the cell is a mammalian cell. In a further embodiment, the cell is non-neuronal in origin. In a further embodiment, the cell is a COS-7 cell, a 293 human embryonic kidney cell, a LM(tk-) cell, or an NIH-3T3 cell.

This invention provides a method of screening drugs to identify drugs which specifically bind to a Y5 receptor on the surface of a cell which comprises contacting a

-49-

cell transfected with and expressing DNA encoding a Y5  
receptor with a plurality of drugs under conditions  
permitting binding of drugs to the Y5 receptor,  
determining those drugs which specifically bind to the  
5 transfected cell, and thereby identifying drugs which  
specifically bind to the Y5 receptor.

This invention provides a method of screening drugs to  
identify drugs which specifically bind to a human Y5  
10 receptor on the surface of a cell which comprises  
contacting a cell transfected with and expressing DNA  
encoding a human Y5 receptor with a plurality of drugs  
under conditions permitting binding of drugs to the human  
Y5 receptor, determining those drugs which specifically  
15 bind to the transfected cell, and thereby identifying  
drugs which specifically bind to the human Y5 receptor.

This invention provides a method of screening drugs to  
identify drugs which act as agonists of a Y5 receptor  
20 which comprises contacting a cell transfected with and  
expressing DNA encoding a Y5 receptor with a plurality of  
drugs under conditions permitting the activation of a  
functional Y5 receptor response, determining those drugs  
which activate such receptor in the cell, and thereby  
25 identify drugs which act as Y5 receptor agonists.

This invention provides a method of screening drugs to  
identify drugs which act as agonists of a human Y5  
receptor which comprises contacting a cell transfected  
30 with and expressing DNA encoding a human Y5 receptor with  
a plurality of drugs under conditions permitting the  
activation of a functional human Y5 receptor response,  
determining those drugs which activate such receptor in  
the cell, and thereby identify drugs which act as human  
35 Y5 receptor agonists.

This invention provides a method of screening drugs to

-50-

identify drugs which act as Y5 receptor antagonists which comprises contacting cells transfected with and expressing DNA encoding a Y5 receptor with a plurality of drugs in the presence of a known Y5 receptor agonist, such as PYY or NPY, under conditions permitting the activation of a functional Y5 receptor response, determining those drugs which inhibit the activation of the receptor in the mammalian cell, and thereby identifying drugs which act as Y5 receptor antagonists.

This invention provides a method of screening drugs to identify drugs which act as human Y5 receptor antagonists which comprises contacting cells transfected with and expressing DNA encoding a human Y5 receptor with a plurality of drugs in the presence of a known human Y5 receptor agonist, such as PYY or NPY, under conditions permitting the activation of a functional human Y5 receptor response, determining those drugs which inhibit the activation of the receptor in the mammalian cell, and thereby identifying drugs which act as human Y5 receptor antagonists. In an embodiment, the cell is non-neuronal in origin. In a further embodiment, the cell is a Cos-7 cell, a 293 human embryonic kidney cell, an LM(tk-) cell or an NIH-3T3 cell.

This invention provides a pharmaceutical composition comprising a drug identified by the above-described method and a pharmaceutically acceptable carrier.

This invention provides a method of detecting expression of Y5 receptor by detecting the presence of mRNA coding for the Y5 receptor which comprises obtaining total mRNA from the cell and contacting the mRNA so obtained with the above-described nucleic acid probe under hybridizing conditions, detecting the presence of mRNA hybridized to the probe, and thereby detecting the expression of the Y5 receptor by the cell.

-51-

This invention provides a method of treating an abnormality in a subject, wherein the abnormality is alleviated by the inhibition of a Y5 receptor which comprises administering to a subject an effective amount of the above-described pharmaceutical composition effective to inhibit the Y5 receptor by the subject.

This invention provides a method of treating an abnormality in a subject wherein the abnormality is alleviated by the activation of a Y5 receptor which comprises administering to a subject an effective amount of the above-described pharmaceutical composition effective to activate the Y5 receptor in the subject.

This invention provides a method of treating an abnormality in a subject, wherein the abnormality is alleviated by the inhibition of a Y5 receptor which comprises administering to a subject an effective amount of Y5 receptor antagonist.

This invention provides a method of treating an abnormality in a subject wherein the abnormality is alleviated by the activation of a Y5 receptor which comprises administering to a subject an effective amount of a Y5 receptor agonist. In a further embodiment, the abnormal condition is anorexia. In a separate embodiment, the abnormal condition is a sexual/reproductive disorder. In another embodiment, the abnormal condition is depression. In another embodiment, the abnormal condition is anxiety.

In an embodiment, the abnormal condition is gastric ulcer. In a further embodiment, the abnormal condition is memory loss. In a further embodiment, the abnormal condition is migraine. In a further embodiment, the abnormal condition is pain. In a further embodiment, the abnormal condition is epileptic seizure. In a further

-52-

embodiment, the abnormal condition is hypertension. In a further embodiment, the abnormal condition is cerebral hemorrhage. In a further embodiment, the abnormal condition is shock. In a further embodiment, the abnormal condition is congestive heart failure. In a further embodiment, the abnormal condition is sleep disturbance. In a further embodiment, the abnormal condition is nasal congestion. In a further embodiment, the abnormal condition is diarrhea.

10

This invention provides a method of treating obesity in a subject which comprises administering to the subject an effective amount of a Y5 receptor antagonist.

15 This invention provides a method of treating anorexia in a subject which comprises administering to the subject an effective amount of a Y5 receptor agonist.

20 This invention provides a method of treating bulimia nervosa in a subject which comprises administering to the subject an effective amount of a Y5 receptor antagonist.

25 This invention provides a method of inducing a subject to eat which comprises administering to the subject an effective amount of a Y5 receptor agonist. In one embodiment, the subject is a vertebrate. In another embodiment, the subject is a human.

30 This invention provides a method of increasing the consumption of a food product by a subject which comprises a composition of the food product and an effective amount of a Y5 receptor agonist. In one embodiment, the subject is a vertebrate. In another embodiment, the subject is a human.

35

This invention provides a method of treating abnormalities which are alleviated by reduction of

-53-

activity of a human Y5 receptor which comprises administering to a subject an amount of the above-described pharmaceutical composition effective to reduce the activity of human Y5 receptor and thereby alleviate abnormalities resulting from overactivity of a human Y5 receptor.

This invention provides a method of treating an abnormal condition related to an excess of Y5 receptor activity which comprises administering to a subject an amount of the pharmaceutical composition effective to block binding of a ligand to the Y5 receptor and thereby alleviate the abnormal condition.

This invention provides a method of detecting the presence of a human Y5 receptor on the surface of a cell which comprises contacting the cell with the antibody capable of binding to the human Y5 receptor under conditions permitting binding of the antibody to the receptor, detecting the presence of the antibody bound to the cell, and thereby detecting the presence of a human Y5 receptor on the surface of the cell.

This invention provides a method of determining the physiological effects of varying levels of activity of a human Y5 receptors which comprises producing a transgenic nonhuman mammal whose levels of human Y5 receptor activity are varied by use of an inducible promoter which regulates human Y5 receptor expression.

This invention provides a method of determining the physiological effects of varying levels of activity of a human Y5 receptors which comprises producing a panel of transgenic nonhuman mammals each expressing a different amount of human Y5 receptor.

This invention provides a method for identifying a

-54-

substance capable of alleviating the abnormalities resulting from overactivity of a human Y5 receptor comprising administering a substance to the above-described transgenic nonhuman mammals, and determining  
5 whether the substance alleviates the physical and behavioral abnormalities displayed by the transgenic nonhuman mammal as a result of overactivity of a human Y5 receptor.

10 This invention provides a method for treating the abnormalities resulting from overactivity of a human Y5 receptor which comprises administering to a subject an amount of the above-described pharmaceutical composition effective to alleviate the abnormalities resulting from  
15 overactivity of a human Y5 receptor.

This invention provides a method for identifying a substance capable of alleviating the abnormalities resulting from underactivity of a human Y5 receptor comprising administering the substance to the above-described transgenic nonhuman mammals and determining  
20 whether the substance alleviates the physical and behavioral abnormalities displayed by the transgenic nonhuman mammal as a result of underactivity of a human Y5 receptor.  
25

This invention provides a method for treating the abnormalities resulting from underactivity of a human Y5 receptor which comprises administering to a subject an amount of the above-described pharmaceutical composition effective to alleviate the abnormalities resulting from  
30 underactivity of a human Y5 receptor.

This invention provides a method for diagnosing a predisposition to a disorder associated with the activity  
35 of a specific human Y5 receptor allele which comprises:  
a. obtaining DNA of subjects suffering from the disorder;



-55-

performing a restriction digest of the DNA with a panel of restriction enzymes; c. electrophoretically separating the resulting DNA fragments on a sizing gel; d. contacting the resulting gel with a nucleic acid probe capable of specifically hybridizing to DNA encoding a human Y5 receptor and labelled with a detectable marker; e. detecting labelled bands which have hybridized to the DNA encoding a human Y5 receptor labelled with a detectable marker to create a unique band pattern specific to the DNA of subjects suffering from the disorder; f. preparing DNA obtained for diagnosis by steps a-e; and g. comparing the unique band pattern specific to the DNA of subjects suffering from the disorder from step e and the DNA obtained for diagnosis from step f to determine whether the patterns are the same or different and to diagnose thereby predisposition to the disorder if the patterns are the same. In an embodiment, a disorder associated with the activity of a specific human Y5 receptor allele is diagnosed.

This invention provides a method of preparing an isolated Y5 receptor which comprises: a. inducing cells to express the Y5 receptor; b. recovering the receptor from the resulting cells; and c. purifying the receptor so recovered.

This invention provides a method of preparing the isolated Y5 receptor which comprises: a. inserting nucleic acid encoding Y5 receptor in a suitable vector which comprises the regulatory elements necessary for expression of the nucleic acid operatively linked to the nucleic acid encoding a Y5 receptor; b. inserting the resulting vector in a suitable host cell so as to obtain a cell which produces the Y5 receptor; c. recovering the receptor produced by the resulting cell; and d. purifying the receptor so recovered.

-56-

This invention will be better understood from the Experimental Details which follow. However, one skilled in the art will readily appreciate that the specific methods and results discussed are merely illustrative of the invention as described more fully in the claims which follow thereafter.

## Experimental Details

### **MATERIALS AND METHODS**

#### 5    cDNA Cloning

Total RNA was prepared by a modification of the guanidine thiocyanate method (Kingston, 1987), from 5 grams of rat hypothalamus (Rockland, Gilbertsville, PA). Poly A<sup>+</sup>RNA was  
10 purified with a FastTrack kit (Invitrogen Corp., San Diego, CA). Double stranded (ds) cDNA was synthesized from 7 µg of poly A<sup>+</sup> RNA according to Gubler and Hoffman (Gubler and Hoffman, 1983), except that ligase was omitted in the second strand cDNA synthesis. The  
15 resulting DS cDNA was ligated to BstxI/EcoRI adaptors (Invitrogen Corp.), the excess of adaptors was removed by chromatography on Sephacryl 500 HR (Pharmacia-LKB) and the ds-cDNA size selected on a Gen-Pak Fax HPLC column (Millipore Corp., Milford, MA). High molecular weight  
20 fractions were ligated in pEXJ.BS (A cDNA cloning expression vector derived from pcEXV-3; Okayama and Berg, 1983; Miller and Germain, 1986) cut by BstxI as described by Aruffo and Seed (Aruffo and Seed, 1987). The ligated DNA was electroporated in E.Coli MC 1061 F<sup>+</sup> (Gene Pulser, Biorad). A total of  $3.4 \times 10^6$  independent clones with an  
25 insert mean size of 2.7 kb could be generated. The library was plated on Petri dishes (Ampicillin selection) in pools of  $6.9$  to  $8.2 \times 10^3$  independent clones. After 18 hours amplification, the bacteria from each pool were  
30 scraped, resuspended in 4 mL of LB media and 1.5 mL processed for plasmid purification with a QIAprep-8 plasmid kit (Qiagen Inc, Chatsworth, CA). 1 ml aliquots of each bacterial pool were stored at -85°C in 20% glycerol.

35

Isolation of a cDNA clone encoding an atypical rat hypothalamic NPY5 receptor

-58-

DNA from pools of ~ 7500 independent clones was transfected into COS-7 cells by a modification of the DEAE-dextran procedure (Warden and Thorne, 1968). COS-7 cells were grown in Dulbecco's modified Eagle medium (DMEM) supplemented with 10% fetal calf serum, 100 U/ml of penicillin, 100  $\mu$ g/ml of streptomycin, 2mM L-glutamine (DMEM-C) at 37°C in 5% CO<sub>2</sub>. The cells were seeded one day before transfection at a density of 30,000 cells/cm<sup>2</sup> on Lab-Tek chamber slides (1 chamber, Permanox slide from Nunc Inc., Naperville, IL). On the next day, cells were washed twice with PBS, 735  $\mu$ l of transfection cocktail was added containing 1/10 of the DNA from each pool and DEAE-dextran (500  $\mu$ g/ml) in Opti-MEM I serum free media (Gibco®BRL LifeTechnologies Inc. Grand Island, NY). After a 30 min. incubation at 37°C, 3 ml of chloroquine (80  $\mu$ M in DMEM-C) was added and the cells incubated a further 2.5 hours at 37°C. The media was aspirated from each chamber and 2 ml of 10% DMSO in DMEM-C added. After 2.5 min. incubation at room temperature, the media was aspirated, each chamber washed once with 2 ml PBS, the cells incubated 48 hours in DMEM-C and the binding assay was performed on the slides. After one wash with PBS, positive pools were identified by incubating the cells with 1 nM (3x10<sup>6</sup> cpm per slide) of porcine [<sup>125</sup>I]-PYY (NEN; SA=2200Ci/mmmole) in 20 mM Hepes-NaOH pH 7.4, CaCl<sub>2</sub> 1.26 mM, MgSO<sub>4</sub> 0.81 mM, KH<sub>2</sub>PO<sub>4</sub> 0.44 mM, KCL 5.4, NaCl 10mM, .1% BSA, 0.1% bacitracin for 1 hour at room temperature. After six washes (three seconds each) in binding buffer without ligand, the monolayers were fixed in 2.5% glutaraldehyde in PBS for five minutes, washed twice for two minutes in PBS, dehydrated in ethanol baths for two minutes each (70, 80, 95, 100%) and air dried. The slides were then dipped in 100% photoemulsion (Kodak type NTB2) at 42°C and exposed in the dark for 48 hours at 4°C in light proof boxes containing drierite. Slides were developed for three minutes in Kodak D19 developer (32 g/l of water), rinsed in water, fixed in Kodak fixer for

-59-

5 minutes, rinsed in water, air dried and mounted with Aqua-Mount (Lerner Laboratories, Pittsburgh, PA). Slides were screened at 25x total magnification. A single clone, CG-18, was isolated by SIB selection as described (Mc Cormick, 1987). DS-DNA was sequenced with a Sequenase kit (US Biochemical, Cleveland, OH) according to the manufacturer. Nucleotide and peptide sequence analysis were performed with GCG programs (Genetics Computer group, Madison, WI).

10

#### Isolation of the human Y5 homolog

Using rat oligonucleotide primers in TM 3 (sense primer; position 484-509 in fig. 1A) and in TM 6 (antisense primer; position 1219-1243 in fig. 3A), applicants screened a human hippocampal cDNA library using the polymerase chain reaction. 1  $\mu$ l ( $4 \times 10^6$  bacteria) of each of 450 amplified pools containing each ~5000 independent clones and representing a total of  $2.2 \times 10^6$  was subjected directly to 40 cycles of PCR and the resulting products analyzed by agarose gel electrophoresis. One of three positive pools was analyzed further and by sib selection a single cDNA clone was isolated and characterized. This cDNA turned out to be full length and in the correct orientation for expression. DS-DNA was sequenced with a sequenase kit (US Biochemical, Cleveland, OH) according to the manufacturer.

30

#### Isolation of the canine Y5 homolog

An alignment of the coding nucleotide sequences of the rat and human Y5 receptors was used to synthesize a pair of PCR primers. A region upstream of TM III which is 100% conserved between rat and human was chosen to synthesize the forward primer CH 156:

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-60-

5'-TGGATCAGTGGATGTTTGGCAAAG-3' (Seq. I.D. No. 7).

5 A region at the carboxy end of the 5-6 loop, immediately upstream of TM6, which is also 100% conserved between rat and human sequences was chosen to synthesize the reverse primer CH153:

5'-GTCTGTAGAAAACACTTCGAGATCTCTT-3' (Seq. I.D. No. 8).

10 The primers CH156-CH153 were used to amplify 10 ng of poly (A+) RNA from rat brain that was reverse transcribed using the SSII reverse transcriptase (GibcoBRL, Gaithersburg, MD). PCR was performed on single-stranded  
15 cDNA with Taq Polymerase (Perkin Elmer-Roche Molecular Systems, Branchburg, NJ) under the following conditions: 94°C for 1 min, 60°C for 1 min and 72°C for 1 min for 40 cycles. The resulting 798 bp PCR DNA fragment was subcloned in pCR Script (Stratagene, La Jolla, CA) and sequenced using a sequenase kit (USB, Cleveland, OH) and  
20 is designated Y5-bd-5.

### 3' and 5' RACE

The missing 3' and 5' ends of the beagle dog Y5 receptor sequences were isolated by 3' and 5' RACE using a  
25 Marathon cDNA amplification kit (Clontech, Palo Alto, CA). From the sequence of the beagle dog PCR DNA fragment described above, the following PCR primers were synthesized:

30 (3' RACE)

CH 204:

5'-CTTCCAGTGTTCACAGTCTGGTGG-3' (Seq. I.D. No. 9);

CH 218 (nested primer):

35 5'-CTGAGCAGCAGGTATTTATGTGTTG-3' (Seq. I.D. No. 10);

(5' RACE)

-61-

CH 219:

5'-CTGGATGAAGAATGCTGACTTCTTACAG-3' (Seq. I.D. No. 11);

5 CH 245 (nested primer):

5'-TTCTTGAGTGGTTCTCTTGAGGAGG-3' (Seq. I.D. No. 12).

The 3' and 5' RACE reactions were carried out on beagle dog thalamic cDNA according to the kit specifications, with the primers described above. The resulting PCR DNA products (smear of 0.7 to 10 kb) were purified from an agarose gel and reamplified using the nested primers described above. The resulting DNA bands were again purified from an agarose gel and subcloned in pCR Script (Stratagene, La Jolla, CA).

The nucleotide sequence corresponding to the 3' end of the cDNA was determined and the plasmid designated Y5-bd-8. The nucleotide sequence corresponding to the 5' end will be determined in the near future. Those nucleotide sequences will then be used to synthesize exact primers against the initiation and stop codon regions and those exact primers will then be used to amplify canine thalamic cDNA to generate a PCR product corresponding to the full length coding region of the canine Y5 receptor, using the Expand High Fidelity polymerase (Boehringer Mannheim Corporation, Indianapolis, IN). The resulting PCR DNA product will be subcloned in the expression vector pEXJ and the entire coding region of the canine Y5 nucleotide sequence will be determined using a Sequenase Kit (USB, Cleveland, OH).

#### Northern Blots

Human brain multiple tissue northern blots (MTN blots II and III, Clontech, Palo Alto, CA) carrying mRNA purified from various human brain areas was hybridized at high stringency according to the manufacturer specifications.

-62-

The probe was a 0.8 kb DNA PCR fragment corresponding to the TM III - carboxy end of the 5-6 loop in the coding region of the human Y5 receptor subtype.

- 5 A rat multiple tissue northern blot (rat MTN blot, Clontech, Palo Alto, CA) carrying mRNA purified from various rat tissues was hybridized at high stringency according to the manufacturer specifications. The probe was a 0.8 kb DNA PCR fragment corresponding to the TM III  
10 - carboxy end of the 5-6 loop in the coding region of the rat Y5 receptor subtype.

#### Southern Blot

- 15 Southern blots (Geno-Blot, clontech, Palo Alto, CA) containing human or rat genomic DNA cut with five different enzymes (8  $\mu$ g DNA per lane) was hybridized at high stringency according to the manufacturer specifications. The probe was a .8 kb DNA PCR fragment  
20 in the coding region of the human and rat Y5 receptor subtypes.

#### Production of Recombinant Baculovirus

- 25 A Bam HI site directly 5' to the starting methionine of human Y5 was genetically engineered by replacing the beginning  $\sim$ 100 base pairs of hY5 (i.e. from the starting methionine to an internal EcoRI site) with two overlapping synthetically-derived oligonucleotides ( $\sim$ 100 bases each), containing a 5' Bam HI site and a 3' EcoRI  
30 site. This permitted the isolation of an  $\sim$ 1.5 kb Bam HI/Hind III fragment containing the coding region of hY5. This fragment was subcloned into pBlueBacIII<sup>TM</sup> into the Bam HI/Hind III sites found in the polylinker (construct called pBB/hY5). To generate baculovirus, 0.5  $\mu$ g of  
35 viral DNA (BaculoGold<sup>TM</sup>) and 3  $\mu$ g of pBB/hY5 were co-transfected into  $2 \times 10^6$  Spodoptera frugiperda insect Sf9 cells by calcium phosphate co-precipitation method, as



-63-

outlined in by Pharmingen (in "Baculovirus Expression Vector System: Procedures and Methods Manual"). The cells were incubated for 5 days at 27°C. The supernatant of the co-transfection plate was collected by centrifugation and the recombinant virus (hY5BB3) was plaque purified. The procedure to infect cells with virus, to prepare stocks of virus and to titer the virus stocks were as described in Pharmingen's manual.

#### 10 Cell Culture

COS-7 cells were grown on 150 mm plates in D-MEM with supplements (Dulbecco's Modified Eagle Medium with 10% bovine calf serum, 4 mM glutamine, 100 units/ml penicillin/100 µg/ml streptomycin) at 37°C, 5% CO<sub>2</sub>. Stock plates of COS-7 cells were trypsinized and split 1:6 every 3-4 days. Human embryonic kidney 293 cells were grown on 150 mm plates in D-MEM with supplements (minimal essential medium) with Hanks' salts and supplements (Dulbecco's Modified Eagle Medium with 10% bovine calf serum, 4 mM glutamine, 100 units/ml penicillin/100 µg/ml streptomycin) at 37 °C, 5% CO<sub>2</sub>. Stock plates of 293 cells were trypsinized and split 1:6 every 3-4 days. Mouse fibroblast LM(tk-) cells were grown on 150 mm plates in D-MEM with supplements (Dulbecco's Modified Eagle Medium with 10% bovine calf serum, 4 mM glutamine, 100 units/mL penicillin/100 µg/mL streptomycin) at 37 °C, 5% CO<sub>2</sub>. Stock plates of LM(tk-) cells were trypsinized and split 1:10 every 3-4 days.

LM(tk-) cells stably transfected with the human Y5 receptor were routinely converted from an adherent monolayer to a viable suspension. Adherent cells were harvested with trypsin at the point of confluence, resuspended in a minimal volume of complete DMEM for a cell count, and further diluted to a concentration of 10<sup>6</sup> cells/ml in suspension media (10% bovine calf serum, 10% 10X Medium 199 (Gibco), 9 mM NaHCO<sub>3</sub>, 25 mM glucose, 2 mM

-64-

L-glutamine, 100 units/ml penicillin/100  $\mu$ g/ml streptomycin, and 0.05% methyl cellulose). The cell suspension was maintained in a shaking incubator at 37 °C, 5% CO<sub>2</sub> for 24 hours. Membranes harvested from cells grown in this manner may be stored as large, uniform batches in liquid nitrogen. Alternatively, cells may be returned to adherent cell culture in complete DMEM by distribution into 96-well microtiter plates coated with poly-D-lysine (0.01 mg/ml) followed by incubation at 37 °C, 5% CO<sub>2</sub> for 24 hours. Cells prepared in this manner yielded a robust and reliable NPY-dependent response in cAMP radio-immunoassays as further described hereinbelow.

Mouse embryonic fibroblast NIH-3T3 cells were grown on 150 mm plates in Dulbecco's Modified Eagle Medium (DMEM) with supplements (10% bovine calf serum, 4 mM glutamine, 100 units/ml penicillin/100  $\mu$ g/ml streptomycin) at 37 °C, 5% CO<sub>2</sub>. Stock plates of NIH-3T3 cells were trypsinized and split 1:15 every 3-4 days.

Sf9 and Sf21 cells were grown in monolayers on 150 mm tissue culture dishes in TMN-FH media supplemented with 10% fetal calf serum, at 27°C, no CO<sub>2</sub>. High Five insect cells were grown on 150 mm tissue culture dishes in Ex-Cell 400™ medium supplemented with L-Glutamine, also at 27°C, no CO<sub>2</sub>.

#### Transient Transfection

All receptor subtypes studied (human and rat Y1, human and rat Y2, human and rat Y4, human and rat Y5) were transiently transfected into COS-7 cells by the DEAE-dextran method, using 1  $\mu$ g of DNA /10<sup>6</sup> cells (Cullen, 1987). The human Y1 receptor was prepared using known methods (Larhammar, et al., 1992).

#### Stable Transfection

-65-

Human Y1, human Y2, and rat Y5 receptors were co-transfected with a G-418 resistant gene into the human embryonic kidney 293 cell line by a calcium phosphate transfection method (Cullen, 1987). Stably transfected cells were selected with G-418. Human Y4 and human Y5 receptors were similarly transfected into mouse fibroblast LM(tk-) cells and NIH-3T3 cells.

#### Expression of other G-protein coupled receptors

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**$\alpha_1$  Human Adrenergic Receptors:** To determine the binding of compounds to human  $\alpha_1$  receptors, LM(tk-) cell lines stably transfected with the genes encoding the  $\alpha_{1a}$ ,  $\alpha_{1b}$ , and  $\alpha_{1d}$  receptors were used. The nomenclature describing the  $\alpha_1$  receptors was changed recently, such that the receptor formerly designated  $\alpha_{1a}$  is now designated  $\alpha_{1d}$ , and the receptor formerly designated  $\alpha_{1c}$  is now designated  $\alpha_{1a}$  (ref). The cell lines expressing these receptors were deposited with the ATCC before the nomenclature change and reflect the subtype designations formerly assigned to these receptors. Thus, the cell line expressing the receptor described herein as the  $\alpha_{1a}$  receptor was deposited with the ATCC on September 25, 1992, under ATCC Accession No. CRL 11140 with the designation L- $\alpha_{1c}$ . The cell line expressing receptor described herein as the  $\alpha_{1d}$  receptor was deposited with the ATCC on September 25, 1992, under ATCC Accession No. CRL 11138 with the designation L- $\alpha_{1A}$ . The cell line expressing the  $\alpha_{1b}$  receptor is designated L- $\alpha_{1B}$ , and was deposited on September 25, 1992, under ATCC Accession No. CRL 11139.

**$\alpha_2$  Human Adrenergic Receptors:** To determine the binding of compounds to human  $\alpha_2$  receptors, LM(tk-) cell lines stably transfected with the genes encoding the  $\alpha_{2A}$ ,  $\alpha_{2B}$ , and  $\alpha_{2C}$  receptors were used. The cell line expressing the  $\alpha_{2A}$  receptor is designated L- $\alpha_{2A}$ , and was

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-66-

deposited on November 6, 1992, under ATCC Accession No. CRL 11180. The cell line expressing the  $\alpha_{2B}$  receptor is designated L-NGC- $\alpha_{2B}$ , and was deposited on October 25, 1989, under ATCC Accession No. CRL 10275. The cell  
5 line expressing the  $\alpha_{2C}$  receptor is designated L- $\alpha_{2C}$ , and was deposited on November 6, 1992, under ATCC Accession No. CRL-11181. Cell lysates were prepared as described below (see Radioligand Binding to Membrane  
10 Suspensions), and suspended in 25mM glycylglycine buffer (pH 7.6 at room temperature). Equilibrium competition binding assay were performed using [ $^3$ H]rauwolscine (0.5nM), and nonspecific binding was determined by incubation with 10 $\mu$ M phentolamine. The  
15 bound radioligand was separated by filtration through GF/B filters using a cell harvester.

**Human Histamine  $H_1$  Receptor:** The coding sequence of the human histamine  $H_1$  receptor, homologous to the bovine  $H_1$  receptor, was obtained from a human  
20 hippocampal cDNA library, and was cloned into the eukaryotic expression vector pCEXV-3. The plasmid DNA for the  $H_1$  receptor is designated pcEXV-H1, and was deposited on November 6, 1992, under ATCC Accession No. 75346. This construct was transfected into COS-7 cells  
25 by the DEAE-dextran method. Cells were harvested after 72 hours and lysed by sonication in 5mM Tris-HCl, 5mM EDTA, pH 7.5. The cell lysates were centrifuged at 1000 rpm for 5 min at 4°C, and the supernatant was centrifuged at 30,000 x g for 20 min. at 4°C. The  
30 pellet was suspended in 37.8 mM NaHPO<sub>4</sub>, 12.2 mM KH<sub>2</sub>PO<sub>4</sub>, pH 7.5. The binding of the histamine  $H_1$  antagonist [ $^3$ H]mepyramine (1nM, specific activity: 24.8 Ci/mM) was done in a final volume of 0.25 mL and incubated at room temperature for 60 min. Nonspecific binding was  
35 determined in the presence of 10  $\mu$ M mepyramine. The bound radioligand was separated by filtration through GF/B filters using a cell harvester.

-67-

**Human Histamine H<sub>2</sub> Receptor:** The coding sequence of the human H<sub>2</sub> receptor was obtained from a human placenta genomic library, and cloned into the cloning site of PCEXV-3 eukaryotic expression vector. The plasmid DNA for the H<sub>2</sub> receptor is designated pcEXV-H2, and was deposited on November 6, 1992 under ATCC Accession No. 75345. This construct was transfected into COS-7 cells by the DEAE-dextran method. Cells were harvested after 72 hours and lysed by sonication in 5mM Tris-HCl, 5mM EDTA, pH 7.5. The cell lysates were centrifuged at 1000 rpm for 5 min at 4°C, and the supernatant was centrifuged at 30,000 x g for 20 min at 4 °C. The pellet was suspended in 37.8 mM NaHPO<sub>4</sub>, 12.2 mM K<sub>2</sub>PO<sub>4</sub>, pH 7.5. The binding of the histamine H<sub>2</sub> antagonist [<sup>3</sup>H]tiotidine (5nM, specific activity: 70 Ci/mM) was done in a final volume of 0.25 ml and incubated at room temperature for 60 min. Nonspecific binding was determined in the presence of 10 μM histamine. The bound radioligand was separated by filtration through GF/B filters using a cell harvester.

**Human Serotonin Receptors:**

5HT<sub>1Dα</sub>, 5HT<sub>1Dβ</sub>, 5HT<sub>1E</sub>, 5HT<sub>1F</sub> Receptors: LM(tk-) clonal cell lines stably transfected with the genes encoding each of these 5HT receptor subtypes were prepared as described above. The cell line for the 5HT<sub>1Dα</sub> receptor, designated as Ltk-8-30-84, was deposited on April 17, 1990, and accorded ATCC Accession No. CRL 10421. The cell for the 5HT<sub>1Dβ</sub> receptor, designated as Ltk-11, was deposited on April 17, 1990, and accorded ATCC Accession No. CRL 10422. The cell line for the 5HT<sub>1E</sub> receptor, designated 5 HT<sub>1E</sub>-7, was deposited on November 6, 1991, and accorded ATCC Accession No. CRL 10913. The cell line for the 5HT<sub>1F</sub> receptor, designated L-5-HT<sub>1F</sub>, was deposited on December 27, 1991, and accorded ATCC Accession No. ATCC 10957. Membrane preparations comprising these receptors were prepared

-68-

as described below, and suspended in 50mM Tris-HCl buffer (pH 7.4 at 37°C) containing 10 mM MgCl<sub>2</sub>, 0.2 mM EDTA, 10μM pargyline, and 0.1% ascorbate. The binding of compounds was determined in competition binding assays by incubation for 30 minutes at 37°C in the presence of 5nM [<sup>3</sup>H]serotonin. Nonspecific binding was determined in the presence of 10μM serotonin. The bound radioligand was separated by filtration through GF/B filters using a cell harvester.

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**Human 5HT<sub>2</sub> Receptor:** The coding sequence of the human 5HT<sub>2</sub> receptor was obtained from a human brain cortex cDNA library, and cloned into the cloning site of pCEXV-3 eukaryotic expression vector. This construct was transfected into COS-7 cells by the DEAE-dextran method. Cells were harvested after 72 hours and lysed by sonication in 5mM Tris-HCl, 5mM EDTA, pH 7.5. This cell line was deposited with the ATCC on October 31, 1989, designated as L-NGC-5HT<sub>2</sub>, and was accorded ATCC Accession No. CRL 10287. The cell lysates were centrifuged at 1000 rpm for 5 minutes at 4°C, and the supernatant was centrifuged at 30,000 x g for 20 minutes at 4°C. The pellet was suspended in 50mM Tris-HCl buffer (pH 7.7 at room temperature) containing 10 mM MgSO<sub>4</sub>, 0.5mM EDTA, and 0.1% ascorbate. The potency of alpha-1 antagonists at 5HT<sub>2</sub> receptors was determined in equilibrium competition binding assays using [3H]ketanserin (1nM). Nonspecific binding was defined by the addition of 10μM mianserin. The bound radioligand was separated by filtration through GF/B filters using a cell harvester.

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**Human 5-HT<sub>1</sub> Receptor:** A LM(tk-) clonal cell line stably transfected with the gene encoding the 5HT<sub>1</sub> receptor subtype was prepared as described above. The cell line for the 5HT<sub>1</sub> receptor, designated as L-5HT<sub>1B</sub>, was deposited on October 20, 1992, and accorded ATCC

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Accession No. CRL 11166.

**Human Dopamine D<sub>3</sub> Receptor:** The binding of compounds to the human D<sub>3</sub> receptor was determined using membrane preparations from COS-7 cells transfected with the gene encoding the human D<sub>3</sub> receptor. The human dopamine D<sub>3</sub> receptor was prepared according to known methods (Sokoloff, P. et al. Nature, 347, 146, 1990, deposited with the EMBL Genbank as X53944). Cells were harvested after 72 hours and lysed by sonication in 5mM Tris-HCl, 5mM EDTA, pH 7.5. The cell lysates were centrifuged at 1000 rpm for 5 minutes at 4°C, and the supernatant was centrifuged at 30,000 x g for 20 minutes at 4°C. The pellet was suspended in 50 mM Tris-HCl (pH 7.4) containing 1mM EDTA, 5mM KCl, 1.5mM CaCl<sub>2</sub>, 4mM MgCl<sub>2</sub>, and 0.1% ascorbic acid. The cell lysates were incubated with [<sup>3</sup>H]spiperone (2nM), using 10μM (+)Butaclamol to determine nonspecific binding.

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#### Membrane Harvest

Membranes were harvested from COS-7 cells 48 hours after transient transfection. Adherent cells were washed twice in ice-cold phosphate buffered saline (138 mM NaCl, 8.1 mM Na<sub>2</sub>HPO<sub>4</sub>, 2.5 mM KCl, 1.2 mM KH<sub>2</sub>PO<sub>4</sub>, 0.9 mM CaCl<sub>2</sub>, 0.5 mM MgCl<sub>2</sub>, pH 7.4) and lysed by sonication in ice-cold sonication buffer (20 mM Tris-HCl, 5 mM EDTA, pH 7.7). Large particles and debris were cleared by low speed centrifugation (200 x g, 5 min, 4 °C). Membranes were collected from the supernatant fraction by centrifugation (32,000 x g, 18 min, 4 °C), washed with ice-cold hypotonic buffer, and collected again by centrifugation (32,000 x g, 18 min, 4 °C). The final membrane pellet was resuspended by sonication into a small volume of ice-cold binding buffer (~1 ml for every 5 plates: 10 mM NaCl, 20 mM HEPES, 0.22 mM KH<sub>2</sub>PO<sub>4</sub>, 1.26 mM CaCl<sub>2</sub>, 0.81 mM MgSO<sub>4</sub>, pH 7.4). Protein

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-70-

concentration was measured by the Bradford method (Bradford, 1976) using Bio-Rad Reagent, with bovine serum albumin as a standard. Membranes were held on ice for up to one hour and used fresh, or flash-frozen and stored in liquid nitrogen.

Membranes were prepared similarly from 293, LM(tk-), and NIH-3T3 cells. To prepare membranes from baculovirus infected cells,  $2 \times 10^7$  Sf21 cells were grown in 150mm tissue culture dishes and infected with a high-titer stock of hY5BB3. Cells were incubated for 2-4 days at 27°C, no CO<sub>2</sub> before harvesting and membrane preparation as described above.

Membranes were prepared similarly from dissected rat hypothalamus. Frozen hypothalami were homogenized for 20 seconds in ice-cold sonication buffer with the narrow probe of a Virtishear homogenizer at 1000 rpm (Virtis, Gardiner, NY). Large particles and debris were cleared by centrifugation (200 x g, 5 min, 4 °C) and the supernatant fraction was reserved on ice. Membranes were further extracted from the pellet by repeating the homogenization and centrifugation procedure two more times. The supernatant fractions were pooled and subjected to high speed centrifugation (100,000 x g, 20 min. 4 °C). The final membrane pellet was resuspended by gentle homogenization into a small volume of ice-cold binding buffer (1 mL/ gram wet weight tissue) and held on ice for up to one hour, or flash-frozen and stored in liquid nitrogen.

#### Radioligand Binding to Membrane Suspensions

Membrane suspensions were diluted in binding buffer supplemented with 0.1% bovine serum albumin to yield an optimal membrane protein concentration so that <sup>125</sup>I-PYY (or alternative radioligand such as <sup>125</sup>I-NPY, <sup>125</sup>I-PYY<sub>3-36</sub>, or <sup>125</sup>I-[Leu<sup>31</sup>Pro<sup>34</sup>]PYY) bound by membranes in the assay



-71-

was less than 10% of  $^{125}\text{I}$ -PYY (or alternative radioligand) delivered to the sample (100,000 dpm/sample = 0.08 nM for competition binding assays).  $^{125}\text{I}$ -PYY (or alternative radioligand) and peptide competitors were also diluted to desired concentrations in supplemented binding buffer. Individual samples were then prepared in 96-well polypropylene microtiter plates by mixing  $^{125}\text{I}$ -PYY (25  $\mu\text{L}$ ) (or alternative radioligand), competing peptides or supplemented binding buffer (25  $\mu\text{L}$ ), and finally, membrane suspensions (200  $\mu\text{l}$ ). Samples were incubated in a 30 °C water bath with constant shaking for 120 min. Incubations were terminated by filtration over Whatman GF/C filters (pre-coated with 1% polyethyleneimine and air-dried before use), followed by washing with 5 mL of ice-cold binding buffer. Filter-trapped membranes were impregnated with MultiLex solid scintillant (Wallac, Turku, Finland) and counted for  $^{125}\text{I}$  in a Wallac Beta-Plate Reader. Non-specific binding was defined by 300 nM human NPY for all receptors except the Y4 subtypes; 100 nM human PP was used for the human Y4 and 100 nM rat PP for the rat Y4. Specific binding in time course and competition studies was typically 80%; most non-specific binding was associated with the filter. Binding data were analyzed using nonlinear regression and statistical techniques available in the GraphPAD Prism package (San Diego, CA).

#### Functional Assay: Radioimmunoassay of cAMP

Stably transfected cells were seeded into 96-well microtiter plates and cultured until confluent. To reduce the potential for receptor desensitization, the serum component of the media was reduced to 1.5% for 4 to 16 hours before the assay. Cells were washed in Hank's buffered saline, or HBS (150 mM NaCl, 20 mM HEPES, 1 mM  $\text{CaCl}_2$ , 5 mM KCl, 1 mM  $\text{MgCl}_2$ , and 10 mM glucose) supplemented with 0.1% bovine serum albumin

-72-

plus 5 mM theophylline and pre-equilibrated in the same solution for 20 min at 37 °C in 5% CO<sub>2</sub>. Cells were then incubated 5 min with 10 μM forskolin and various concentrations of receptor-selective ligands. The assay was terminated by the removal of HBS and acidification of the cells with 100 mM HCl. Intracellular cAMP was extracted and quantified with a modified version of a magnetic bead-based radioimmunoassay (Advanced Magnetics, Cambridge, MA). The final antigen/antibody complex was separated from free <sup>125</sup>I-cAMP by vacuum filtration through a PVDF filter in a microtiter plate (Millipore, Bedford, MA). Filters were punched and counted for <sup>125</sup>I in a Packard gamma counter. Binding data were analyzed using nonlinear regression and statistical techniques available in the GraphPAD Prism package (San Diego, CA).

#### Functional Assay: Intracellular calcium mobilization

The intracellular free calcium concentration was measured by microspectrofluorometry using the fluorescent indicator dye Fura-2/AM (ref). Stably transfected cells were seeded onto a 35 mm culture dish containing a glass coverslip insert. Cells were washed with HBS and loaded with 100 μl of Fura-2/AM (10 μM) for 20 to 40 min. After washing with HBS to remove the Fura-2/AM solution, cells were equilibrated in HBS for 10 to 20 min. Cells were then visualized under the 40X objective of a Leitz Fluovert FS microscope and fluorescence emission was determined at 510 nm with excitation wave lengths alternating between 340 nm and 380 nm. Raw fluorescence data were converted to calcium concentrations using standard calcium concentration curves and software analysis techniques.

#### Tissue preparation for neuroanatomical studies

Male Sprague-Dawley rats (Charles Rivers) were

-73-

decapitated and the brains rapidly removed and frozen in isopentane. Coronal sections were cut at 11  $\mu$ m on a cryostat and thaw-mounted onto poly-L-lysine coated slides and stored at -80° C until use. Prior to hybridization, tissues were fixed in 4% paraformaldehyde, treated with 5 mM dithiothreitol, acetylated in 0.1 M triethanolamine containing 0.25% acetic anhydride, delipidated with chloroform, and dehydrated in graded ethanols.

### Probes

The oligonucleotide probes employed to characterize the distribution of the rat NPY Y5 mRNA were complementary to nucleotides 1121 to 1165 in the 5,6-loop of the rat Y5 mRNA (fig. 3A) 45mer antisense and sense oligonucleotide probes were synthesized on a Millipore Expedite 8909 Nucleic Acid Synthesis System. The probes were then lyophilized, reconstituted in sterile water, and purified on a 12% polyacrylamide denaturing gel. The purified probes were again reconstituted to a concentration of 100 ng/ $\mu$ l, and stored at -20°C.

### In Situ Hybridization

Probes were 3'-end labeled with <sup>35</sup>S-dATP (1200 Ci/mmol, New England Nuclear, Boston, MA) to a specific activity of 10<sup>9</sup> dpm/ $\mu$ g using terminal deoxynucleotidyl transferase (Pharmacia). The radiolabeled probes were purified on Biospin 6 chromatography columns (Bio-Rad; Richmond, CA), and diluted in hybridization buffer to a concentration of 1.5 x 10<sup>4</sup> cpm/ $\mu$ l. The hybridization buffer consisted of 50% formamide, 4X sodium citrate buffer (1X SSC = 0.15 M NaCl and 0.015 M sodium citrate), 1X Denhardt's solution (0.2% polyvinylpyrrolidone, 0.2% Ficoll, 0.2% bovine serum albumin), 50 mM dithiothreitol, 0.5 mg/ml salmon sperm DNA, 0.5 mg/ml yeast tRNA, and 10% dextran sulfate. One hundred  $\mu$ l of the diluted radiolabeled probe was

-74-

applied to each section, which was then covered with a Parafilm coverslip. Hybridization was carried out overnight in humid chambers at 40 to 55°C. The following day the sections were washed in two changes of 2X SSC for one hour at room temperature, in 2X SSC for 30 min at 50-60°C, and finally in 0.1X SSC for 30 min at room temperature. Tissues were dehydrated in graded ethanols and apposed to Kodak XAR-5 film for 3 days to 3 weeks at -20°C, then dipped in Kodak NTB3 autoradiography emulsion diluted 1:1 with 0.2% glycerol water. After exposure at 4°C for 2 to 8 weeks, the slides were developed in Kodak D-19 developer, fixed, and counterstained with cresyl violet.

#### 15     Hybridization controls

Controls for probe/hybridization specificity included hybridization with the radiolabeled sense probe, and the use of transfected cell lines. Briefly, COS-7 cells were transfected (see above) with receptor cDNAs for the rat Y1, Y2 (disclosed in US patent application Serial No. 08/192,288, filed on February 3, 1994), Y4 (disclosed in US patent application Serial No. 08/176,412, filed on December 28 1993), or Y5. As described above, the transfected cells were treated and hybridized with the radiolabeled Y5 antisense and sense oligonucleotide probes, washed, and apposed to film for 1-7 days.

#### Analysis of hybridization signals

30     Sections through the rat brain were analyzed for hybridization signals in the following manner. "Hybridization signal" as used in the present context indicates the relative number of silver grains observed over neurons in a selected area of the rat brain. Two independent observers rated the intensity of the hybridization signal in a given brain area as nonexistent, low, moderate, or high. These were then

-75-

converted to a subjective numerical scale as 0, +1, +2, or +3 (see Table 10), and mapped on to schematic diagrams of coronal sections through the rat brain (see Fig. 11).

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#### Chemical synthetic methods

##### **Compound 28**

##### **2-(Naphthalen-1-ylamino)-3-phenylpropionitrile**

10 To a solution of 1-naphthalenemethylamine (2.9 g, 20 mmol) and benzylaldehyde (2.0 g, 17 mmol) in 30 ml of  $\text{CHCl}_3$  and 10 ml of MeOH was added TMSCN (6.6 ml, 51 mmol) and the resulting solution was stirred for 12 h at 25 °C. The reaction mixture was concentrated in vacuo, yielding an oil which was subjected to column chromatography (EtOAc, neat) to provide 3.5 g (74%) of the desired product as a colorless oil. Product was identified by NMR.

##### **2-(Naphthalen-1-yl)-3-phenylpropane-1,2-diamine**

20 To a solution of the nitrile (0.5 g, 1.8 mmol) in THF was added 6.9 ml of 1N  $\text{LiAlH}_4$  in THF dropwise and the resulting solution was stirred for 2 h. The reaction was quenched by adding a few pieces of ice into the solution. The reaction mixture was diluted with EtOAc and filtered through pad of Celite. Organic filtrate was concentrated in vacuo to provide a oily residue which was subjected to column chromatography (EtOAc, neat) to provide 0.28 g (57%) of the desired product as a colorless oil. The product was identified by NMR.

#### In vivo Studies in rats

##### Food intake in satiated rats

35 For these determinations food intake maybe measured in normal satiated rats after intracerebroventricular application (i.c.v.) of NPY in the presence or absence

-76-

- of the test compound. Male Sprague Dawley rats (Ciba-Geigy AG, Sisseln, Switzerland) weighing between 180g and 220 g are used for all experiments. The rats are individually housed in stainless steel cages and maintained on an 11:13 h light-dark cycle (lights off at 18:00 h) at a controlled temperature of 21-23 °C at all times. Water and food (NAFAG lab chow pellets, NAFAG, Gossau, Switzerland) are available ad libidum.
- 10 Rats under pentobarbital anesthesia are stereotaxically implanted with a stainless steel guide cannula targeted at the right lateral ventricle. Stereotaxic coordinates, with the incisor bar set -2.0mm below interaural line, are: -0.8mm anterior and +1.3mm lateral to bregma. The guide cannula is placed on the dura. Injection cannulas extend the guide cannulas - 3.8mm ventrally to the skull surface. Animals are allowed at least 4 days of recovery postoperatively before being used in the experiments. Cannula placement is checked postoperatively by testing all rats for their drinking response to a 50 ng intracerebroventricular (i.c.v.) injection of angiotensin II. Only rats which drink at least 2.5 ml of water within 30 min. after angiotensin II injection are used in the feeding studies.
- All injections are made in the morning 2 hours after light onset. Peptides are injected in artificial cerebrospinal fluid (ACSF) in a volume of 5µl. ACSF contains: NaCl 124mM, KCl 3.75 mM, CaCl<sub>2</sub> 2.5 mM, MgSO<sub>4</sub> 2.0 mM, KH<sub>2</sub>PO<sub>4</sub> 0.22mM, NaHCO<sub>3</sub> 26 mM and glucose 10 mM. porcine-NPY is dissolved in artificial cerebrospinal fluid (ACS). For i.c.v. injection the test compounds are preferably dissolved in DMSO/water (10%, v/v). The vehicle used for intraperitoneal (i.p.) , subcutaneous (s.c.) or oral (p.o.) delivery of compounds is preferably water, physiological saline or DMSO/water

-77-

(10% v/v), or cremophor/water (20% v/v), respectively.

Animals which are treated with both test compounds and p-NPY are treated first with the test compound. Then,  
5 10 min. after i.c.v. application of the test compound or vehicle (control), or 30-60 min after i.p., s.c. and p.o. application of the test compound or vehicle, 300 pmol of NPY is administered by intracerebroventricular (i.c.v.) application.

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Food intake may be measured by placing preweighed pellets into the cages at the time of NPY injection. Pellets are removed from the cage subsequently at each selected time point and replaced with a new set of  
15 preweighed pellets. The food intake of animals treated with test compound may be calculated as a percentage of the food intake of control animals, i.e., animals treated with vehicle. Alternatively, food intake for a group of animals subjected to the same experimental  
20 condition may be expressed as the mean  $\pm$  S.E.M. Statistical analysis is performed by analysis of variance using the Student-Newman-Keuls test.

#### Food intake in food-deprived rats

25

Food-deprivation experiments are conducted with male Sprague-Dawley rats weighing between 220 and 250 g. After receipt, the animals are individually housed for the duration of the study and allowed free access to normal food together with tap water. The animals are  
30 maintained in a room with a 12 h light/dark cycle (8:00 a.m. to 8:00 p.m. light) at 24 °C and monitored humidity. After placement into individual cages the rats undergo a 4 day equilibration period, during which they are habituated to their new environment and to  
35 eating a powdered or pellet diet (NAFAG, Gossau, Switzerland).

-78-

At the end of the equilibration period, food is removed from the animals for 24 hours starting at 8:00 a.m. At the end of the fasting period compound or vehicle may be administered to the animals orally or by injection intraperitoneally or intravenously. After 10 - 60 min. food is returned to the animals and their food intake monitored at various time periods during the following 24 hour period. The food intake of animals treated with test compound may be calculated as a percentage of the food intake of control animals (i.e., animals treated with vehicle). Alternatively, food intake for a group of animals subjected to the same experimental conditions may be expressed as the mean  $\pm$  S.E.M.

Food intake in obese Zucker rats

The antiobesity efficacy of the compounds according to the present invention might also be manifested in Zucker obese rats, which are known in the as an animal model of obesity. These studies are conducted with male Zucker fatty rats (fa/fa Harlan CPB, Austerlitz NL) weighing between 480g and 500g. Animals are individually housed in metabolism cages for the duration of the study and allowed free access to normal powdered food and water. The animals are maintained in a room with a 12 h light/dark cycle (light from 8:00 A.M. to 8:00 P.M.) at 24°C and monitored humidity. After placement into the metabolism cages the rats undergo a 6 day equilibration period, during which they are habituated to their new environment and to eating a powdered diet. At the end of the equilibration period, food intake during the light and dark phases is determined. After a 3 day control period, the animals are treated with test compounds or vehicle (preferably water or physiological saline or DMSO/water (10%, v/v) or cremophor/water (20%, v/v). Food intake is then monitored over the following 3 day period to determine the effect of administration of test compound



-79-

or vehicle alone. As in the studies described hereinabove, food intake in the presence of drug may be expressed as a percentage of the food intake of animals treated with vehicle.

5

### Materials

Cell culture media and supplements were from Specialty Media (Lavallette, NJ). Cell culture plates (150 mm and 96-well microtiter) were from Corning (Corning, NY).  
10 Sf9, Sf21, and High Five insect cells, as well as the baculovirus transfer plasmid, pBlueBacIII™, were purchased from Invitrogen (San Diego, CA). TMN-FH insect medium complemented with 10% fetal calf serum, and the baculovirus DNA, BaculoGold™, was obtained from  
15 Pharmingen (San Diego, CA). Ex-Cell 400™ medium with L-Glutamine was purchased from JRH Scientific. Polypropylene 96-well microtiter plates were from Costar (Cambridge, MA). All radioligands were from New England Nuclear (Boston, MA). Commercially available  
20 NPY and related peptide analogs were either from Bachem California (Torrance, CA) or Peninsula (Belmont, CA); [D-Trp<sup>32</sup>]NPY and PP C-terminal fragments were synthesized by custom order from Chiron Mimotopes Peptide Systems (San Diego, CA). Bio-Rad Reagent was  
25 from Bio-Rad (Hercules, CA). Bovine serum albumin (ultra-fat free, A-7511) was from Sigma (St. Louis, MO). All other materials were reagent grade.

-80-

**EXPERIMENTAL RESULTS**cDNA Cloning

In order to clone a rat hypothalamic "atypical" NPY receptor subtype, applicants used an expression cloning strategy in COS-7 cells (Gearing et al, 1989; Kluxen et al, 1992; Kiefer et al, 1992). This strategy was chosen for its extreme sensitivity since it allows detection of a single "receptor positive" cell by direct microscopic autoradiography. Since the "atypical" receptor has only been described in feeding behavior studies involving injection of NPY and NPY related ligands in rat hypothalamus (see introduction), applicants first examined its binding profile by running competitive displacement studies of  $^{125}\text{I}$ -PYY and  $^{125}\text{I}$ -PYY<sub>3-36</sub> on membranes prepared from rat hypothalamus. The competitive displacement data indicate: 1) Human PP is able to displace 20% of the bound  $^{125}\text{I}$ -PYY with an  $\text{IC}_{50}$  of 11 nM (Fig. 1 and Table 2). As can be seen in table 5, this value does not fit with the isolated rat Y1, Y2 and Y4 clones and could therefore correspond to another NPY/PYY receptor subtype. 2) [Leu<sub>31</sub>, Pro<sub>34</sub>] NPY (a Y1 specific ligand) is able to displace with high affinity ( $\text{IC}_{50}$  of 0.38) 27% of the bound  $^{125}\text{I}$ -PYY<sub>3-36</sub> ligand (a Y2 specific ligand) (Fig. 2 and table 2). These data provide the first evidence based on a binding assay that rat hypothalamic membranes could carry an NPY receptor subtype with a mixed Y1/Y2 pharmacology (referred to as the "atypical" subtype) which fits with the pharmacology defined in feeding behavior studies.

-81-

**TABLE 2: Pharmacological profile of the rat hypothalamus.**

Binding data reflect competitive displacement of  $^{125}\text{I}$ -PYY and  $^{125}\text{I}$ -PYY<sub>3-36</sub> from rat hypothalamic membranes. Peptides were tested at concentrations ranging from 0.001 nM to 100 nM unless noted. The IC<sub>50</sub> value corresponding to 50% displacement, and the percentage of displacement relative to that produced by 300 nM human NPY, were determined by nonlinear regression analysis. Data shown are representative of at least two independent experiments.

**TABLE 2**

Peptide	IC <sub>50</sub> Values, nM (% NPY-produced displacement)	
	$^{125}\text{I}$ -PYY	$^{125}\text{I}$ -PYY <sub>3-36</sub>
human NPY	0.82 (100%)	1.5 (100%)
human NPY <sub>2-36</sub>	2.3 (100%)	1.2 (100%)
human [Leu <sup>31</sup> , Pro <sup>34</sup> ]NPY	0.21 (44%) 340 (56%)	0.38 (27%) 250 (73%)
human PYY	1.3 (100%)	0.29 (100%)
human PP	11 (20%)	untested

Based on the above data, a rat hypothalamic cDNA library of  $3 \times 10^6$  independent recombinants with a 2.7 kb average insert size was fractionated into 450 pools of ~7500 independent clones. All pools were tested in a binding assay with  $^{125}\text{I}$ -PYY as previously described (U.S. Serial No. 08/192/288). Seven pools gave rise to positive cells in the screening assay (#'s 81, 92, 147, 246, 254, 290, 312). Since Y1, Y2, Y4 and Y5 receptor subtypes (by PCR or binding analysis) are expressed in

-82-

rat hypothalamus, applicants analyzed the DNA of positive pools by PCR with rat Y1, Y2 and Y4 specific primers. Pools # 147, 246, 254 and 312 turned out to contain cDNAs encoding a Y1 receptor, pool # 290 turned out to contain cDNA encoding a Y2 receptor subtype, but pools # 81 and 92 were negative by PCR analysis for Y1, Y2 and Y4 and therefore likely contained a cDNA encoding a new rat hypothalamic NPY receptor (Y5). Pools # 81 and 92 later turned out to contain an identical NPY receptor cDNA. Pool 92 was subjected to sib selection as described in U.S. Serial No. 08/192,288 until a single clone was isolated (designated CG-18).

The isolated clone carries a 2.8 kb cDNA. This cDNA contains an open reading frame between nucleotides 779 and 2146 that encodes a 456 amino acid protein. The long 5' untranslated region could be involved in the regulation of translation efficiency or mRNA stability. The flanking sequence around the putative initiation codon does not conform to the Kozak consensus sequence for optimal translation initiation (Kozak, 1989, 1991). The hydrophobicity plot displayed seven hydrophobic, putative membrane spanning regions which makes the rat hypothalamic Y5 receptor a member of the G-protein coupled superfamily. The nucleotide and deduced amino acid sequences are shown in Figures 3 and 4, respectively. Like most G-protein coupled receptors, the Y5 receptor contains consensus sequences for N-linked glycosylation in the amino terminus (position 21 and 28) involved in the proper expression of membrane proteins (Kornfeld and Kornfeld, 1985). The Y5 receptor carries two highly conserved cysteine residues in the first two extracellular loops that are believed to form a disulfide bond stabilizing the functional protein structure (Probst et al, 1992). The Y5 receptor shows 9 potential phosphorylation sites for

-83-

protein kinase C in positions 204, 217, 254, 273, 285, 301, 328, 336 and 409; and 2 cAMP- and cGMP-dependent protein kinase phosphorylation sites in positions 298 and 370. It should be noted that 8 of these 11 potential phosphorylation sites are located in the third intra-cellular loop, two in the second intra-cellular loop and one in the carboxy terminus of the receptor and could, therefore, play a role in regulating functional characteristics of the Y5 receptor (Probst et al, 1992). In addition, the rat Y5 receptor carries a leucine zipper motif in its first putative transmembrane domain (Landschulz et al, 1988). A tyrosine kinase phosphorylation site is found in the middle of the leucine zipper.

Localization studies (see below) show that the Y5 mRNA is present in several areas of the rat hippocampus. Assuming a comparable localization in human brain, applicants screened a human hippocampal cDNA library as described in U.S. Serial No. 08/192,288 with rat oligonucleotide primers which were shown to yield a DNA band of the expected size in a PCR reaction run on human hippocampal cDNA (C. Gerald, unpublished results). Using this PCR screening strategy (Gerald et al, 1994, submitted for publication), three positive pools were identified. One of these pools was analyzed further, and an isolated clone was purified by sib selection. The isolated clone (CG-19) turned out to contain a full length cDNA cloned in the correct orientation for functional expression (see below). The human Y5 nucleotide and deduced amino acid sequences are shown in Figures 5 and 6, respectively. When compared to the rat Y5 receptor, the human sequence shows 84.1% nucleotide identity (Fig. 7A to 7E) and 87.2% amino acid identity (Fig. 7F and 7G). The rat protein sequence is one amino acid longer at the very end of both amino and carboxy tails of the receptor

-84-

when compared to the rat. The human 5-6 loop is one amino acid longer than the rat and shows multiple non conservative substitutions. Even though the 5-6 loops show significant changes between the rat and human homologs, all of the protein motifs found in the rat receptor are present in the human homolog. All putative transmembrane domains and extra cellular loop regions are highly conserved (Fig. 7F and 7G). Therefore, both pharmacological profiles and functional characteristics of the rat and human Y5 receptor subtype homologs may be expected to match closely.

When the human and rat Y5 receptor sequences were compared to other NPY receptor subtypes or to other human G protein-coupled receptor subtypes, both overall and transmembrane domain identities are very low, showing that the Y5 receptor genes are not closely related to any other previously characterized cDNAs. Even among the human NPY receptor family, Y1, Y2, Y4 and Y5 members show unusually low levels of amino acid identity (Fig. 8A through 8C).

**TABLE 3: Human Y5 transmembrane domains identity with other human NPY receptor subtypes and other human G-protein coupled receptors**

	<u>Receptor subtype</u>	<u>% TM identity</u>
	Y-4	40
	Y-2	42
	Y-1	42
30	MUSGIR	32
	DroNPY	31
	Beta-1	30
	Endothelin-1	30
	Dopamine D2	29
35	Adenosine A2b	28
	Subst K	28
	Alpha-2A	27
	5-HT1Dalpha	26
	Alpha-1A	26
40	IL-8	26
	5-HT2	25
	Subst P	24

-85-

Northern blot analysis

Using the rat Y5 probe, northern hybridizations reveal a strong signal at 2.7 kb and a weak band at 8 kb in rat whole brain. A weak signal is observed at 2.7 kb in  
5 testis. No signal was seen in heart, spleen, lung, liver, skeletal muscle and kidney after a three day exposure (Figure 16A). This is in good agreement with the 2.7 kb cDNA that we isolated by expression cloning  
10 is full length. The 8 kb band seen in whole brain probably corresponds to unspliced pre-mRNA.

With the human Y5 probe, northern hybridizations (Figures 16B and 16C) showed a strong signal at 3.5 kb  
15 with a much weaker band at 2.2 and 1.1 kb in caudate nucleus, putamen and cerebral cortex, a medium signal in frontal lobe and amygdala and a weak signal in hippocampus, occipital and temporal lobes, spinal cord, medulla, thalamus, subthalamic nucleus, and substantia  
20 nigra. No signal at 3.5 kb was detectable in cerebellum or corpus callosum after a 48 h exposure. It should be noted that Clontech's MTN II and III blots do not carry any mRNA from hypothalamus, periaquiductalgray, superior colliculus and raphe.

25 Southern blot analysis on human genomic DNA reveals a unique band pattern in 4 of the 5 restriction digests (Figure 17A). The two bands observed in the PstI digest can be explained by the presence of a PstI site in the  
30 coding region of the human Y5 gene. Rat southern blotting analysis showed a unique band pattern in all five restriction digests tested (Figure 17B). These analyses are consistent with the human and rat genomes containing a single copy of the Y5 receptor gene.

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Canine Y5 homolog

-86-

The canine nucleotide sequence obtained to date (PCR and 3' RACE products) spans the canine Y5 receptor from the first extracellular loop immediately upstream of TM III into the 3' untranslated region (Figure 14). In the coding region, this nucleotide sequence is highly identical to both the human and the rat sequences (91% and 83.3% respectively). The deduced canine Y5 amino acid sequence is shown in Figure 15. This amino acid sequence is again highly identical to both the human and rat Y5 sequences (94.6% and 89.5% respectively), with most amino acid changes located in the 5-6 loop. Therefore the pharmacological profile of the canine Y5 receptor subtype is expected to closely resemble the human and rat Y5 profiles.

#### Binding Studies

The cDNA for the rat hypothalamic Y5 receptor was transiently expressed in COS-7 cells for full pharmacological evaluation.  $^{125}\text{I}$ -PYY bound specifically to membranes from COS-7 cells transiently transfected with the rat Y5 receptor construct. The time course of specific binding was measured in the presence of 0.08 nM  $^{125}\text{I}$ -PYY at 30 °C (Fig. 9). The association curve was monophasic, with an observed association rate ( $K_{\text{obs}}$ ) of 0.06 min<sup>-1</sup> and a  $t_{1/2}$  of 11 min; equilibrium binding was 99% complete within 71 min and stable for at least 180 min. All subsequent binding assays were carried out for 120 min at 30 °C. The binding of  $^{125}\text{I}$ -PYY to transiently expressed rat Y5 receptors was saturable over a radioligand concentration range of 0.4 pM to 2.7 nM. Binding data were fit to a one-site binding model with an apparent  $K_d$  of 0.29 nM ( $\text{p}K_d = 9.54 \pm 0.13$ ,  $n = 4$ ). A receptor density of between 5 and 10 pmol/mg membrane protein was measured on membranes which had been frozen and stored in liquid nitrogen (Fig. 10). Membranes from mock-transfected cells, when prepared and analyzed in the same way as those from CG-



- 87 -

18-transfected cells, displayed no specific binding of  $^{125}\text{I}$ -PYY (data not shown). Applicants conclude that the  $^{125}\text{I}$ -PYY binding sites observed under the described conditions were derived from the rat Y5 receptor construct.

A closely related peptide analog, porcine  $^{125}\text{I}$ -[Leu<sup>31</sup>,Pro<sup>34</sup>]PYY, also bound specifically to membranes from COS-7 cells transiently transfected with rat Y5 receptor cDNA. The time course of specific binding was measured at room temperature in both standard binding buffer ( $[\text{Na}^+] = 10 \text{ mM}$ ) and isotonic binding buffer ( $[\text{Na}^+] = 138 \text{ mM}$ ) using  $0.08 \text{ nM } ^{125}\text{I}$ -[Leu<sup>31</sup>,Pro<sup>34</sup>]PYY (Figure 18). The association curve in  $10 \text{ mM } [\text{Na}^+]$  was monophasic, with an observed association rate ( $K_{\text{obs}}$ ) of  $0.042 \text{ min}^{-1}$  and a  $t_{1/2}$  of  $17 \text{ min}$ ; equilibrium binding was 99% complete within  $110 \text{ min}$  and stable for at least  $210 \text{ min}$  (specific binding was maximal at  $480 \text{ fmol/mg membrane protein}$ ). The association curve in  $138 \text{ mM } [\text{Na}^+]$  was also monophasic with a slightly slower time course: ( $K_{\text{obs}}$ ) of  $0.029 \text{ min}^{-1}$  and a  $t_{1/2}$  of  $24 \text{ min.}$ ; equilibrium binding was 99% complete within  $160 \text{ min.}$  and stable for at least  $210 \text{ min.}$  (specific binding was maximal at  $330 \text{ fmol/mg membrane protein}$ ). Note that the specific binding was reduced as  $[\text{Na}^+]$  was increased; a similar phenomenon has been observed for other G protein coupled receptors and may reflect a general property of this receptor family to be modulated by  $\text{Na}^+$  (Horstman et al., 1990). Saturation binding studies were performed with  $^{125}\text{I}$ -[Leu<sup>31</sup>,Pro<sup>34</sup>]PYY in isotonic buffer at room temperature over a  $120 \text{ minute period}$ . Specific binding to transiently expressed rat Y5 receptors was saturable over a radioligand concentration range of  $0.6 \text{ pM}$  to  $1.9 \text{ nM}$ . Binding data were fit to a one-site binding model with an apparent  $K_d$  of  $0.072 \text{ nM}$  ( $\text{pKd} = 10.14 \pm 0.07$ ,  $n = 2$ ). A receptor density of  $560 \pm 150 \text{ pmol/mg}$  on

-88-

membranes which had been frozen and stored in liquid nitrogen. That  $^{125}\text{I}$ -[Leu<sup>31</sup>,Pro<sup>34</sup>]PYY can bind to the rat Y5 receptor with high affinity at room temperature in isotonic buffer makes it a potentially useful ligand for characterizing the native Y5 receptor in rat tissues using autoradiographic techniques. Care must be taken, however, to use appropriate masking agents to block potential radiolabeling of other receptors such as Y1 and Y4 receptors (note in Table 5 that rat Y1 and Y4 bind the structural homolog [Pro<sup>34</sup>]PYY). Previously published reports of  $^{125}\text{I}$ -[Leu<sup>31</sup>,Pro<sup>34</sup>]PYY as a Y1-selective radioligand should be re-evaluated in light of new data obtained with the rat Y5 receptor (Dumont, et al., 1995).

The pharmacological profile of the rat Y5 receptor was first studied by using pancreatic polypeptide analogs in membrane binding assays. The rank order of affinity for selected compounds was derived from competitive displacement of  $^{125}\text{I}$ -PYY (Fig. 11). The rat Y5 receptor was compared with cloned Y1, Y2, and Y4 receptors from human (Table 4) and rat (Table 5), all expressed transiently in COS-7 cells. One receptor subtype absent from our panel was the Y3, human or rat, as no model suitable for radioligand screening has yet been identified.

**TABLE 4: Pharmacological profile of the rat Y5 receptor vs. Y-type receptors cloned from human.**

Binding data reflect competitive displacement of  $^{125}\text{I}$ -PYY from membranes of COS-7 cells transiently expressing rat Y5 and human subtype clones. Peptides were tested at concentrations ranging from 0.001 nM to 1000 nM unless noted.  $\text{IC}_{50}$  values corresponding to 50% displacement were determined by nonlinear regression analysis and converted to  $K_i$  values according to the Cheng-Prusoff equation. The data shown are

- 89 -

representative of at least two independent experiments.

**TABLE 4**

	Peptide	K <sub>i</sub> Values (nM)			
		Rat Y5	Human Y4	Human Y1	Human Y2
5	rat/human NPY	0.68	2.2	0.07	0.74
	porcine NPY	0.66	1.1	0.05	0.81
	human NPY <sub>2-36</sub>	0.86	16	3.9	2.0
10	porcine NPY <sub>2-36</sub>	1.2	5.6	2.4	1.2
	porcine NPY <sub>13-36</sub>	73	38	60	2.5
15	porcine NPY <sub>26-36</sub>	> 1000	304	> 1000	380
	porcine C2-NPY	470	120	79	3.5
20	human [Leu <sup>31</sup> , Pro <sup>34</sup> ] NPY	1.0	1.1	0.17	> 130
	human [D-Trp <sup>32</sup> ] NPY	53	> 760	> 1000	> 1000
	human NPY free acid	480	> 1000	490	> 1000
25	rat/porcine PYY	0.64	0.14	0.35	1.26
	human PYY	0.87	0.87	0.18	0.36
	human PYY <sub>3-36</sub>	8.4	15	41	0.70
30	human PYY <sub>13-36</sub>	190	46	33	1.5
	human [Pro <sup>34</sup> ] PYY	0.52	0.12	0.14	> 310
	human PP	5.0	0.06	77	> 1000
35	human PP <sub>2-36</sub> *	not tested	0.06	> 40	> 100

Table 4 continued

-91-

human PP <sub>13-36</sub> *	<b>not tested</b>	39	> 100	> 100
rat PP	<b>180</b>	0.16	450	> 1000
salmon PP	<b>0.31</b>	3.2	0.11	0.17

5 \*Tested only up to 100 nM.

-92-

**TABLE 5: Pharmacological profile of the rat Y5 receptor vs. Y-type receptors cloned from rat.**

5 Binding data reflect competitive displacement of  $^{125}\text{I}$ -PYY from membranes of COS-7 cells transiently expressing rat Y5 and rat subtype clones. Peptides were tested at concentrations ranging from 0.001 nM to 1000 nM.  $\text{IC}_{50}$  values corresponding to 50% displacement were  
 10 determined by nonlinear regression analysis and converted to  $K_i$  values according to the Cheng-Prusoff equation. The data shown are representative of at least two independent experiments. Exception: new peptides (marked with a double asterisk) were tested in one or  
 15 more independent experiments.

**TABLE 5**

Peptide	$K_i$ Values (nM)			
	Rat Y5	Rat Y4	Rat Y1	Rat Y2
20 rat/human NPY	0.68	1.7	0.12	1.3
porcine NPY **	0.66	1.78	0.06	1.74
25 frog NPY ** (melanostatin)	0.71		0.09	0.65
human $\text{NPY}_{2-36}$	0.86	5.0	12	2.6
porcine $\text{NPY}_{2-36}$ **	1.1	18	1.6	1.6
30 porcine $\text{NPY}_{3-36}$ **	7.7	36	91	3.7
porcine $\text{NPY}_{13-36}$	73	140	190	31
35 porcine $\text{NPY}_{16-36}$ **	260	200	140	35
porcine $\text{NPY}_{18-36}$ **	> 1000		470	12

Table 5 Continued

-93-

	Peptide	K <sub>i</sub> Values (nM)			
		Rat Y5	Rat Y4	Rat Y1	Rat Y2
	porcine NPY <sub>20-36</sub> **	> 100		360	93
	porcine NPY <sub>22-36</sub> **	> 1000		> 1000	54
5	porcine NPY <sub>26-36</sub> **	> 1000		> 1000	> 830
	human [Leu <sup>31</sup> , Pro <sup>34</sup> ] NPY	1.0	0.59	0.10	> 1000
10	porcine ** [Leu <sup>31</sup> , Pro <sup>34</sup> ] NPY	1.6	0.32	0.25	840
15	human (O- Methyl- Tyr <sup>21</sup> ) NPY **	1.6			2.3
	human NPY free acid **	> 610	> 1000	720	> 980
	porcine C2- NPY **	> 260	22	140	2.6
20	human NPY <sub>1-24</sub> amide **	> 1000		> 320	> 1000
	human [D- Trp <sup>32</sup> ] NPY	35	> 630	> 1000	760
25	rat/porcine PYY	0.64	0.58	0.21	0.28
	human PYY **	0.87		0.12	0.30
	human PYY <sub>3-36</sub> **	8.4	15		0.48
30	human PYY <sub>13-36</sub> **	290		130	14
	human [Pro <sup>34</sup> ] PYY	0.52	0.19	0.25	> 1000
35	porcine [Pro <sup>34</sup> ] PYY **	0.64	0.24	0.07	> 980

Table 5 Continued

-94-

Peptide	K <sub>i</sub> Values (nM)			
	Rat Y5	Rat Y4	Rat Y1	Rat Y2
avian PP **	> 930	> 81	> 320	> 1000
human PP	5.0	0.04	43	> 1000
human PP <sub>13-36</sub> **	84		> 1000	> 650
5 human PP <sub>31-36</sub> **	> 1000	26	> 10 000	> 10 000
human PP <sub>31-36</sub> free acid **	>10,000	> 100		
bovine PP **	8.4	0.19	120	> 1000
10 frog PP (rana temporaria) **	> 550	> 1000	720	> 980
rat PP	230	0.19	350	> 1000
15 salmon PP	0.33	3.0	0.30	0.16
PYX-1 **	920			
PYX-2 **	> 1000			
FLRF-amide **	5500		45 000	
20 FMRF-amide **	18000			
W(nor-L)RF- amide **	8700			

25

30

The rat Y5 receptor possessed a unique pharmacological profile when compared with human and rat Y-type receptors. It displayed a preference for structural analogs of rat/human NPY (K<sub>i</sub> = 0.68 nM) and rat/porcine PYY (K<sub>i</sub> = 0.64 nM) over most PP derivatives. The high affinity for salmon PP (K<sub>i</sub> = 0.31 nM) reflects the close similarity between salmon PP and rat NPY, sharing 81% of their amino acid sequence and maintaining identity at key positions: Tyr<sup>1</sup>, Gln<sup>34</sup>, and Tyr<sup>36</sup>. Both



-95-

N- and C-terminal peptide domains are apparently important for receptor recognition. The N-terminal tyrosine of NPY or PYY could be deleted without an appreciable loss in binding affinity ( $K_i = 0.86$  nM for rat/human NPY<sub>2-36</sub>), but further N-terminal deletion was disruptive ( $K_i = 73$  nM for porcine NPY<sub>13-36</sub>). This pattern places the binding profile of the Y5 receptor somewhere between that of the Y2 receptor (which receptor can withstand extreme N-terminal deletion) and that of the Y1 receptor (which receptor is sensitive to even a single-residue N-terminal deletion). Note that the human Y4 receptor can be described similarly ( $K_i = 0.06$  nM for human PP,  $0.06$  nM for human PP<sub>2-36</sub>, and  $39$  nM for human PP<sub>13-36</sub>). The Y5 receptor resembled both Y1 and Y4 receptors in its tolerance for ligands containing Pro<sup>34</sup> (as in human [Leu<sup>31</sup>,Pro<sup>34</sup>]NPY, human [Pro<sup>34</sup>]-PYY, and human PP). Interestingly, the rat Y5 receptor displayed a preference for human PP ( $K_i = 5.0$  nM) over rat PP ( $K_i = 180$  nM). This pattern distinguishes the rat Y5 from the rat Y4 receptor, which binds both human and rat PP with  $K_i$  values  $< 0.2$  nM. Hydrolysis of the carboxy terminal amide to free carboxylic acid, as in NPY free acid, was disruptive for binding affinity for the rat Y5 receptor ( $K_i = 480$  nM). The terminal amide appears to be a common structural requirement for pancreatic polypeptide family/receptor interactions.

Several peptides shown previously to stimulate feeding behavior in rats bound to the rat Y5 receptor with  $K_i \leq 5.0$  nM. These include rat/human NPY ( $K_i = 0.68$  nM), rat/porcine PYY ( $K_i = 0.64$  nM), rat/human NPY<sub>2-36</sub> ( $K_i = 0.86$  nM), rat/human [Leu<sup>31</sup>,Pro<sup>34</sup>]NPY ( $K_i = 1.0$  nM), and human PP ( $K_i = 5.0$  nM). Conversely, peptides which were relatively less effective as orexigenic agents bound weakly to CG-18. These include porcine NPY<sub>13-36</sub> ( $K_i = 73$  nM), porcine C2-NPY ( $K_i = 470$  nM) and human NPY

-96-

free acid ( $K_i = 480$  nM). The rank order of  $K_i$  values are in agreement with rank orders of potency and activity for stimulation of feeding behavior when peptides are injected i.c.v. or directly into rat hypothalamus (Clark et al., 1984; Stanley et al., 1985; Kalra et al., 1991; Stanley et al., 1992). The rat Y5 receptor also displayed moderate binding affinity for [D-Trp<sup>32</sup>]NPY ( $K_i = 53$  nM), the modified peptide reported to regulate NPY-induced feeding by Balasubramaniam and co-workers (1994). It is noteworthy that [D-Trp<sup>32</sup>]NPY was  $\geq 10$ -fold selective for CG-18 over the other cloned receptors studied, whether human or rat. These data clearly and definitively link the cloned Y5 receptor to the feeding response.

The cDNA corresponding to the human Y5 homolog isolated from human hippocampus was transiently expressed in COS-7 cells for membrane binding studies. The binding of <sup>125</sup>I-PYY to the human Y5 receptor (CG-19) was saturable over a radioligand concentration range of 8 pM to 1.8 nM. Binding data were fit to a one-site binding model with an apparent  $K_d$  of 0.10 nM in the first experiment. Repeated testing yielded an apparent  $K_d$  of 0.18 nM ( $pK_d = 9.76 \pm 0.11$ ,  $n = 4$ ). A maximum receptor density of 500 fmol/mg membrane protein was measured on fresh membranes. As determined by using peptide analogs within the pancreatic polypeptide family, the human Y5 pharmacological profile bears a striking resemblance to the rat Y5 receptor (Tables 6 and 7).

-97-

**TABLE 6: Pharmacological profile of the rat Y5 receptor vs. the human Y5 receptor, as expressed both transiently in COS-7 and stably in LM(tk-) cells.**

5 Binding data reflect competitive displacement of radioligand (either  $^{125}\text{I}$ -PYY or  $^{125}\text{I}$ -PYY<sub>3-36</sub> as indicated) from membranes of COS-7 cells transiently expressing the rat Y5 receptor and its human homolog or from  
10 LM(tk-) cells stably expressing the human Y5 receptor. Peptides were tested at concentrations ranging from 0.001 nM to 1000 nM. IC<sub>50</sub> values corresponding to 50% displacement were determined by nonlinear regression analysis and converted to K<sub>i</sub> values according to the  
15 Cheng-Prusoff equation. New peptides are marked with a double asterisk.

**TABLE 6**

Peptide	K <sub>i</sub> Values (nM)			
	Rat Y5 (COS-7, $^{125}\text{I}$ -PYY)	Human Y5 (COS-7, $^{125}\text{I}$ -PYY)	Human Y5 (LM(tk-), $^{125}\text{I}$ -PYY)	Human Y5 (LM(tk-), $^{125}\text{I}$ -PYY <sub>3-36</sub> )
rat/human NPY	0.68	0.15	0.89	0.65
25 porcine NPY **		0.68	1.4	
human NPY <sub>2-36</sub>	0.86	0.33	1.6	0.51
30 porcine NPY <sub>2-36</sub> **	0.66	0.58	1.2	
porcine NPY <sub>13-36</sub>	73	110		39
porcine NPY <sub>16-36</sub> **	260	300		180
35 porcine NPY <sub>18-36</sub> **	> 1000	> 470		310

Table 6 continued

-98-

Peptide	K <sub>i</sub> Values (nM)			
	Rat Y5 (COS-7, <sup>125</sup> I-PYY)	Human Y5 (COS-7, <sup>125</sup> I-PYY)	Human Y5 (LM(tk-), <sup>125</sup> I-PYY)	Human Y5 (LM(tk-), <sup>125</sup> I-PYY <sub>3-36</sub> )
porcine NPY <sub>22-36</sub> **	> 1000	> 1000		
porcine NPY <sub>26-36</sub> **	> 1000	> 1000		
5 human [Leu <sup>31</sup> , Pro <sup>34</sup> ] NPY	1.0	0.72	3.0	
10 human [Leu <sup>31</sup> , Pro <sup>34</sup> ] NPY **			2.4	1.4
15 human NPY free acid **	> 610	> 840		
porcine C2-NPY **	260	370	260	220
human [D- Trp <sup>32</sup> ] NPY	35	35	16	10
20 rat/porci ne PYY	0.64	0.75		
human PYY **	0.87	0.44	1.3	0.43
25 human PYY <sub>3-36</sub> **	8.4	17	8.1	1.6
human [Pro <sup>34</sup> ] PYY	0.52	0.34	1.7	1.7
human PP	5.0	1.7	3.0	1.2
30 human PP <sub>2-36</sub> **		2.1		
human PP <sub>13-36</sub> **	290	720		

Table 6 continued

-99-

Peptide	K <sub>i</sub> Values (nM)			
	Rat Y5 (COS- 7, <sup>125</sup> I- PYY)	Human Y5 (COS-7, <sup>125</sup> I- PYY)	Human Y5 (LM(tk- ) , <sup>125</sup> I- PYY)	Human Y5 (LM(tk- ) , <sup>125</sup> I- PYY <sub>3-36</sub> )
human PP <sub>31-36</sub> **	> 10 000	> 10 000		41 000
human [Ile <sup>31</sup> , Gln <sup>34</sup> ] PP **		2.0		
bovine PP **	8.4	1.6	7.9	5.0
rat PP	230	630		130
salmon PP	0.33	0.27		0.63

5

10

-100-

**TABLE 7: Pharmacological profile of the human Y5 receptor vs. Y-type receptors cloned from human.**

5 Binding data reflect competitive displacement of  $^{125}\text{I}$ -PYY from membranes of COS-7 cells transiently expressing human Y5 other sub-type clones. Peptides were tested at concentrations ranging from 0.001 nM to 1000 nM unless noted.  $\text{IC}_{50}$  values corresponding to 50% displacement were determined by nonlinear regression analysis and converted to  $K_i$  values according to the Cheng-Prusoff equation. The data shown are representative of at least two independent experiments.

TABLE 7

Peptide	$K_i$ Values (nM)			
	Human Y5	Human Y4	Human Y1	Human Y2
rat/human NPY	0.46	2.2	0.07	0.74
porcine NPY	0.68	1.1	0.05	0.81
human NPY <sub>2-36</sub>	0.75	16	3.9	2.0
20 porcine NPY <sub>2-36</sub>	0.58	5.6	2.4	1.2
porcine NPY <sub>13-36</sub>	110	38	60	2.5
porcine NPY <sub>26-36</sub>	> 1000	304	> 1000	380
porcine C2-NPY	370	120	79	3.5
25 human [Leu <sup>31</sup> , Pro <sup>34</sup> ]NPY	1.6	1.1	0.17	> 130
human [D-Trp <sup>32</sup> ]NPY	35	> 760	> 1000	> 1000
human NPY free acid	> 840	> 1000	490	> 1000
30 rat/porcine PYY	0.58	0.14	0.35	1.26
human PYY	0.44	0.87	0.18	0.36
human PYY <sub>3-36</sub>	17	15	41	0.70

Table 7 Continued

-101-

Peptide	K <sub>i</sub> Values (nM)			
	Human Y5	Human Y4	Human Y1	Human Y2
human PYY <sub>13-36</sub>	<b>not tested</b>	46	33	1.5
human [Pro <sup>34</sup> ] PYY	<b>0.77</b>	0.12	0.14	> 310
human PP	<b>1.4</b>	0.06	77	> 1000
human PP <sub>2-36</sub> *	<b>2.1</b>	0.06	> 40	> 100
human PP <sub>13-36</sub> *	<b>720</b>	39	> 100	> 100
rat PP	<b>630</b>	0.16	450	> 1000
salmon PP	<b>0.46</b>	3.2	0.11	0.17

\*Tested only up to 100 nM.

-102-

Binding Studies of hY5 Expressed in Insect Cells

Tests were initially performed to optimize expression of hY5 receptor. Infecting Sf9, Sf21, and High Five cells with hY5BB3 virus at a multiplicity of infection (MOI) of 5 and preparing membranes for binding analyses at 45 hours postinfection, we observed  $B_{\max}$  ranges from 417 to 820 fmoles/mg protein, with the highest expression being hY5BB3 in Sf21 cells. Therefore, our next series of experiments used Sf21 cells. We next examined optimal multiplicity of infection (MOI, the ratio of viral particles to cells) by testing MOI of 1, 2, 5 and 10. The  $B_{\max}$  values were ~1.1-1.2 pmoles/mg protein for any of the MOIs, suggesting that increasing the number of viral particles per cell is neither deleterious nor advantageous. Since viral titer calculations are approximate, we used MOI=5 for future experiments. The last parameter we tested was hours postinfection for protein expression, ranging from 45-96 hours postinfection. We found that optimal expression occurred 45-73 hours postinfection. In summary, we have created a hY5 recombinant baculovirus which binds  $^{125}\text{I}$ -PYY with a  $B_{\max}$  of ~1.2 pmoles/mg protein.

Human Y5 Homolog: Transient Expression in Baculovirus-Infected Sf21 Insect Ovary Cells

Sf21 cells infected with a human Y5 baculovirus construct were harvested as membrane homogenates and screened for specific binding of  $^{125}\text{I}$ -PYY using 0.08 nM radioligand. Specific binding was greatest (500 fmol/mg membrane protein) for sample D-2/[4], derived from Sf-21 cells. No specific binding was observed after infection with the baculovirus plasmid alone (data not shown). If we make the assumption that the binding affinity of porcine  $^{125}\text{I}$ -PYY for the human Y5 receptor is the same whether the expression system is COS-7 or baculovirus/Sf-21 (0.18 nM), the specific binding in



-103-

sample D-2/[4] predicts an apparent  $B_{max}$  of 1600 fmol/mg membrane protein. The Y5 receptor yield in the baculovirus/Sf21 expression system is therefore as good or better than that in COS-7. We conclude that the baculovirus offers an alternative transfection technique amenable to large batch production of the human Y5 receptor.

#### Stable Expression Systems for Y5 Receptors:

##### Characterization in Binding Assays

The cDNA for the rat Y5 receptor was stably transfected into 293 cells which were pre-screened for the absence of specific  $^{125}\text{I}$ -PYY binding (data not shown). After co-transfection with the rat Y5 cDNA plus a G-418-resistance gene and selection with G-418, surviving colonies were screened as membrane homogenates for specific binding of  $^{125}\text{I}$ -PYY using 0.08 nM radioligand. A selected clone (293 clone # 12) bound 65 fmol  $^{125}\text{I}$ -PYY /mg membrane protein and was isolated for further study in functional assays.

The cDNA for the human Y5 receptor was stably transfected into both NIH-3T3 and LM(tk-) cells, each of which were pre-screened for the absence of specific  $^{125}\text{I}$ -PYY binding (data not shown). After co-transfection with the human Y5 cDNA plus a G-418-resistance gene and selection with G-418, surviving colonies were screened as membrane homogenates for specific binding of  $^{125}\text{I}$ -PYY using 0.08 nM radioligand. NIH-3T3 clone #8 bound 46 fmol  $^{125}\text{I}$ -PYY/mg membrane protein and LM(tk-) clone #7 bound 32 fmol  $^{125}\text{I}$ -PYY/mg membrane protein. These two clones were isolated for further characterization in binding and cAMP functional assays. A third clone which bound 25 fmol/mg membrane protein, LM(tk-) #3, was evaluated in calcium mobilization assays.

-104-

5 The human Y5 stably expressed in NIH-3T3 cells (clone #8) was further characterized in saturation binding assays using  $^{125}\text{I}$ -PYY. The binding was saturable over a concentration range of 0.4 pM to 1.9 nM. Binding data were fit to a one-site binding model with an apparent  $K_d$  of 0.30 nM ( $\text{p}K_d = 9.53$ ,  $n = 1$ ) and an apparent  $B_{\text{max}}$  of 2100 fmol/mg membrane protein using fresh membranes.

10 The human Y5 stably expressed in LM(tk-) cells (clone #7) was further characterized in saturation binding assays using  $^{125}\text{I}$ -PYY,  $^{125}\text{I}$ -PYY<sub>3-36</sub>, and  $^{125}\text{I}$ -NPY.  $^{125}\text{I}$ -PYY binding was saturable according to a 1-site model over a concentration range of 0.4 pM to 1.9 nM, with an apparent  $K_d$  of 0.47 nM ( $\text{p}K_d = 9.32 \pm 0.07$ ,  $n = 5$ ) and an apparent  $B_{\text{max}}$  of up to 8 pmol/mg membrane protein when membranes had been frozen and stored in liquid nitrogen. Peptide  $K_i$  values derived from  $^{125}\text{I}$ -PYY binding to human Y5 receptors from LM(tk-) were comparable to those derived from the previously described human and rat Y5 expression systems (Table 6).  $^{125}\text{I}$ -PYY<sub>3-36</sub> binding to the human Y5 in LM(tk-) cells was also saturable according to a 1-site model over a concentration range of 0.5 pM to 2.09 nM, with an apparent  $K_d$  of 0.40 nM ( $\text{p}K_d = 9.40$ ,  $n = 1$ ) and an apparent  $B_{\text{max}}$  of 490 fmol/mg membrane protein when membranes had been frozen and stored in liquid nitrogen. Peptide ligands appeared to bind with comparable affinity to human Y5 receptors in LM(tk-) cells whether the radioligand used was  $^{125}\text{I}$ -PYY or  $^{125}\text{I}$ -PYY<sub>3-36</sub> (Table 6). Finally,  $^{125}\text{I}$ -NPY binding to the human Y5 in LM(tk-) cells was saturable according to a 1-site model over a concentration range of 0.4 pM to 1.19 nM, with an apparent  $K_d$  of 0.28 and an apparent  $B_{\text{max}}$  of 360 fmol/mg membrane protein when membranes had been frozen and stored in liquid nitrogen.

Considering the saturation binding studies for the

-105-

human and rat Y5 receptor homologs as a whole, the data provide evidence that the Y5 receptor is a target for multiple radioiodinated peptide analogs in the pancreatic polypeptide family, including  $^{125}\text{I}$ -PYY,  $^{125}\text{I}$ -NPY,  $^{125}\text{I}$ -PYY<sub>3-36</sub>, and  $^{125}\text{I}$ -[Leu<sup>31</sup>,Pro<sup>34</sup>]PYY. The so-called Y1 and Y2-selective radioligands such as  $^{125}\text{I}$ -[Leu<sup>31</sup>,Pro<sup>34</sup>]PYY and  $^{125}\text{I}$ -PYY<sub>3-36</sub>, respectively (Dumont, et al., 1995) should be used with caution when probing native tissues for Y-type receptor expression.

10

#### Receptor/G protein Interactions: Effects of Guanine Nucleotides

For a given G protein-coupled receptor, a portion of the receptor population can typically be characterized in the high affinity ligand binding site using discriminating agonists. The binding of GTP or a non-hydrolyzable analog to the G protein causes a conformational change in the receptor which favors a low affinity ligand binding state. We investigated whether the non-hydrolyzable GTP analog, Gpp(NH)p, would alter the binding of  $^{125}\text{I}$ -PYY to Y5 in COS-7 and LM(tk-) cells (Fig 19).  $^{125}\text{I}$ -PYY binding to both human and rat Y5 receptors in COS-7 cells was relatively insensitive to increasing concentrations of Gpp(NH)p ranging from 1 nM to 100  $\mu\text{M}$ . The human Y5 receptor in LM(tk-) cells, however, displayed a concentration dependent decrease in radioligand binding (-85 fmol/mg membrane protein over the entire concentration range). The difference between the receptor preparations could be explained by several factors, including 1) the types of G proteins available in the host cell for supporting a high affinity receptor-agonist complex, 2) the level of receptor reserve in the host cell, and 3) the efficiency of receptor/G protein coupling, and 4) the intrinsic ability of the agonist (in this case,  $^{125}\text{I}$ -PYY) to distinguish between multiple conformations of the receptor.

-106-

Functional Assay

Activation of all Y-type receptors described thus far is thought to involve coupling to pertussis toxin-sensitive G-proteins which are inhibitory for adenylate cyclase activity ( $G_i$  or  $G_o$ ) (Wahlestedt and Reis, 1993). That the atypical Y1 receptor is linked to cyclase inhibition was prompted by the observation that pertussis toxin inhibited NPY-induced feeding *in vivo* (Chance et al., 1989); a more definitive analysis was impossible in the absence of the isolated receptor. Based on these prior observations, applicants investigated the ability of NPY to inhibit forskolin-stimulated cAMP accumulation in human embryonic kidney 293 cells stably transfected with rat Y5 receptors. Incubation of intact cells with 10  $\mu$ M forskolin produced a 10-fold increase in cAMP accumulation over a 5 minute period, as determined by radioimmunoassay. Simultaneous incubation with rat/human NPY decreased the forskolin-stimulated cAMP accumulation by 67% in stably transfected cells (Fig. 12), but not in untransfected cells (data not shown). Applicants conclude that the rat Y5 receptor activation results in decreased cAMP accumulation, very likely through inhibition of adenylate cyclase activity. This result is consistent with the proposed signalling pathway for all Y-type receptors and for the atypical Y1 receptor in particular.

Peptides selected for their ability to stimulate feeding behavior in rats were able to activate the rat Y5 receptor with  $EC_{50} < 10$  nM (Kalra et al., 1991; Stanley et al., 1992; Balasubramaniam et al., 1994). These include rat/human NPY ( $EC_{50} = 1.8$  nM), rat/human NPY<sub>2-36</sub> ( $EC_{50} = 2.0$  nM), rat/human [Leu<sup>31</sup>, Pro<sup>34</sup>]NPY ( $EC_{50} = 0.6$  nM), rat/porcine PYY ( $EC_{50} = 4.0$  nM), and rat/human [D-Trp<sup>32</sup>]NPY ( $EC_{50} = 7.5$  nM) (Table 8).  $K_i$  values derived from rat Y5-dependent binding of <sup>125</sup>I-PYY

-107-

and peptide ligands (Table 5) were in close range of  $EC_{50}$  values derived from rat Y5-dependent regulation of cAMP accumulation (Table 8). The maximal suppression of cAMP produced by all peptides in Table 6 was between 84% and 120% of that produced by human NPY, except in the case of FLRFamide (42%). Of particular interest is the Y5-selective peptide [D-Trp<sup>32</sup>]NPY. This is a peptide which was shown to stimulate food intake when injected into rat hypothalamus, and which also attenuated NPY-induced feeding in the same paradigm (Balasubramaniam, 1994). Applicants observed that [D-Trp<sup>32</sup>]NPY bound weakly to other Y-type clones with  $K_i > 500$  nM (Tables 4 and 5) and displayed no activity in functional assays (Table 10). In striking contrast, [D-Trp<sup>32</sup>]NPY bound to the rat Y5 receptor with a  $K_i = 53$  nM and was fully able to mimic the inhibitory effect of NPY on forskolin-stimulated cAMP accumulation with an  $EC_{50}$  of 25nM and an  $E_{max} = 72\%$ . That [D-Trp<sup>32</sup>]NPY was able to selectively activate the Y5 receptor while having no detectable activity at the other subtype clones strongly suggests that Y5 receptor activation is responsible for the stimulatory effect of [D-Trp<sup>32</sup>]NPY on feeding behavior in vivo.

**TABLE 8: Functional activation of the rat Y5 receptor.**

Functional data were derived from radioimmunoassay of cAMP accumulation in stably transfected 293 cells stimulated with 10  $\mu$ M forskolin. Peptides were tested for agonist activity at concentrations ranging from 0.03 pM to 0.3  $\mu$ M. The maximum inhibition of cAMP accumulation ( $E_{max}$ ) and the concentration producing a half-maximal effect ( $EC_{50}$ ) were determined by nonlinear regression analysis according to a 4 parameter logistic equation. New peptides are marked with a double asterisk.

TABLE 8

	Peptide	E <sub>max</sub>	EC <sub>50</sub> (nM)
5	rat/human NPY	67 %	1.8
	porcine NPY **		0.79
	rat/human NPY <sub>2-36</sub>	84 %	2.0
	porcine NPY <sub>2-36</sub> **		1.2
	porcine NPY <sub>13-36</sub> **		21
10	rat/human [Leu <sup>31</sup> , Pro <sup>34</sup> ] NPY	70 %	0.6
	porcine [Leu <sup>31</sup> , Pro <sup>34</sup> ] NPY **		1.1
	porcine C2-NPY **		240
	rat/human [D- Trp <sup>32</sup> ] NPY	72 %	9.5
20	rat/porcine PYY	86 %	4.0
	human PYY **		1.5
	human PYY <sub>3-36</sub> **		4.9
	human [Pro <sup>34</sup> ] PYY **		1.8
25	human PP **		1.4
	bovine PP **		5.7

Table 8 continued

-109-

Peptide	$E_{max}$	$EC_{50}$ (nM)
salmon PP **		0.92
rat PP **		130
PYX-1 **		> 300
PYX-2 **		> 300
5 FLRFamide **		13 000

The ability of the human Y5 receptor to inhibit cAMP accumulation was evaluated in NIH-3T3 and LM(tk-) cells, neither of which display an NPY-dependent regulation of [cAMP] without the Y5 construct. Intact cells stably transfected with the human Y5 receptor were analyzed as described above for the rat Y5 cAMP assay. Incubation of stably transfected NIH-3T3 cells with 10 uM forskolin generated an average 21-fold increase in [cAMP] (n = 2). Simultaneous incubation with human NPY decreased the forskolin-stimulated [cAMP] with an  $E_{max}$  of 42% and an  $EC_{50}$  of 8.5 nM (Fig 20). The technique of suspending and then replating the Y5-transfected LM(tk-) cells was correlated with a robust and reliable cellular response to NPY-like peptides and was therefore incorporated into the standard methodology for the functional evaluation of the human Y5 in LM(tk-). Incubation of stably transfected LM(tk-) cells prepared in this manner produced an average 7.4-fold increase in [cAMP] (n = 87). Simultaneous incubation with human NPY decreased the forskolin-stimulated [cAMP] with an  $E_{max}$  of 72% and with an  $EC_{50}$  of 2.4 nM (Fig 21). The human Y5 receptor supported a cellular response to NPY-like peptides in a rank order similar to that described for the rat Y5

-110-

receptor (Table 8, 9). As the rat Y5 receptor is clearly linked by D-Trp32-NPY and other pharmacological tools to the NPY-dependent regulation of feeding behavior, the human Y5 receptor is predicted to function in a similar fashion. Both the human and receptor homologs represent useful models for the screening of compounds intended to modulate feeding behavior by interfering with NPY-dependent pathways.

**TABLE 9: Functional activation of the human Y5 receptor in a cAMP radioimmunoassay.**

Functional data were derived from radioimmunoassay of cAMP accumulation in stably transfected LM(tk-) cells stimulated with 10  $\mu$ M forskolin. Peptides were tested for agonist activity at concentrations ranging from 0.03 pM to 0.3  $\mu$ M. The maximum inhibition of cAMP accumulation ( $E_{max}$ ) and the concentration producing a half-maximal effect ( $EC_{50}$ ) were determined by nonlinear regression analysis according to a 4 parameter logistic equation.

TABLE 9

Peptide	% inhibition relative to human NPY	$EC_{50}$ (nM)
rat/human NPY	100%	2.7
porcine NPY	107%	0.99
rat/human NPY <sub>2-36</sub>	116%	2.6
porcine NPY <sub>2-36</sub>	85%	0.71
porcine NPY <sub>13-36</sub>		49
rat/human [Leu <sup>31</sup> , Pro <sup>34</sup> ] NPY		3.0



Table 9 continued

-111-

Peptide	% inhibition relative to human NPY	EC <sub>50</sub> (nM)
porcine [Leu <sup>31</sup> ,Pro <sup>34</sup> ]NPY		1.3
rat/human [D- Trp <sup>32</sup> ]NPY	108%	26
5 rat/porcine PYY	109%	3.6
human PYY	111%	4.9
human PYY <sub>3-36</sub>		18
human [Pro <sup>34</sup> ]PYY	108%	2.5
human PP	96%	14
10 human PP <sub>2-36</sub>		2.0
human [Ile <sup>31</sup> ,Gln <sup>34</sup> ]PP		5.6
bovine PP		4.0
salmon PP	96%	4.5

TABLE 10: Binding and functional characterization of  
[D-Trp<sup>32</sup>]NPY.

Binding data were generated as described in Tables 4  
and 5. Functional data were derived from  
radioimmunoassay of cAMP accumulation in stably  
transfected cells stimulated with 10  $\mu$ M forskolin. [D-  
Trp<sup>32</sup>]NPY was tested for agonist activity at  
concentrations ranging from 0.03 pM to 0.3  $\mu$ M.  
Alternatively, [D-Trp<sup>32</sup>]NPY was included as a single  
spike (0.3  $\mu$ M) in the human PYY concentration curve for

-112-

human Y1 and human Y2 receptors, or in the human PP concentration curve for human Y4 receptors, and antagonist activity was detected by the presence of a rightward shift (from  $EC_{50}$  to  $EC_{50}'$ ).  $K_b$  values were calculated according to the equation:  $K_b = [D-Trp^{32}]NPY / ((EC_{50}/EC_{50}') - 1)$ . The data shown are representative of at least two independent experiments.

TABLE 10

Recept or Subtyp e	Species	Binding	Function		
		$K_i$ (nM)	$EC_{50}$ (nM)	$K_b$ (nM)	Activity
Y1	Human	> 1000			None detected
Y2	Human	> 1000			None detected
Y4	Human	> 1000			None detected
Y5	Human	18	26		Not Determined
Y1	Rat	> 1000			Not Determined
Y2	Rat	>1000			Not Determined
Y4	Rat	> 1000			Not Determined
Y5	Rat	53	9.50		Agonist

#### Functional Assay: Intracellular Calcium Mobilization

The intracellular free calcium concentration was increased in LM(tk-) cells stably transfected with the human Y5 receptor within 30 seconds of incubation with 100 nM human NPY ( $\Delta Ca^{2+} = 34$ , Fig 21D). Untransfected LM(tk-) cells did not respond to human NPY (data not shown). The calcium mobilization provides a second pathway through which Y5 receptor activation can be measured. These data also serve to link with the Y5

-113-

receptor with other cloned human Y-type receptors, all of which have been demonstrated to mobilize intracellular calcium in various expression systems (Fig 21).

5

#### Localization Studies

The mRNA for the NPY Y5 receptor was widely distributed in rat brain, and appeared to be moderately abundant (Table 11 and Fig. 13). The midline thalamus contained  
10 many neurons with silver grains over them, particularly the paraventricular thalamic nucleus, the rhomboid nucleus, and the nucleus reunions. In addition, moderately intense hybridization signals were observed over neurons in both the centromedial and anterodorsal  
15 thalamic nuclei. In the hypothalamus, a moderate level of hybridization signal was seen over scattered neurons in the lateral hypothalamus, paraventricular, supraoptic, arcuate, and dorsomedial nuclei. In both  
20 the medial preoptic nucleus and suprachiasmatic nucleus, weak or moderate accumulations of silver grains were present. In the suprachiasmatic nucleus, hybridization signal was restricted mainly to the ventrolateral subdivision. In the paraventricular  
25 hypothalamus, positive neurons were observed primarily in the medial parvicellular subdivision.

TABLE 11: Distribution of NPY Y5 mRNA in the Rat CNS

	REGION	Y5 mRNA
	Cerebral cortex	+1
5	Thalamus	
	paraventricular n.	+3
	rhomboid n.	+3
	reunions n.	+3
	anterodorsal n.	+2
10	Hypothalamus	
	paraventricular n.	+2
	lateral hypoth. area	+2 /+3
	supraoptic n.	+1
	medial preoptic n.	+2
15	suprachiasmatic n.	+1/+2
	arcuate n.	+2
	Hippocampus	
	dentate gyrus	+1
	polymorph dentate gyrus	+2
20	CA1	0
	CA3	+1
	Amygdala	
	central amygd. n., medial	+2
	anterior cortical amygd. n.	+2
25	Olivary pretectal n.	+3
	Anterior pretectal n.	+3
	Substantia nigra, pars compacta	+2
	Superior colliculus	+2
	Central gray	+2
30	Rostral linear raphe	+3
	Dorsal raphe	+1
	Inferior colliculus	+1
	Medial vestibular n.	+2/+3
	Parvicellular ret. n., alpha	+2
35	Gigantocellular reticular n., alpha	+2
	Pontine nuclei	+1/+2

-115-

Moderate hybridization signals were found over most of the neurons in the polymorphic region of the dentate gyrus in the hippocampus, while lower levels were seen over scattered neurons in the CA3 region. In the amygdala, the central nucleus and the anterior cortical nucleus contained neurons with moderate levels of hybridization signal. In the mesencephalon, hybridization signals were observed over a number of areas. The most intense signals were found over neurons in the anterior and olivary pretectal nuclei, periaqueductal gray, and over the rostral linear raphe. Moderate hybridization signals were observed over neurons in the internal gray layer of the superior colliculus, the substantia nigra, pars compacta, the dorsal raphe, and the pontine nuclei. Most of the neurons in the inferior colliculus exhibited a low level of signal. In the medulla and pons, few areas exhibited substantial hybridization signals. The medial vestibular nucleus was moderately labeled, as was the parvocellular reticular nucleus, pars alpha, and the gigantocellular reticular nucleus.

Little or no hybridization signal was observed on sections hybridized with the radiolabeled sense oligonucleotide probe. More importantly, in the transfected COS-7 cells, the antisense probe hybridized only to the cells transfected with the rat Y5 cDNA (Table 12). These results indicate that the probe used to characterize the distribution of Y5 mRNA in rat brain is specific for this mRNA, and does not cross-hybridize to any of the other known NPY receptor mRNAs.

-116-

**TABLE 12: Hybridization of antisense oligonucleotide probes to transfected COS-7 cells.**

Hybridization was performed as described in Methods. The NPY Y5 probe hybridizes only to the cells transfected with the Y5 cDNA. ND=not done.

<u>Cells</u>	Mock	rY1	rY2	rY4	rY5
<u>Oligo</u>					
rY1	-	+	-	ND	ND
rY2	-	-	+	-	-
rY4	-	-	-	+	-
rY5	-	-	-	-	+

**In vivo studies with Y5-selective compounds**

The results reported above strongly support a role for the Y5 receptor in regulating feeding behavior. Accordingly, applicants have synthesized and evaluated the binding and functional properties of several compounds at the cloned human Y1, human Y2, human Y4, and human Y5 receptors. As shown below in Table 13, applicants have discovered several compounds which not only bind selectively to the human Y5 receptor but also act as Y5 receptor antagonists, as measured by their ability to block NPY-induced inhibition of cAMP accumulation in forskolin-stimulated LM(tk-) cells stably transfected with the cloned human Y5 receptor. An example of such a compound is shown in Figure 22. Preliminary experiments indicate that compound 28 is a Y5 receptor antagonist.

**Table 13: Evaluation of human Y5 receptor antagonists**

The ability of the compounds to antagonize the Y-type receptors is reported as the  $K_b$ . The  $K_b$  is derived from

-117-

the  $EC_{50}$ , or concentration of half-maximal effect, in the presence ( $EC_{50}$ ) or absence ( $EC_{50}'$ ) of compound, according to the equation:  $K_b = [NPY] / ((EC_{50}/EC_{50}') - 1)$ . Results shown are representative of at least three independent experiments.

5 N.D. = Not determined.

Table 13

	Binding Affinity (K <sub>i</sub> (nM) vs. <sup>125</sup> I-PYY)					
10	Compound	Human Receptor			K <sub>b</sub> (nM)	
	-	Y1	Y2	Y4	Y5	-
15	1	1660	1920	4540	38.9	183
	2	1806	386	1280	17.8	9.6
	5	3860	249	2290	1.27	2.1
	6	4360	4610	32,900	47.5	93
	7	2170	2870	7050	42.0	105
	9	3240	>100,000	3720	108	479
	10	1070	>100,000	5830	40.7	2.8
	11	1180	>100,000	7130	9.66	1.5
20	17	5550	1000	8020	14	6.0
	19	3550	955	11700	11	23
	20	16000	7760	20400	8.3	26
	21	13000	1610	18500	9.8	16
	22	17200	7570	27500	11	3.0
25	23	14500	617	21500	26	38
	25	3240	851	13100	17	311
	26	23700	58200	19300	14	50

Table 13 continued

-118-

	Binding Affinity (K <sub>i</sub> (nM) vs. <sup>125</sup> I-PYY)				
27	48700	5280	63100	28	49
28	>100,000	>75,000	>100,000	19,000	N.D.

5        These compounds were further tested using in vivo  
animal models of feeding behavior. Since NPY is the  
strongest known stimulant of feeding behavior,  
experiments were performed with several compounds to  
evaluate the effect of the compounds described above on  
10       NPY-induced feeding behavior in satiated rats.

First, 300 pmole of porcine NPY in vehicle (A.C.S.F.)  
was administered by intracerebroventricular (i.c.v.)  
injection, along with i.p. administration of compound  
15       vehicle (10% DMSO/water), and the food intake of NPY-  
stimulated animals was compared to food intake in  
animals treated with the vehicles. The 300 pmole  
injection of NPY was found to significantly induce food  
intake ( $p < 0.05$ ; Student-Newman-Keuls).

20       Using the 300 pmole dose of NPY found to be effective  
to stimulate feeding, other animals were treated with  
the compounds by intraperitoneal (i.p.) administration,  
followed 30-60 minutes later by i.c.v. NPY  
25       administration, and measurement of subsequent food  
intake. As shown in Table 14, NPY-induced food intake  
was significantly reduced in animals first treated with  
the compounds ( $p < 0.05$ ; Student-Newman-Keuls). These  
experiments demonstrate that NPY-induced food intake is  
30       significantly reduced by administration to animals of  
a compound which is a Y5-selective antagonist.

Table 14. NPY-induced cumulative food intake in rats  
treated with either the i.c.v. and i.p. vehicles



-119-

(control), 300 pmole NPY alone (NPY), or in rats treated first with compound and then NPY (NPY + compound). Food intake was measured 4 hours after stimulation with NPY. Food intake is reported as the mean  $\pm$  S.E.M. intake for a group of animals.

Table 14

	Food intake (g) mean $\pm$ S.E.M.			
	1	5	17	19
Compound				
Compound Dose (mg/kg i.p.)	10	10	10	30
control (vehicles only)	3.7 $\pm$ 0.6	2.4 $\pm$ 0.5	2.4 $\pm$ 0.7	2.9 $\pm$ 0.8
NPY	7.4 $\pm$ 0.5	6.8 $\pm$ 1.0	5.8 $\pm$ 0.5	4.9 $\pm$ 0.4
NPY + compound	4.6 $\pm$ 0.6	4.1 $\pm$ 0.4	3.8 $\pm$ 0.4	1.5 $\pm$ 0.6

Since food deprivation induces an increase in the hypothalamic NPY levels, it has been postulated that food intake following a period of food deprivation is NPY-mediated. Therefore, the Y5 antagonists of Table 13 were administered to conscious rats following a 24h food deprivation. Each of the human Y5 receptor antagonists shown in Table 13 was able to significantly reduce NPY-induced food intake in the animals, as shown below in Table 15. The food intake of animals treated with test compound is reported as a percentage of the food intake measured for control animals (treated with vehicle), i.e., 25% means the animals treated with the compound consumed only 25% as much food as the control animals. Measurements were performed two hours after administration of the test

-120-

compound.

**Table 15 Two-hour food intake of NPY-stimulated rats.**  
Food intake is expressed as the percentage  
of intake compared to control rats.

Compound	Mean (%)	Compound	Mean (%)
1	34	19	36
2	42	20	35
5	87	21	80
6	38	22	55
7	47	23	58
9	40	25	32
10	74	26	73
11	15	27	84
17	27	28	N.D.

These experiments indicate that the compounds of the present invention inhibit food intake in rats, especially when administered in a range of about 0.01 to about 100 mg/kg rat, by either oral, intraperitoneal or intravenous administration. The animals appeared normal during these experiments, and no ill effects on the animals were observed after the termination of the feeding experiments.

The binding properties of the compounds were also

-121-

evaluated with respect to other cloned human G-protein coupled receptors. As shown in Table 16, below, the Y5-selective compounds described hereinabove exhibited lower affinity for receptors other than the Y-type  
5 receptors.

-122-

Table 16 Cross-reactivity of compounds at other cloned human receptors

Compound	Receptor (pKi)									
	$\alpha_{1d}$	$\alpha_{1b}$	$\alpha_{1a}$	$\alpha_{2a}$	$\alpha_{2b}$	$\alpha_{2c}$	H1	H2	D3	
1	6.25	6.23	6.15	6.28	6.01	6.34	5.59	6.32	5.69	
2	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	
5	7.24	7.36	7.63	7.39	7.29	7.63	6.65	6.68	7.24	
6	5.68	5.73	6.54	7.14	5.79	6.35	N.D.	N.D.	N.D.	
7	6.46	6.08	6.06	7.16	6.09	6.85	N.D.	N.D.	N.D.	
9	6.45	6.26	6.57	7.04	5.00	6.81	N.D.	N.D.	N.D.	
10	6.12	5.82	6.27	8.94	5.62	6.18	N.D.	N.D.	N.D.	
11	7.03	5.6	6.05	7.38	5.60	6.00	N.D.	N.D.	N.D.	
17	6.68	7.17	7.08	6.52	6.51	7.07	6.33	5.92	6.61	
19	6.90	7.35	7.47	6.74	6.58	7.07	7.04	6.29	6.69	
20	7.01	7.22	7.72	7.31	6.96	7.39	6.73	5.85	6.35	
21	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	
22	6.80	6.98	7.34	7.05	6.43	7.15	6.22	5.72	6.29	
23	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	N.D.	
25	6.66	6.67	7.07	6.21	5.95	6.79	6.43	6.43	5.93	







-126-

**EXPERIMENTAL DISCUSSION**

In order to isolate new NPY receptor subtypes applicants choose an expression cloning approach where a functional receptor is actually detected with  
5 exquisite sensitivity on the surface of transfected cells, using a highly specific iodinated ligand. Using this strategy, applicants have identified a rat hypothalamic cDNA encoding a novel Y-type receptor (Y5). The fact that applicants had to screen  $3.5 \times 10^6$   
10 independent clones with a 2.7 kb average insert size to find two clones reveals either a very strong bias against Y5 cDNA cloning in the cDNA library construction procedure or that the Y5 mRNA is expressed at very low levels in rat hypothalamic tissue. The  
15 longest reading frame in the rat Y5 cDNA (CG-18) encodes a 456 amino acid protein with an estimated molecular weight of 50.1 kD. Given there are two N-linked glycosylation site in the amino terminus, the apparent molecular weight could be slightly higher. Applicants have isolated the human Y5 homolog from a  
20 human hippocampal cDNA library. The longest reading frame in the human Y5 cDNA (CG-19) encodes a 455 amino acid protein with an estimated molecular weight of 50 kD. The human Y5 receptor is one amino acid shorter  
25 than the rat Y5 and shows significant amino acid differences both in the N-terminal and the middle of the third intracellular loop portions of the protein. The seven transmembrane domains and the extracellular loops, however, are virtually identical and the protein motifs found in both species homologs are identical.  
30 Both human and rat Y5 receptors carry a large number of potential phosphorylation sites in their second and third intra- cellular loops which could be involved in the regulation of their functional characteristics.

35

The rat and human Y5 receptors both carry a leucine zipper in the first putative transmembrane domain. In



-127-

such a structure, it has been proposed that segments containing periodic arrays of leucine residues exist in an alpha-helical conformation. The leucine side chains extending from one alpha-helix interact with those from a similar alpha helix of a second polypeptide, facilitating dimerization by the formation of a coiled coil (O'Shea et al, 1989). Usually, such patterns are associated with nuclear DNA binding protein like c-myc, c-fos and c-jun, but it is possible that in some proteins the leucine repeat simply facilitates dimerization and has little to do with positioning a DNA-binding region. Further evidence supporting the idea that dimerization of specific seven transmembrane receptors can occur comes from coexpression studies with muscarinic/adrenergic receptors where intermolecular "cross-talk" between chimeric G-protein coupled receptors has been described (Maggio et al., 1993). The tyrosine phosphorylation site found in the middle of this leucine zipper in transmembrane domain one (TM I) could be involved in regulating dimerization of the Y5 receptor. The physiological significance of G-protein coupled receptor dimerization remains to be elucidated but by analogy with peptide hormone receptors oligomerization, it could be involved in receptor activation and signal transduction (Wells, 1994).

The nucleotide and amino acid sequence analysis of Y5 (rat and human) reveals low identity levels with all 7 TM receptors including the Y1, Y2 and Y4 receptors, even in the transmembrane domains which are usually highly conserved within receptor subfamilies. Applicants have named CG-18 and CG-19 "Y5" receptors because of their unique amino acid sequence (87.2% identical with each other,  $\leq$  42% identical with the TM regions of previously cloned "Y" receptor subtypes) and pharmacological profile. The name is not biased toward

-128-

any one member of the pancreatic polypeptide family. The "Y" has its roots in the original classification of Y1 and Y2 receptor subtypes (Wahlestedt et al., 1987). The letter reflects the conservation in pancreatic polypeptide family members of the C-terminal tyrosine, described as "Y" in the single letter amino acid code. The number is the next available in the Y-type series, position number three having been reserved for the pharmacologically defined Y3 receptor. Applicants note that the cloned human Y1 receptor was introduced by Larhammar and co-workers as a "human neuropeptide Y/peptide YY receptor of the Y1 type" (Larhammar et al., 1992). Similarly, the novel clones described herein can be described as rat and human neuropeptide Y/peptide YY receptors of the Y5 type.

The rat hypothalamic Y5 receptor displays a very similar pharmacological profile to the pharmacologically described "atypical" Y1 receptor thought to mediate NPY-induced food intake in rat hypothalamus. Both the Y5 receptor and the "feeding receptor" display a preference for NPY and PYY-like analogs, a sensitivity to N-terminal peptide deletion, and a tolerance for Pro<sup>34</sup>. Each would be considered Y1-like except for the anomalous ability of NPY<sub>2-36</sub> to bind and activate as well as NPY. Each appears to be sensitive to changes in the mid-region of the peptide ligand. For example, a study by Kalra and colleagues (1991) indicated that replacement of the NPY midregion by an amino-octanoic chain to produce NPY<sub>1-4</sub>-Aca-<sub>25-36</sub> dramatically reduced activity in a feeding behavioral assay. Likewise, applicants note that the robust difference in human PP binding ( $K_i = 5.0$  nM) and rat PP binding ( $K_i = 230$ ) to the rat Y5 receptor can be attributed to a series of 8 amino acid changes between residues 6-30 in the peptide ligands, with human PP bearing the closer resemblance to human NPY. Note also

-129-

that FLRFamide, a structural analog of the FMRFamide peptide which is reported to stimulate feeding in rats, was able to bind and activate the rat Y5 receptor albeit at relatively high concentrations (Orosco, et al., 1989). These matching profiles, combined with a selective activation of the rat Y5 by the reported feeding "modulator" [D-Trp<sup>32</sup>]NPY, support the identity of the rat Y5 as the "feeding receptor" first proposed to explain NPY-induced feeding in rat hypothalamus.

That the human Y5 receptor has a pharmacological profile like that of the rat Y5 in both binding and functional assays suggests that the two receptors may have similar functions in vivo.

The distribution of Y5 mRNA in rat brain further extends the argument for a role of Y5 receptors in feeding behavior. The anatomical locus of the feeding response, for example, has been suggested to reside at least in part in the paraventricular hypothalamic nucleus (PVN) and also in the lateral hypothalamus, two places where Y5 mRNA was detected in abundance. Post-synaptic localization of the Y5 receptor in both of these regions can regulate the response to endogenously released NPY in vivo. The paraventricular nucleus receives projections from NPY-containing neurons in the arcuate nucleus, another region where Y5 mRNA was detected. This indicates a pre-synaptic role for the Y5 receptor in the control of NPY release via the arcuato-paraventricular projection, and consequently in the control of feeding behavior. The localization of the Y5 mRNA in the midline thalamic nuclei is also important. The paraventricular thalamic nucleus/centromedial nucleus complex projects heavily to the paraventricular hypothalamus and to the amygdala. As such, the Y5 receptor is a substrate for the emotional aspect of appetitive behaviors.

-130-

Y5 receptors are highly attractive targets for appetite and weight control based on several lines of research (Sahu and Kalra, 1993). NPY is the most potent stimulant of feeding behavior yet described (Clark et al., 1984; Levine and Morley, 1984; Stanley and Leibowitz, 1984). Direct injection of NPY into the hypothalamus of rats can increase food intake ~ 10-fold over a 4-hour period (Stanley et al., 1992). NPY-stimulated rats display a preference for carbohydrates over protein and fat (Stanley et al., 1985). Interestingly, NPY and NPY mRNA are increased in food-deprived rats (Brady et al., 1990; O' Shea and Gundlach, 1991) and also in rats which are genetically obese (Sanacora et al., 1990) or made diabetic by treatment with streptozotocin (White et al., 1990). One potential explanation is that NPY, a potent stimulant of feeding behavior in normal rats, is disregulated in the overweight or diabetic animal so that food intake is increased, accompanied by obesity. The physiological stress of obesity increases the risk for health problems such as cardiovascular malfunction, osteoarthritis, and hyperinsulinemia, together with a worsened prognosis for adult-onset diabetes. A nonpeptide antagonist targeted to the Y5 receptor could therefore be effective as a way to control not only appetite and body weight but an entire range of obesity- and diabetes-related disorders (Dryden et al., 1994). There is also neurochemical evidence to suggest that NPY-mediated functions are disregulated in eating disorders such as bulimia and anorexia nervosa, so that they too could be responsive to treatment by a Y5-selective drug. It has been proposed, for example, that food intake in NPY-stimulated rats mimics the massive food consumption associated with binge eating in bulimia (Stanley, 1993). CSF levels of PYY but not NPY were elevated in bulimic patients who abstained from binging, and then diminished when binging was allowed

-131-

(Berrettini et al., 1988). Conversely, NPY levels were elevated in underweight anorectic patients and then diminished as body weight was normalized (Kaye et al., 1990).

5

As described above, the human and rat *in vitro* expression models were used in combination to screen for compounds intended to modulate NPY-dependent feeding behavior. Using this approach, applicants have discovered several compounds which inhibit feeding behavior in animal models, which should lead to additional drug discoveries. The compounds according to the present invention inhibit food intake in Zucker obese rats in a range especially of about 0.01 to about 100 mg/kg after oral, intraperitoneal or intravenous administration.

10  
15

The Y5 pharmacological profile further offers a new standard by which to review the molecular basis of all NPY-dependent processes; examples are listed in Table 11. Such an exercise suggests that the Y5 receptor is likely to have a physiological significance beyond feeding behavior. It has been reported, for example, that a Y-type receptor can regulate luteinizing hormone releasing hormone (LHRH) release from the median eminence of steroid-primed rats *in vitro* with an atypical Y1 pharmacological profile. NPY, NPY<sub>2-36</sub>, and LP-NPY were all effective at 1uM but deletion of as few as four amino acids from the N-terminus of NPY destroyed biological activity. The Y5 may therefore represent a therapeutic target for sexual or reproductive disorders. Preliminary *in situ* hybridization of rat Y5 mRNA in hippocampus and elsewhere further suggest that additional roles will be uncovered, for example, in the regulation of memory. It is worth while considering that the Y5 is so similar

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-132-

in pharmacological profile to the other Y-type receptors that it may have been overlooked among a mixed population of Y1, Y2 and Y4 receptors. Certain functions now associated with these subtypes could therefore be reassigned to Y5 as our pharmacological tools grow more sophisticated (Table 18). By offering new insight into NPY receptor pharmacology, the Y5 thereby provides a greater clarity and focus in the field of drug design.

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-133-

**TABLE 17: Pathophysiological Conditions Associated With NPY**

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The following pathological conditions have been linked to either 1) application of exogenous NPY, or 2) changes in levels of endogenous NPY.

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1	obesity	Sahu and Kalra, 1993
2	eating disorders (anorexia and bulimia nervosa)	Stanley, 1993
3	sexual/reproduct ive function	Clark, 1994
4	depression	Heilig and Weiderlov, 1990
5	anxiety	Wahlestedt et al., 1993
6	cocaine addiction	Wahlestedt et al., 1991
7	gastric ulcer	Penner et al., 1993
8	memory loss	Morley and Flood, 1990
9	pain	Hua et al., 1991
10	epileptic seizure	Rizzi et al., 1993
11	hypertension	Zukowska-Grojec et al., 1993
12	subarachnoid hemorrhage	Abel et al., 1988
13	shock	Hauser et al., 1993
14	circadian rhythm	Albers and Ferris, 1984
15	nasal congestion	Lacroix et al., 1988
16	diarrhea	Cox and Cuthbert, 1990
17	neurogenic voiding dysfunction	Zoubek et al., 1993

-134-

A successful strategy for the design of a Y5-receptor based drug or for any drug targeted to single G protein-coupled receptor subtype involves the screening of candidate compounds 1) in radioligand binding assays so as to detect affinity for cross-reactive G protein-coupled receptors, and 2) in physiological assays so as to detect undesirable side effects. In the specific process of screening for a Y5-selective drug, the receptor subtypes most likely to cross-react and therefore most important for radioligand binding screens include the other "Y-type" receptors, Y1, Y2, Y3, and Y4. Cross-reactivity between the Y5 and any of the other subtypes could result in potential complications as suggested by the pathophysiological indications listed in Table 17. In designing a Y5 antagonist for obesity and appetite control, for example, it is important not to design a Y1 antagonist resulting in hypertension or increased anxiety, a Y2 antagonist resulting in memory loss, or a Y4 antagonist resulting in increased appetite.



**TABLE 18: Y-Type Receptor Indications**

5	Y-type Receptor Indications	Receptor Subtype	Drug Activity	Reference
	obesity, appetite disorder	atypical Y1	antagonist	Sahu and Kalra, 1993
10	adult onset diabetes	atypical Y1	antagonist	Sahu and Kalra, 1993
	bulimia nervosa	atypical Y1	antagonist	Stanley, 1993
15	pheochromoc ytoma- induced hypertensio n	Y1	antagonist	Grouzman et al., 1989
20	subarachnoi d hemorrhage	Y1	antagonist	Abel et al., 1988
	neurogenic vascular hypertrophy	Y1 Y2	antagonist antagonist	Zukowska- Grojec et al., 1993
25	epileptic seizure	Y2	antagonist	Rizzi et al., 1993
30	hypertensio n: central, peripheral regulation	peripheral Y1 central Y3 central Y2	antagonist agonist antagonist	Grundemar and Hakanson, 1993 Barraco et al., 1991
	obesity, appetite disorder	Y4 or PP	agonist	Malaisse- Lagae et al., 1977
35	anorexia nervosa	atypical Y1	agonist	Berrettin i et al., 1988
	anxiety	Y1	agonist	Wahlested t et al., 1993

Table 18 continued

-136-

	cocaine addiction	Y1	agonist	Wahlestedt et al., 1991
5	stress-induced gastric ulcer	Y1 Y4 or PP	agonist agonist	Penner et al., 1993
	memory loss	Y2	agonist	Morley and Flood, 1990
	pain	Y2	agonist	Hua et al., 1991
	shock	Y1	agonist	Hauser et al., 1993
10	sleep disturbances, jet lag	Y2	not clear	Albers and Ferris, 1984
15	nasal decongestion	Y1 Y2	agonist agonist	Lacroix et al., 1988
	diarrhea	Y2	agonist	Cox and Cuthbert, 1990

-137-

The cloning of the Y5 receptor from human and rat is especially valuable for receptor characterization based on *in situ* localization, anti-sense functional knock-out, and gene induction. These studies will generate  
5 important information related to Y5 receptor function and its therapeutic significance. The cloned Y5 receptor lends itself to mutagenesis studies in which receptor/ligand interactions can be modeled. The Y5 receptor further allows us to investigate the  
10 possibility of other Y-type receptors through homology cloning. These could include new receptor subtypes as well as Y5 species homologs for the establishment of experimental animal models with relevance for human pathology. The Y5 receptor therefore represents an  
15 enormous opportunity for the development of novel and selective drug therapies, particularly those targeted to appetite and weight control, but also for memory loss, depression, anxiety, gastric ulcer, epileptic seizure, pain, hypertension, subarachnoid hemorrhage,  
20 sleeping disturbances, nasal congestion, neurogenic voiding dysfunction, and diarrhea.

In particular, the discovery of Y5-selective antagonists which inhibit food intake in rats provides a method of  
25 modifying feeding behavior in a wide variety of vertebrate animals.

-138-

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## SEQUENCE LISTING

## (1) GENERAL INFORMATION:

5

(i) APPLICANT: Synaptic Pharmaceutical Corporation

10

(ii) TITLE OF INVENTION: METHODS OF MODIFYING FEEDING  
BEHAVIOR, COMPOUNDS USEFUL IN SUCH  
METHODS, AND DNA ENCODING A  
HYPOTHALAMIC ATYPICAL NEUROPEPTIDE  
Y/PEPTIDE YY RECEPTOR (Y5) AND USES  
THEREOF

15

(iii) NUMBER OF SEQUENCES: 12

(iv) CORRESPONDENCE ADDRESS:

20

(A) ADDRESSEE: Cooper & Dunham LLP  
(B) STREET: 1185 Avenue of the Americas  
(C) CITY: New York  
(D) STATE: New York  
(E) COUNTRY: United States of America  
(F) ZIP: 10036

25

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk  
(B) COMPUTER: IBM PC compatible  
(C) OPERATING SYSTEM: PC-DOS/MS-DOS  
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25

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(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:  
(B) FILING DATE:  
(C) CLASSIFICATION:

35

(viii) ATTORNEY/AGENT INFORMATION:

(A) NAME: White, John P.  
(B) REGISTRATION NUMBER: 28,678  
(C) REFERENCE/DOCKET NUMBER: 1795/46166-A-PCT

40

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: (212) 278-0400  
(B) TELEFAX: (212) 391-0525

45

## (2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

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(A) LENGTH: 1501 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

55

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

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(ix) FEATURE:

(A) NAME/KEY: CDS  
(B) LOCATION: 61..1432

65

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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70

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	Met	Asp	Val	Leu	Phe	Phe	His	Gln	Asp	Ser	Ser	Met	Glu	Phe	Lys	Leu	
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	Glu	Glu	His	Phe	Asn	Lys	Thr	Phe	Val	Thr	Glu	Asn	Asn	Thr	Ala	Ala	
10				20					25					30			
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	300																
	Leu	Gly	Phe	Met	Gly	Asn	Leu	Leu	Ile	Leu	Met	Ala	Val	Met	Lys	Lys	
25		65				70					75				80		
	CGC	AAT	CAG	AAG	ACT	ACA	GTG	AAC	TTT	CTC	ATA	GGC	AAC	CTG	GCC	TTC	
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	Val	Leu	Leu	Asp	Gln	Trp	Met	Phe	Gly	Lys	Ala	Met	Cys	His	Ile	Met	
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-151-

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 225 230 235 240  
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 AGT CAT ACC AGC GTC TGC CGA AGC ATA AGC TGT GGA TTG TCC CAC AAA  
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 245 250 255  
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 GAA AAC AGA CTC GAA GAA AAT GAG ATG ATC AAC TTA ACC CTA CAG CCA  
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 260 265 270  
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 TCC AAA AAG AGC AGG AAC CAG GCA AAA ACC CCC AGC ACT CAA AAG TGG  
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 Ser Lys Lys Ser Arg Asn Gln Ala Lys Thr Pro Ser Thr Gln Lys Trp  
 275 280 285  
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 Ala Val Pro Glu Asn Pro Ala Ser Val Arg Ser Gln Leu Ser Pro Ser  
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 1116  
 Ser Lys Val Ile Pro Gly Val Pro Ile Cys Phe Glu Val Lys Pro Glu  
 340 345 350  
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 355 360 365  
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 Ile Lys Lys Arg Ser Arg Ser Val Phe Tyr Arg Leu Thr Ile Leu Ile  
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 ACT GAC TTC AAT GAT AAC TTG ATT TCC AAT AGG CAT TTC AAG CTG GTA  
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 Tyr Cys Ile Cys His Leu Leu Gly Met Met Ser Cys Cys Leu Asn Pro  
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-152-

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1452

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10 (2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 457 amino acids

15 (B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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Ala Arg Asn Ala Ala Phe Pro Ala Trp Glu Asp Tyr Arg Gly Ser Val  
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Asp Asp Leu Gln Tyr Phe Leu Ile Gly Leu Tyr Thr Phe Val Ser Leu  
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Leu Gly Phe Met Gly Asn Leu Leu Ile Leu Met Ala Val Met Lys Lys  
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Arg Asn Gln Lys Thr Thr Val Asn Phe Leu Ile Gly Asn Leu Ala Phe  
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Ser Asp Ile Leu Val Val Leu Phe Cys Ser Pro Phe Thr Leu Thr Ser  
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-153-

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 30 Ile Leu Tyr Gly Phe Leu Asn Asn Gly Ile Lys Ala Asp Leu Arg Ala  
 435 440 445  
 35 Leu Ile His Cys Leu His Met Ser \*  
 450 455

## (2) INFORMATION FOR SEQ ID NO:3:

40 (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 1457 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear  
 45 (ii) MOLECULE TYPE: cDNA  
 (iii) HYPOTHETICAL: NO  
 50 (iv) ANTI-SENSE: NO  
 (ix) FEATURE:  
 (A) NAME/KEY: CDS  
 (B) LOCATION: 61..1432  
 55 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:  
 60 GTTCCCTCT GAATAGATTA ATTTAAAGTA GTCATGTAAT GTTTTTTTGG TTGCTGACAA  
 60  
 ATG TCT TTT TAT TCC AAG CAG GAC TAT AAT ATG GAT TTA GAG CTC GAC  
 108  
 Met Ser Phe Tyr Ser Lys Gln Asp Tyr Asn Met Asp Leu Glu Leu Asp  
 65 1 5 10 15  
 GAG TAT TAT AAC AAG ACA CTT GCC ACA GAG AAT AAT ACT GCT GCC ACT  
 156  
 Glu Tyr Tyr Asn Lys Thr Leu Ala Thr Glu Asn Asn Thr Ala Ala Thr  
 70 20 25 30

-154-

CGG AAT TCT GAT TTC CCA GTC TGG GAT GAC TAT AAA AGC AGT GTA GAT  
 204  
 Arg Asn Ser Asp Phe Pro Val Trp Asp Asp Tyr Lys Ser Ser Val Asp  
 35 40 45  
 5  
 GAC TTA CAG TAT TTT CTG ATT GGG CTC TAT ACA TTT GTA AGT CTT CTT  
 252  
 Asp Leu Gln Tyr Phe Leu Ile Gly Leu Tyr Thr Phe Val Ser Leu Leu  
 50 55 60  
 10  
 GGC TTT ATG GGG AAT CTA CTT ATT TTA ATG GCT CTC ATG AAA AAG CGT  
 300  
 Gly Phe Met Gly Asn Leu Leu Ile Leu Met Ala Leu Met Lys Lys Arg  
 65 70 75 80  
 15  
 AAT CAG AAG ACT ACG GTA AAC TTC CTC ATA GGC AAT CTG GCC TTT TCT  
 348  
 Asn Gln Lys Thr Thr Val Asn Phe Leu Ile Gly Asn Leu Ala Phe Ser  
 85 90 95  
 20  
 GAT ATC TTG GTT GTG CTG TTT TGC TCA CCT TTC ACA CTG ACG TCT GTC  
 396  
 Asp Ile Leu Val Val Leu Phe Cys Ser Pro Phe Thr Leu Thr Ser Val  
 100 105 110  
 25  
 TTG CTG GAT CAG TGG ATG TTT GGC AAA GTC ATG TGC CAT ATT ATG CCT  
 444  
 Leu Leu Asp Gln Trp Met Phe Gly Lys Val Met Cys His Ile Met Pro  
 115 120 125  
 30  
 TTT CTT CAA TGT GTG TCA GTT TTG GTT TCA ACT TTA ATT TTA ATA TCA  
 492  
 Phe Leu Gln Cys Val Ser Val Leu Val Ser Thr Leu Ile Leu Ile Ser  
 130 135 140  
 35  
 ATT GCC ATT GTC AGG TAT CAT ATG ATA AAA CAT CCC ATA TCT AAT AAT  
 540  
 Ile Ala Ile Val Arg Tyr His Met Ile Lys His Pro Ile Ser Asn Asn  
 145 150 155 160  
 40  
 TTA ACA GCA AAC CAT GGC TAC TTT CTG ATA GCT ACT GTC TGG ACA CTA  
 588  
 Leu Thr Ala Asn His Gly Tyr Phe Leu Ile Ala Thr Val Trp Thr Leu  
 165 170 175  
 45  
 GGT TTT GCC ATC TGT TCT CCC CTT CCA GTG TTT CAC AGT CTT GTG GAA  
 636  
 Gly Phe Ala Ile Cys Ser Pro Leu Pro Val Phe His Ser Leu Val Glu  
 180 185 190  
 50  
 CTT CAA GAA ACA TTT GGT TCA GCA TTG CTG AGC AGC AGG TAT TTA TGT  
 684  
 Leu Gln Glu Thr Phe Gly Ser Ala Leu Leu Ser Ser Arg Tyr Leu Cys  
 195 200 205  
 55  
 GTT GAG TCA TGG CCA TCT GAT TCA TAC AGA ATT GCC TTT ACT ATC TCT  
 732  
 Val Glu Ser Trp Pro Ser Asp Ser Tyr Arg Ile Ala Phe Thr Ile Ser  
 210 215 220  
 60  
 TTA TTG CTA GTT CAG TAT ATT CTG CCC TTA GTT TGT CTT ACT GTA AGT  
 780  
 Leu Leu Leu Val Gln Tyr Ile Leu Pro Leu Val Cys Leu Thr Val Ser  
 225 230 235 240  
 65  
 CAT ACA AGT GTC TGC AGA AGT ATA AGC TGT GGA TTG TCC AAC AAA GAA  
 828  
 His Thr Ser Val Cys Arg Ser Ile Ser Cys Gly Leu Ser Asn Lys Glu  
 245 250 255  
 70

-155-

AAC AGA CTT GAA GAA AAT GAG ATG ATC AAC TTA ACT CTT CAT CCA TCC  
 876  
 Asn Arg Leu Glu Glu Asn Glu Met Ile Asn Leu Thr Leu His Pro Ser  
 260 265 270  
 5  
 AAA AAG AGT GGG CCT CAG GTG AAA CTC TCT GGC AGC CAT AAA TGG AGT  
 924  
 Lys Lys Ser Gly Pro Gln Val Lys Leu Ser Gly Ser His Lys Trp Ser  
 275 280 285  
 10  
 TAT TCA TTC ATC AAA AAA CAC AGA AGA AGA TAT AGC AAG AAG ACA GCA  
 972  
 Tyr Ser Phe Ile Lys Lys His Arg Arg Arg Tyr Ser Lys Lys Thr Ala  
 290 295 300  
 15  
 TGT GTG TTA CCT GCT CCA GAA AGA CCT TCT CAA GAG AAC CAC TCC AGA  
 1020  
 Cys Val Leu Pro Ala Pro Glu Arg Pro Ser Gln Glu Asn His Ser Arg  
 305 310 315 320  
 20  
 ATA CTT CCA GAA AAC TTT GGC TCT GTA AGA AGT CAG CTC TCT TCA TCC  
 1068  
 Ile Leu Pro Glu Asn Phe Gly Ser Val Arg Ser Gln Leu Ser Ser Ser  
 325 330 335  
 25  
 AGT AAG TTC ATA CCA GGG GTC CCC ACT TGC TTT GAG ATA AAA CCT GAA  
 1116  
 Ser Lys Phe Ile Pro Gly Val Pro Thr Cys Phe Glu Ile Lys Pro Glu  
 340 345 350  
 30  
 GAA AAT TCA GAT GTT CAT GAA TTG AGA GTA AAA CGT TCT GTT ACA AGA  
 1164  
 Glu Asn Ser Asp Val His Glu Leu Arg Val Lys Arg Ser Val Thr Arg  
 355 360 365  
 35  
 ATA AAA AAG AGA TCT CGA AGT GTT TTC TAC AGA CTG ACC ATA CTG ATA  
 1212  
 Ile Lys Lys Arg Ser Arg Ser Val Phe Tyr Arg Leu Thr Ile Leu Ile  
 370 375 380  
 40  
 TTA GTA TTT GCT GTT AGT TGG ATG CCA CTA CAC CTT TTC CAT GTG GTA  
 1260  
 Leu Val Phe Ala Val Ser Trp Met Pro Leu His Leu Phe His Val Val  
 385 390 395 400  
 45  
 ACT GAT TTT AAT GAC AAT CTT ATT TCA AAT AGG CAT TTC AAG TTG GTG  
 1308  
 Thr Asp Phe Asn Asp Asn Leu Ile Ser Asn Arg His Phe Lys Leu Val  
 405 410 415  
 50  
 TAT TGC ATT TGT CAT TTG TTG GGC ATG ATG TCC TGT TGT CTT AAT CCA  
 1356  
 Tyr Cys Ile Cys His Leu Leu Gly Met Met Ser Cys Cys Leu Asn Pro  
 420 425 430  
 55  
 ATT CTA TAT GGG TTT CTT AAT AAT GGG ATT AAA GCT GAT TTA GTG TCC  
 1404  
 Ile Leu Tyr Gly Phe Leu Asn Asn Gly Ile Lys Ala Asp Leu Val Ser  
 435 440 445  
 60  
 CTT ATA CAC TGT CTT CAT ATG TAA TAA TTCTCACTGT TTACCAAGGA  
 1452  
 Leu Ile His Cys Leu His Met \* \*  
 450 455  
 65  
 AAGAAC  
 1457  
 70 (2) INFORMATION FOR SEQ ID NO:4:

-156-

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 457 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

5

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

10	Met	Ser	Phe	Tyr	Ser	Lys	Gln	Asp	Tyr	Asn	Met	Asp	Leu	Glu	Leu	Asp
	1				5					10					15	
	Glu	Tyr	Tyr	Asn	Lys	Thr	Leu	Ala	Thr	Glu	Asn	Asn	Thr	Ala	Ala	Thr
				20					25					30		
15	Arg	Asn	Ser	Asp	Phe	Pro	Val	Trp	Asp	Asp	Tyr	Lys	Ser	Ser	Val	Asp
			35					40					45			
20	Asp	Leu	Gln	Tyr	Phe	Leu	Ile	Gly	Leu	Tyr	Thr	Phe	Val	Ser	Leu	Leu
	50						55					60				
	Gly	Phe	Met	Gly	Asn	Leu	Leu	Ile	Leu	Met	Ala	Leu	Met	Lys	Lys	Arg
	65					70				75						80
25	Asn	Gln	Lys	Thr	Thr	Val	Asn	Phe	Leu	Ile	Gly	Asn	Leu	Ala	Phe	Ser
					85					90						95
	Asp	Ile	Leu	Val	Val	Leu	Phe	Cys	Ser	Pro	Phe	Thr	Leu	Thr	Ser	Val
				100					105					110		
30	Leu	Leu	Asp	Gln	Trp	Met	Phe	Gly	Lys	Val	Met	Cys	His	Ile	Met	Pro
			115					120					125			
35	Phe	Leu	Gln	Cys	Val	Ser	Val	Leu	Val	Ser	Thr	Leu	Ile	Leu	Ile	Ser
	130						135					140				
	Ile	Ala	Ile	Val	Arg	Tyr	His	Met	Ile	Lys	His	Pro	Ile	Ser	Asn	Asn
	145				150					155						160
40	Leu	Thr	Ala	Asn	His	Gly	Tyr	Phe	Leu	Ile	Ala	Thr	Val	Trp	Thr	Leu
					165					170					175	
	Gly	Phe	Ala	Ile	Cys	Ser	Pro	Leu	Pro	Val	Phe	His	Ser	Leu	Val	Glu
				180					185					190		
45	Leu	Gln	Glu	Thr	Phe	Gly	Ser	Ala	Leu	Leu	Ser	Ser	Arg	Tyr	Leu	Cys
			195					200					205			
50	Val	Glu	Ser	Trp	Pro	Ser	Asp	Ser	Tyr	Arg	Ile	Ala	Phe	Thr	Ile	Ser
	210						215					220				
	Leu	Leu	Leu	Val	Gln	Tyr	Ile	Leu	Pro	Leu	Val	Cys	Leu	Thr	Val	Ser
	225				230					235						240
55	His	Thr	Ser	Val	Cys	Arg	Ser	Ile	Ser	Cys	Gly	Leu	Ser	Asn	Lys	Glu
					245					250					255	
	Asn	Arg	Leu	Glu	Glu	Asn	Glu	Met	Ile	Asn	Leu	Thr	Leu	His	Pro	Ser
				260				265						270		
60	Lys	Lys	Ser	Gly	Pro	Gln	Val	Lys	Leu	Ser	Gly	Ser	His	Lys	Trp	Ser
			275					280					285			
65	Tyr	Ser	Phe	Ile	Lys	Lys	His	Arg	Arg	Arg	Tyr	Ser	Lys	Lys	Thr	Ala
	290					295						300				
	Cys	Val	Leu	Pro	Ala	Pro	Glu	Arg	Pro	Ser	Gln	Glu	Asn	His	Ser	Arg
	305					310					315					320
70	Ile	Leu	Pro	Glu	Asn	Phe	Gly	Ser	Val	Arg	Ser	Gln	Leu	Ser	Ser	Ser

325 330 335  
 Ser Lys Phe Ile Pro Gly Val Pro Thr Cys Phe Glu Ile Lys Pro Glu  
 340 345 350  
 Glu Asn Ser Asp Val His Glu Leu Arg Val Lys Arg Ser Val Thr Arg  
 355 360 365  
 Ile Lys Lys Arg Ser Arg Ser Val Phe Tyr Arg Leu Thr Ile Leu Ile  
 370 375 380  
 Leu Val Phe Ala Val Ser Trp Met Pro Leu His Leu Phe His Val Val  
 385 390 395 400  
 Thr Asp Phe Asn Asp Asn Leu Ile Ser Asn Arg His Phe Lys Leu Val  
 405 410 415  
 Tyr Cys Ile Cys His Leu Leu Gly Met Met Ser Cys Cys Leu Asn Pro  
 420 425 430  
 Ile Leu Tyr Gly Phe Leu Asn Asn Gly Ile Lys Ala Asp Leu Val Ser  
 435 440 445  
 Leu Ile His Cys Leu His Met \* \*  
 450 455  
 (2) INFORMATION FOR SEQ ID NO:5:  
 (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 1054 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear  
 (ii) MOLECULE TYPE: DNA (genomic)  
 (ix) FEATURE:  
 (A) NAME/KEY: CDS  
 (B) LOCATION: 3..1004  
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:  
 TC ATG TGT CAC ATT ATG CCT TTT CTT CAA TGT GTG TCA GTT CTG GTT  
 47  
 Met Cys His Ile Met Pro Phe Leu Gln Cys Val Ser Val Leu Val  
 1 5 10 15  
 TCA ACT TTA ATT CTA ATA TCA ATT GCC ATT GTC AGG TAT CAT ATG ATC  
 95  
 Ser Thr Leu Ile Leu Ile Ser Ile Ala Ile Val Arg Tyr His Met Ile  
 20 25 30  
 AAG CAT CCT ATA TCT AAC AAT TTA ACA GCA AAC CAT GGC TAC TTC CTG  
 143  
 Lys His Pro Ile Ser Asn Asn Leu Thr Ala Asn His Gly Tyr Phe Leu  
 35 40 45  
 ATT GCT ACT GTC TGG ACA CTA GGT TTT GCG ATT TGT TCT CCC CTT CCA  
 191  
 Ile Ala Thr Val Trp Thr Leu Gly Phe Ala Ile Cys Ser Pro Leu Pro  
 50 55 60  
 GTG TTT CAC AGT CTG GTG GAA CTT CAG GAA ACA TTT GAC TCC GCA TTG  
 239  
 Val Phe His Ser Leu Val Glu Leu Gln Glu Thr Phe Asp Ser Ala Leu  
 65 70 75  
 CTG AGC AGC AGG TAT TTA TGT GTT GAG TCG TGG CCA TCT GAT TCG TAC  
 287

-158-

Leu Ser Ser Arg Tyr Leu Cys Val Glu Ser Trp Pro Ser Asp Ser Tyr  
 80 85 90 95  
 5 AGA ATC GCT TTT ACT ATC TCT TTA TTG CTA GTC CAG TAT ATT CTT CCC  
 335  
 Arg Ile Ala Phe Thr Ile Ser Leu Leu Leu Val Gln Tyr Ile Leu Pro  
 100 105 110  
 10 TTG GTG TGT CTA ACT GTG AGC CAT ACC AGT GTC TGC AGG AGT ATA AGC  
 383  
 Leu Val Cys Leu Thr Val Ser His Thr Ser Val Cys Arg Ser Ile Ser  
 115 120 125  
 15 TGC GGG TTG TCC AAC AAA GAA AAC AAA CTG GAA GAA AAC GAG ATG ATC  
 431  
 Cys Gly Leu Ser Asn Lys Glu Asn Lys Leu Glu Glu Asn Glu Met Ile  
 130 135 140  
 20 AAC TTA ACT CTT CAA CCA TTC AAA AAG AGT GGG CCT CAG GTG AAA CTT  
 479  
 Asn Leu Thr Leu Gln Pro Phe Lys Lys Ser Gly Pro Gln Val Lys Leu  
 145 150 155  
 25 TCC AGC AGC CAT AAA TGG AGC TAT TCA TTC ATC AGA AAA CAC AGG AGA  
 527  
 Ser Ser Ser His Lys Trp Ser Tyr Ser Phe Ile Arg Lys His Arg Arg  
 160 165 170 175  
 30 AGG TAC AGC AAG AAG ACG GCG TGT GTC TTA CCT GCT CCA GCA AGA CCT  
 575  
 Arg Tyr Ser Lys Lys Thr Ala Cys Val Leu Pro Ala Pro Ala Arg Pro  
 180 185 190  
 35 CCT CAA GAG AAC CAC TCA AGA ATG CTT CCA GAA AAC TTT GGT TCT GTA  
 623  
 Pro Gln Glu Asn His Ser Arg Met Leu Pro Glu Asn Phe Gly Ser Val  
 195 200 205  
 40 AGA AGT CAG CAT TCT TCA TCC AGT AAG TTC ATA CCG GGG GTC CCC ACC  
 671  
 Arg Ser Gln His Ser Ser Ser Ser Lys Phe Ile Pro Gly Val Pro Thr  
 210 215 220  
 45 TGC TTT GAG GTG AAA CCT GAA GAA AAC TCG GAT GTT CAT GAC ATG AGA  
 719  
 Cys Phe Glu Val Lys Pro Glu Glu Asn Ser Asp Val His Asp Met Arg  
 225 230 235  
 50 GTA AAC CGT TCT ATC ATG AGA ATC AAA AAG AGA TCC CGA AGT GTT TTC  
 767  
 Val Asn Arg Ser Ile Met Arg Ile Lys Lys Arg Ser Arg Ser Val Phe  
 240 245 250 255  
 55 TAT AGA CTA ACC ATA CTG ATA CTA GTG TTT GCC GTT AGC TGG ATG CCA  
 815  
 Tyr Arg Leu Thr Ile Leu Ile Leu Val Phe Ala Val Ser Trp Met Pro  
 260 265 270  
 60 CTA CAC CTT TTC CAT GTG GTA ACT GAT TTT AAT GAC AAC CTC ATT TCA  
 863  
 Leu His Leu Phe His Val Val Thr Asp Phe Asn Asp Asn Leu Ile Ser  
 275 280 285  
 65 AAC AGG CAT TTC AAA TTG GTG TAT TGC ATT TGT CAT TTG TTA GGC ATG  
 911  
 Asn Arg His Phe Lys Leu Val Tyr Cys Ile Cys His Leu Leu Gly Met  
 290 295 300  
 70 ATG TCC TGT TGT CTT AAT CCT ATT CTG TAT GGT TTT CTC AAT AAT GGG  
 959



-159-

Met Ser Cys Cys Leu Asn Pro Ile Leu Tyr Gly Phe Leu Asn Asn Gly  
 305 310 315

5 ATC AAA GCT GAT TTA ATT TCC CTT ATA CAG TGT CTT CAT ATG TCA  
 1004  
 Ile Lys Ala Asp Leu Ile Ser Leu Ile Gln Cys Leu His Met Ser  
 320 325 330

10 TAATTATTAA TGTTTACCAA GGAGACAACA AATGTTGGGA TCGTCTAAAA  
 1054

## (2) INFORMATION FOR SEQ ID NO:6:

15 (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 334 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

20 (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

25 Met Cys His Ile Met Pro Phe Leu Gln Cys Val Ser Val Leu Val Ser  
 1 5 10 15

Thr Leu Ile Leu Ile Ser Ile Ala Ile Val Arg Tyr His Met Ile Lys  
 20 25 30

30 His Pro Ile Ser Asn Asn Leu Thr Ala Asn His Gly Tyr Phe Leu Ile  
 35 40 45

Ala Thr Val Trp Thr Leu Gly Phe Ala Ile Cys Ser Pro Leu Pro Val  
 50 55 60

35 Phe His Ser Leu Val Glu Leu Gln Glu Thr Phe Asp Ser Ala Leu Leu  
 65 70 75 80

40 Ser Ser Arg Tyr Leu Cys Val Glu Ser Trp Pro Ser Asp Ser Tyr Arg  
 85 90 95

Ile Ala Phe Thr Ile Ser Leu Leu Leu Val Gln Tyr Ile Leu Pro Leu  
 100 105 110

45 Val Cys Leu Thr Val Ser His Thr Ser Val Cys Arg Ser Ile Ser Cys  
 115 120 125

Gly Leu Ser Asn Lys Glu Asn Lys Leu Glu Glu Asn Glu Met Ile Asn  
 130 135 140

50 Leu Thr Leu Gln Pro Phe Lys Lys Ser Gly Pro Gln Val Lys Leu Ser  
 145 150 155 160

55 Ser Ser His Lys Trp Ser Tyr Ser Phe Ile Arg Lys His Arg Arg Arg  
 165 170 175

Tyr Ser Lys Lys Thr Ala Cys Val Leu Pro Ala Pro Ala Arg Pro Pro  
 180 185 190

60 Gln Glu Asn His Ser Arg Met Leu Pro Glu Asn Phe Gly Ser Val Arg  
 195 200 205

Ser Gln His Ser Ser Ser Ser Lys Phe Ile Pro Gly Val Pro Thr Cys  
 210 215 220

65 Phe Glu Val Lys Pro Glu Glu Asn Ser Asp Val His Asp Met Arg Val  
 225 230 235 240

70 Asn Arg Ser Ile Met Arg Ile Lys Lys Arg Ser Arg Ser Val Phe Tyr  
 245 250 255

-160-

5	Arg	Leu	Thr	Ile	Leu	Ile	Leu	Val	Phe	Ala	Val	Ser	Trp	Met	Pro	Leu
				260					265					270		
	His	Leu	Phe	His	Val	Val	Thr	Asp	Phe	Asn	Asp	Asn	Leu	Ile	Ser	Asn
			275					280					285			
10	Arg	His	Phe	Lys	Leu	Val	Tyr	Cys	Ile	Cys	His	Leu	Leu	Gly	Met	Met
		290					295					300				
	Ser	Cys	Cys	Leu	Asn	Pro	Ile	Leu	Tyr	Gly	Phe	Leu	Asn	Asn	Gly	Ile
	305				310					315					320	
	Lys	Ala	Asp	Leu	Ile	Ser	Leu	Ile	Gln	Cys	Leu	His	Met	Ser		
				325						330						

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 24 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

TGGATCAGTG GATGTTTGGC AAAG  
24

(2) INFORMATION FOR SEO ID NO: 8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 28 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEO ID NO:8:

GTCTGTAGAA AACACTTCGA GATCTCTT  
28

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

CTTCCAGTGT TTCACAGTCT GGTGG  
25

(2) INFORMATION FOR SEQ ID NO:10:

70

-161-

- 5 (i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 25 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- 10 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:
- 15 CTGAGCAGCA GGTATTTATG TGTTG  
25
- (2) INFORMATION FOR SEQ ID NO:11:
- 20 (i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 28 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear
- 25 (ii) MOLECULE TYPE: cDNA
- 30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:
- CTGGATGAAG AATGCTGACT TCTTAGAG  
28
- 35 (2) INFORMATION FOR SEQ ID NO:12:
- 40 (i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 25 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- 45 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:
- 50 TTCTTGAGTG GTTCTCTTGA GGAGG  
25

-162-

What is claimed is:

- 5 1. A method of modifying feeding behavior of a subject which comprises administering to the subject an amount of a compound which is a Y5 receptor agonist or antagonist effective to increase or decrease the consumption of food by the subject so as to thereby modify feeding behavior of the subject.  
10
2. The method of claim 1, wherein the compound is a Y5 receptor antagonist and the amount is effective to decrease the consumption of food by the subject.  
15
3. The method of either of claims 1 or 2, wherein the compound is administered in combination with food.
- 20 4. The method of claim 1, wherein the compound is a Y5 receptor agonist and the amount is effective to increase the consumption of food by the subject.
- 25 5. The method of either of claims 1 or 4, wherein the compound is administered in combination with food.
6. The method of claim 1, wherein the subject is a vertebrate, a mammal, a human or a canine.
- 30 7. A method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a non-peptidyl compound which is a Y5 receptor antagonist effective to inhibit the activity of the subject's Y5 receptor, wherein  
35 the binding of the compound to the human Y5 receptor is characterized by a  $K_i$  less than 100 nanomolar when measured in the presence of  $^{125}\text{I}$ -

-163-

PYY.

8. The method of claim 7, wherein the compound has a  $K_i$  less than 50 nanomolar.
- 5 9. The method of claim 8, wherein the compound has a  $K_i$  less than 10 nanomolar.
- 10 10. The method of claim 9, wherein binding of the compound to any other human Y-type receptor is characterized by a  $K_i$  greater than 10 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.
- 15 11. The method of claim 9, wherein the binding of the compound to each of the human Y1, human Y2 and human Y4 receptors is characterized by a  $K_i$  greater than 10 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.
- 20 12. The method of claim 10, wherein the binding of the compound to any other human Y-type receptor is characterized by a  $K_i$  greater than 50 nanomolar.
- 25 13. The method of claim 12, wherein the binding of the compound to any other human Y-type receptor is characterized by a  $K_i$  greater than 100 nanomolar.
- 30 14. The method of claim 7, wherein the compound binds to the human Y5 receptor with an affinity greater than ten-fold higher than the affinity with which the compound binds to any other human Y-type receptor.
- 35 15. The method of claim 7, wherein the compound binds to the human Y5 receptor with an affinity greater than ten-fold higher than the affinity with which the compound binds to each of the human Y1, human

-164-

Y2 and human Y4 receptors.

16. The method of claim 7, wherein the feeding disorder is obesity or bulimia.
- 5
17. The method of claim 7, wherein the subject is a vertebrate, a mammal, a human or a canine.
18. A method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a peptidyl compound which is a Y5 receptor antagonist effective to inhibit the activity of the subject's Y5 receptor, wherein the compound's binding to the human Y5 receptor is characterized by a  $K_i$  less than 10 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.
- 10
19. The method of claim 18, wherein the compound's binding is characterized by a  $K_i$  less than 1 nanomolar.
- 15
20. The method of claim 18, wherein the compound's binding to any other human Y-type receptor is characterized by a  $K_i$  greater than 10 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.
- 20
21. The method of claim 18, wherein the compound's binding to each of the human Y1, human Y2 and human Y4 receptors is characterized by a  $K_i$  greater than 10 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.
- 25
22. The method of claim 20, wherein the compound's binding to any other human Y-type receptor is characterized by a  $K_i$  greater than 50 nanomolar.
- 30
23. The method of claim 22, wherein the compound's
- 35

-165-

binding to any other human Y-type receptor is characterized by a  $K_i$  greater than 100 nanomolar.

5        24. The method of claim 18, wherein the compound binds to the human Y5 receptor with an affinity greater than ten-fold higher than the affinity with which the compound binds to any other human Y-type receptor.

10       25. The method of claim 18, wherein the compound binds to the human Y5 receptor with an affinity greater than ten-fold higher than the affinity with which the compound binds to each of the human Y1, human Y2 and human Y4 receptors.

15       26. The method of claim 18, wherein the feeding disorder is obesity or bulimia.

20       27. The method of claim 18, wherein the subject is a vertebrate, a mammal, a human or a canine.

25       28. A method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a non-peptidyl compound which is a Y5 receptor agonist effective to increase the activity of the subject's Y5 receptor, wherein

30           (a) the binding of the compound to the human Y5 receptor is characterized by a  $K_i$  less than 100 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY; and

35           (b) the binding of the compound to any other human Y-type receptor is characterized by a  $K_i$  greater than 1000 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.

-166-

29. The method of claim 28, wherein the binding of the compound to the human Y5 receptor is characterized by a  $K_i$  less than 10 nanomolar.
- 5 30. A method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a non-peptidyl compound which is a Y5 receptor agonist effective to increase the activity of the subject's Y5 receptor, wherein
- 10 (a) the binding of the compound to the human Y5 receptor is characterized by a  $K_i$  less than 1 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY; and
- 15 (b) the compound's binding to any other human Y-type receptor is characterized by a  $K_i$  greater than 100 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.
- 20 31. The method of claim 28, wherein the compound binds to the human Y5 receptor with an affinity greater than ten-fold higher than the affinity with which the compound binds to any other human Y-type receptor.
- 25 32. The method of claim 28, wherein the compound binds to the human Y5 receptor with an affinity greater than ten-fold higher than the affinity with which the compound binds to each of the human Y1, human Y2 and human Y4 receptors.
- 30 33. The method of claim 28, wherein the feeding disorder is anorexia.
- 35 34. The method of claim 28, wherein the subject is a vertebrate, a mammal, a human or a canine.



-167-

35. A method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a peptidyl compound which is a Y5 receptor agonist effective to increase the activity of the subject's Y5 receptor, wherein

(a) the binding of the compound to the human Y5 receptor is characterized by a  $K_i$  less than 1 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY; and

(b) the binding of the compound to any other human Y-type receptor is characterized by a  $K_i$  greater than 25 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.

36. A method of treating a feeding disorder in a subject which comprises administering to the subject an amount of a peptidyl compound which is a Y5 receptor agonist effective to increase the activity of the subject's Y5 receptor, wherein

(a) the binding of the compound to the human Y5 receptor is characterized by a  $K_i$  less than 0.1 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY; and

(b) the binding of the compound to any other human Y-type receptor is characterized by a  $K_i$  greater than 1 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.

37. The method of claim 36, wherein the binding of the agonist to any other human Y-type receptor is characterized by a  $K_i$  greater than 10 nanomolar.

38. A method of treating a feeding disorder in a

-168-

subject which comprises administering to the subject an amount of a peptidyl compound which is a Y5 receptor agonist effective to increase the activity of the subject's Y5 receptor, wherein

5

(a) the binding of the compound to the human Y5 receptor is characterized by a  $K_i$  less than 0.01 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY; and

10

(b) the binding of the compound to any other human Y-type receptor is characterized by a  $K_i$  greater than 1 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.

15

39. The method of claim 35, wherein the compound binds to the human Y5 receptor with an affinity greater than ten-fold higher than the affinity with which the compound binds to any other human Y-type receptor.

20

40. The method of claim 35, wherein the compound binds to the human Y5 receptor with an affinity greater than ten-fold higher than the affinity with which the compound binds to each of the human Y1, human Y2 and human Y4 receptors.

25

41. The method of claim 35, wherein the feeding disorder is anorexia.

30

42. The method of claim 35, wherein the subject is a vertebrate, a mammal, a human or a canine.

43. An isolated nucleic acid encoding a Y5 receptor.

35

44. The nucleic acid of claim 43, wherein the nucleic acid is DNA.

-169-

45. The DNA of claim 44, wherein the DNA is cDNA.
46. The DNA of claim 44, wherein the DNA is genomic DNA.
- 5 47. The nucleic acid of claim 43, wherein the nucleic acid is RNA.
- 10 48. The nucleic acid of claim 43, wherein the nucleic acid encodes a vertebrate Y5 receptor.
49. The nucleic acid of claim 43, wherein the nucleic acid encodes a mammalian Y5 receptor.
- 15 50. The nucleic acid of claim 43, wherein the nucleic acid encodes a human Y5 receptor.
- 20 51. The nucleic acid of claim 50, wherein the nucleic acid encodes a receptor characterized by an amino acid sequence in the transmembrane region which has a homology of 60% or higher to the amino acid sequence in the transmembrane region of the human Y5 receptor shown in Figure 6.
- 25 52. The nucleic acid of claim 50, wherein the human Y5 receptor has substantially the same amino acid sequence as that shown in Figure 6.
- 30 53. The nucleic acid of claim 50, wherein the human Y5 receptor has the amino acid sequence shown in Figure 6.
54. The nucleic acid of claim 43, wherein the nucleic acid encodes a rat Y5 receptor.
- 35 55. The nucleic acid of claim 54, wherein the rat Y5 receptor has substantially the same amino acid

-170-

sequence as that shown in Figure 4.

56. The nucleic acid of claim 54, wherein the rat Y5  
5 receptor has the amino acid sequence shown in  
Figure 4.
57. The nucleic acid of claim 43, wherein the nucleic  
acid encodes a canine Y5 receptor.
- 10 58. The nucleic acid molecule of claim 57, wherein the  
canine Y5 receptor has substantially the same  
amino acid sequence as that shown in Figure 15.
- 15 59. The nucleic acid of claim 57, wherein the canine  
Y5 receptor has the amino acid sequence shown in  
Figure 15.
60. A purified Y5 receptor protein.
- 20 61. A vector comprising the nucleic acid of claim 43.
62. A vector comprising the nucleic acid of claim 50.
63. A vector comprising the nucleic acid of claim 54.  
25
64. A vector comprising the nucleic acid of claim 57.
65. A vector of claim 61 adapted for expression in a  
bacterial cell which comprises the regulatory  
30 elements necessary for expression of the nucleic  
acid in the bacterial cell operatively linked to  
the nucleic acid encoding a Y5 receptor as to  
permit expression thereof.
- 35 66. A vector of claim 61 adapted for expression in a  
yeast cell which comprises the regulatory elements  
necessary for expression of the nucleic acid in

-171-

subject which comprises administering to the subject an amount of a peptidyl compound which is a Y5 receptor agonist effective to increase the activity of the subject's Y5 receptor, wherein

5

(a) the binding of the compound to the human Y5 receptor is characterized by a  $K_i$  less than 0.01 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY; and

10

(b) the binding of the compound to any other human Y-type receptor is characterized by a  $K_i$  greater than 1 nanomolar when measured in the presence of  $^{125}\text{I}$ -PYY.

15

39. The method of claim 35, wherein the compound binds to the human Y5 receptor with an affinity greater than ten-fold higher than the affinity with which the compound binds to any other human Y-type receptor.

20

40. The method of claim 35, wherein the compound binds to the human Y5 receptor with an affinity greater than ten-fold higher than the affinity with which the compound binds to each of the human Y1, human Y2 and human Y4 receptors.

25

41. The method of claim 35, wherein the feeding disorder is anorexia.

30

42. The method of claim 35, wherein the subject is a vertebrate, a mammal, a human or a canine subject.

43. An isolated nucleic acid encoding a Y5 receptor.

35

44. The nucleic acid of claim 43, wherein the nucleic acid is DNA.

-172-

- 5 mammalian cell which comprises the regulatory elements necessary for expression of the DNA in the mammalian cell operatively linked to the DNA encoding the rat Y5 receptor as to permit expression thereof.
75. A vector of claim 74 wherein the vector is a plasmid.
- 10 76. The plasmid of claim 75 designated pcEXV-rY5 (ATCC Accession No. 75944).
- 15 77. A vector of claim 64 adapted for expression in a mammalian cell which comprises the regulatory elements necessary for expression of the DNA in the mammalian cell operatively linked to the DNA encoding the canine Y5 receptor as to permit expression thereof.
- 20 78. The vector of claim 77 designated Y5-bd-8 (ATCC Accession No.        ).
79. The vector of claim 78 designated Y5-bd-5 (ATCC Accession No.        ).
- 25 80. A mammalian cell comprising the vector of any one of claims 70, 71, 74, or 77.
81. A mammalian cell of claim 80, wherein the cell is non-neuronal in origin.
- 30 82. A mammalian cell of claim 80, wherein the mammalian cell is a COS-7 cell.
- 35 83. A mammalian cell of claim 80, wherein the mammalian cell is a 293 human embryonic kidney cell.

-173-

84. The cell of claim 83 designated 293-rY5-14 (ATCC Accession No. CRL 11757).
- 5 85. A mammalian cell of claim 80, wherein the mammalian cell is a NIH-3T3 cell.
86. The cell of claim 81 designated [designation] (ATCC Accession No. CRL [n#]).
- 10 87. A mammalian cell of claim 80, wherein the mammalian cell is a LM(tk-) cell.
88. The cell of claim 87 designated [designation] (ATCC Accession No. CRL [l#]).
- 15 89. An insect cell comprising the vector of claim 67.
90. An insect cell of claim 89, wherein the insect cell is an Sf9 cell.
- 20 91. An insect cell of claim 89, wherein the insect cell is an Sf21 cell.
92. A membrane preparation isolated from the cell of claim 80.
- 25 93. A nucleic acid probe comprising a nucleic acid of at least 15 nucleotides capable of specifically hybridizing with a unique sequence included within the sequence of a nucleic acid encoding a Y5 receptor of claim 43.
- 30 94. A nucleic acid probe of claim 93, wherein the nucleic acid is DNA.
- 35 95. A nucleic acid probe of claim 93, wherein the nucleic acid is RNA.

-174-

- 5           96. An antisense oligonucleotide having a sequence capable of specifically hybridizing to mRNA encoding a Y5 receptor of claim 47 so as to prevent translation of the mRNA.
97. An antisense oligonucleotide having a sequence capable of specifically hybridizing to the genomic DNA of claim 46.
- 10       98. An antisense oligonucleotide of either of claims 96 or 97, wherein the oligonucleotide comprises chemically modified nucleotides or nucleotide analogues.
- 15       99. An antibody capable of binding to a Y5 receptor of claim 43.
100. An antibody of claim 99, wherein the Y5 receptor is a human Y5 receptor.
- 20       101. An antibody capable of competitively inhibiting the binding of the antibody of claim 99 to a Y5 receptor.
- 25       102. An antibody of claim 99 wherein the antibody is a monoclonal antibody.
103. A monoclonal antibody of claim 102 directed to an epitope of a Y5 receptor present on the surface of
- 30       a Y5 receptor expressing cell.
104. A pharmaceutical composition comprising an amount of the oligonucleotide of claim 96 capable of passing through a cell membrane effective to
- 35       reduce expression of a human Y5 receptor and a pharmaceutically acceptable carrier capable of passing through a cell membrane.



-175-

105. A pharmaceutical composition of claim 104, wherein the oligonucleotide is coupled to a substance which inactivates mRNA.
- 5 106. A pharmaceutical composition of claim 105, wherein the substance which inactivates mRNA is a ribozyme.
- 10 107. A pharmaceutical composition of claim 104, wherein the pharmaceutically acceptable carrier comprises a structure which binds to a receptor on a cell capable of being taken up by the cells after binding to the structure.
- 15 108. A pharmaceutical composition of claim 107 wherein the structure of the pharmaceutically acceptable carrier is capable of binding to a receptor which is specific for a selected cell type.
- 20 109. A pharmaceutical composition which comprises an amount of the antibody of claim 99 effective to block binding of a ligand to the Y5 receptor and a pharmaceutically acceptable carrier.
- 25 110. A transgenic nonhuman mammal expressing DNA encoding a human Y5 receptor of claim 50.
- 30 111. A transgenic nonhuman mammal comprising a homologous recombination knockout of the native Y5 receptor.
- 35 112. A transgenic nonhuman mammal whose genome comprises antisense DNA complementary to DNA encoding a human Y5 receptor of claim 50 so placed as to be transcribed into antisense mRNA which is complementary to mRNA encoding a Y5 receptor and which hybridizes to mRNA encoding a Y5 receptor

-176-

thereby reducing its translation.

- 5 113. The transgenic nonhuman mammal of either of claims 110 or 111, wherein the DNA encoding a human Y5 receptor additionally comprises an inducible promoter.
- 10 114. The transgenic nonhuman mammal of either of claims 110 or 112, wherein the DNA encoding a human Y5 receptor additionally comprises tissue specific regulatory elements.
- 15 115. A transgenic nonhuman mammal of any of claims 120, 121 or 122, wherein the transgenic nonhuman mammal is a mouse.
- 20 116. A method for determining whether a ligand can specifically bind to a Y5 receptor which comprises contacting a cell transfected with and expressing DNA encoding the Y5 receptor with the ligand under conditions permitting binding of ligands to such receptor, and detecting the presence of any such ligand specifically bound to the Y5 receptor, so
- 25 as to thereby determine whether the ligand specifically binds to the Y5 receptor.
117. A method of claim 116 wherein the Y5 receptor is a human Y5 receptor.
- 30 118. A method for determining whether a ligand can specifically bind to a Y5 receptor which comprises contacting a cell transfected with and expressing DNA encoding the Y5 receptor with the ligand under conditions permitting binding of ligands to such
- 35 receptor, and detecting the presence of any such ligand specifically bound to the Y5 receptor, so as to thereby determine whether the ligand

-177-

specifically binds to the Y5 receptor, such Y5 receptor being characterized by an amino acid sequence in the transmembrane region, such amino acid sequence having 60% homology or higher to the amino acid sequence in the transmembrane region of the Y5 receptor shown in Figure 6.

119. A method for determining whether a ligand can specifically bind to a human Y5 receptor which comprises contacting a cell transfected with and expressing DNA encoding the human Y5 receptor with the ligand under conditions permitting binding of ligands to such receptor, and detecting the presence of any such ligand specifically bound to the human Y5 receptor, so as to thereby determine whether the ligand specifically binds to the human Y5 receptor, such human Y5 receptor having substantially the same amino acid sequence as that shown in Figure 6.

120. A method for determining whether a ligand can specifically bind to a Y5 receptor which comprises preparing a cell extract from cells transfected with and expressing DNA encoding the Y5 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with the ligand under conditions permitting binding of ligands to such receptor, and detecting the presence of the ligand specifically bound to the Y5 receptor, so as to thereby determine whether the ligand specifically binds to the Y5 receptor.

121. A method of claim 120 wherein the Y5 receptor is a human Y5 receptor.

122. A method for determining whether a ligand can specifically bind to a Y5 receptor which comprises

-178-

5 preparing a cell extract from cells transfected  
with and expressing DNA encoding the Y5 receptor,  
isolating a membrane fraction from the cell  
extract, contacting the membrane fraction with the  
ligand under conditions permitting binding of  
ligands to the Y5 receptor, and detecting the  
presence of the ligand specifically bound to the  
Y5 receptor, so as to thereby determine whether  
the ligand can specifically bind to the Y5  
10 receptor, such Y5 receptor being characterized by  
an amino acid sequence in the transmembrane region  
having 60% homology or higher to the amino acid  
sequence in the transmembrane region of the Y5  
receptor shown in Figure 6.

15  
123. A method for determining whether a ligand can  
specifically bind to a human Y5 receptor which  
comprises preparing a cell extract from cells  
transfected with and expressing DNA encoding the  
human Y5 receptor, isolating a membrane fraction  
20 from the cell extract, contacting the membrane  
fraction with the ligand under conditions  
permitting binding of ligands to the human Y5  
receptor, and detecting the presence of the ligand  
specifically bound to the human Y5 receptor, so as  
25 to thereby determine whether the ligand can  
specifically bind to the human Y5 receptor, such  
human Y5 receptor having substantially the same  
amino acid sequence shown in Figure 6.

30  
124. A method of any one of claims 116, 117, 118, 119,  
120, 121, 122, or 123, wherein the ligand is not  
previously known.

35 125. A ligand determined by the method of claim 124.

126. A method for determining whether a ligand is a Y5

-179-

5 receptor agonist which comprises contacting a cell transfected with and expressing DNA encoding the Y5 receptor with the ligand under conditions permitting activation of the Y5 receptor, and detecting an increase in Y5 receptor activity, so as to thereby determine whether the ligand is a Y5 receptor agonist.

10 127. A method for determining whether a ligand is a Y5 receptor agonist which comprises preparing a cell extract from cells transfected with and expressing DNA encoding the Y5 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with the ligand under conditions  
15 permitting the activation of the Y5 receptor, and detecting an increase in Y5 receptor activity, so as to thereby determine whether the ligand is a Y5 receptor agonist.

20 128. A method of either of claims 126 or 127, wherein the Y5 receptor is a human Y5 receptor.

25 129. A method for determining whether a ligand is a Y5 receptor antagonist which comprises contacting a cell transfected with and expressing DNA encoding the Y5 receptor with the ligand in the presence of a known Y5 receptor agonist, such as PYY, under conditions permitting the activation of the Y5  
30 receptor, and detecting a decrease in Y5 receptor activity, so as to thereby determine whether the ligand is a Y5 receptor antagonist.

35 130. A method for determining whether a ligand is a Y5 receptor antagonist which comprises preparing a cell extract from cells transfected with and expressing DNA encoding the Y5 receptor, isolating a membrane fraction from the cell extract,

-180-

- 5                   contacting the membrane fraction with the ligand  
                  in the presence of a known Y5 receptor agonist,  
                  such as PYY, under conditions permitting the  
                  activation of the Y5 receptor, and detecting a  
10                   decrease in Y5 receptor activity, so as to thereby  
                  determine whether the ligand is a Y5 receptor  
                  antagonist.
- 15                   131. A method of either of claims 129 or 130, wherein  
                  the Y5 receptor is a human Y5 receptor.
132. A method of any one of claims 116, 117, 118, 119,  
                  120, 121, 122, 123, 124, 126, 127, 128, 129, 130,  
                  or 131, wherein the cell is an insect cell.
133. A method of any one of claims 116, 117, 118, 119,  
                  120, 121, 122, 123, 124, 126, 127, 128, 129, 130,  
                  or 131, wherein the cell is a mammalian cell.
- 20                   134. A method of claim 133, wherein the cell is  
                  nonneuronal in origin.
135. A method of claim 134, wherein the nonneuronal  
                  cell is a COS-7 cell, 293 human embryonic kidney  
25                   cell, NIH-3T3 cell or LM(tk-) cell.
136. A method of claim 133 wherein the ligand is not  
                  previously known.
- 30                   137. A Y5 ligand determined by the method of claim 136.
138. A pharmaceutical composition which comprises an  
                  amount of a Y5 receptor agonist determined by the  
                  method of either of claims 126 or 127 effective to  
35                   increase activity of a Y5 receptor and a  
                  pharmaceutically acceptable carrier.

-181-

139. A pharmaceutical composition of claim 138 wherein the Y5 receptor agonist is not previously known.
- 5 140. A pharmaceutical composition which comprises an amount of a Y5 receptor antagonist determined by the method of either of claims 129 or 130 effective to reduce activity of a Y5 receptor and a pharmaceutically acceptable carrier.
- 10 141. A pharmaceutical composition of claim 140 wherein the Y5 receptor antagonist is not previously known.
- 15 142. A method of screening a plurality of chemical compounds not known to bind to a Y5 receptor to identify a compound which specifically binds to the Y5 receptor, which comprises
- 20 (a) contacting a cell transfected with and expressing DNA encoding the Y5 receptor with a compound known to bind specifically to the Y5 receptor;
- 25 (b) contacting the preparation of step (a) with the plurality of compounds not known to bind specifically to the Y5 receptor, under conditions permitting binding of compounds known to bind the Y5 receptor;
- 30 (c) determining whether the binding of the compound known to bind to the Y5 receptor is reduced in the presence of the compounds, relative to the binding of the compound in the absence of the plurality of compounds;
- 35 and if so
- (d) separately determining the binding to the Y5

-182-

receptor of each compound included in the plurality of compounds, so as to thereby identify the compound which specifically binds to the Y5 receptor.

5

143. A method of screening a plurality of chemical compounds not known to bind to a Y5 receptor to identify a compound which specifically binds to the Y5 receptor, which comprises

10

(a) preparing a cell extract from cells transfected with and expressing DNA encoding the Y5 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with a compound known to bind specifically to the Y5 receptor;

15

(b) contacting preparation of step (a) with the plurality of compounds not known to bind specifically to the Y5 receptor, under conditions permitting binding of compounds known to bind the Y5 receptor;

20

(c) determining whether the binding of the compound known to bind to the Y5 receptor is reduced in the presence of the compounds, relative to the binding of the compound in the absence of the plurality of compounds; and if so

25

30

(d) separately determining the binding to the Y5 receptor of each compound included in the plurality of compounds, so as to thereby identify the compound which specifically binds to the Y5 receptor.

35

144. A method of claim 142 or claim 143 wherein the Y5



-183-

receptor is a human Y5 receptor.

145. A method of screening a plurality of chemical compounds not known to activate a Y5 receptor to identify a compound which activates the Y5 receptor which comprises

(a) contacting a cell transfected with and expressing the Y5 receptor with the plurality of compounds not known to bind specifically to the Y5 receptor, under conditions permitting activation of the Y5 receptor;

(b) determining whether the activity of the Y5 receptor is increased in the presence of the compounds; and if so

(c) separately determining whether the activation of the Y5 receptor is increased by each compound included in the plurality of compounds, so as to thereby identify the compound which activates the Y5 receptor.

146. A method of screening a plurality of chemical compounds not known to activate a Y5 receptor to identify a compound which activates the Y5 receptor which comprises

(a) preparing a cell extract from cells transfected with and expressing DNA encoding the Y5 receptor, isolating a membrane fraction from the cell extract, contacting the membrane fraction with the plurality of compounds not known to bind specifically to the Y5 receptor, under conditions permitting activation of the Y5 receptor;

-184-

- (b) determining whether the activity of the Y5 receptor is increased in the presence of the compounds; and if so
- 5 (c) separately determining whether the activation of the Y5 receptor is increased by each compound included in the plurality of compounds, so as to thereby identify the compound which activates the Y5 receptor.
- 10
147. A method of claim 145 or claim 146 wherein the Y5 receptor is a human Y5 receptor.
- 15 148. A method of screening a plurality of chemical compounds not known to inhibit the activation of a Y5 receptor to identify a compound which inhibits the activation of the Y5 receptor, which comprises
- 20 (a) contacting a cell transfected with and expressing the Y5 receptor with the plurality of compounds in the presence of a known Y5 receptor agonist, under conditions permitting activation of the Y5 receptor;
- 25 (b) determining whether the activation of the Y5 receptor is reduced in the presence of the plurality of compounds, relative to the activation of the Y5 receptor in the absence
- 30 of the plurality of compounds; and if so
- (c) separately determining the inhibition of activation of the Y5 receptor for each compound included in the plurality of
- 35 compounds, so as to thereby identify the compound which inhibits the activation of the Y5 receptor.

-185-

149. A method of screening a plurality of chemical compounds not known to inhibit the activation of a Y5 receptor to identify a compound which inhibits the activation of the Y5 receptor, which  
5 comprises

(a) preparing a cell extract from cells transfected with and expressing DNA encoding the Y5 receptor, isolating a membrane  
10 fraction from the cell extract, contacting the membrane fraction with the plurality of compounds in the presence of a known Y5 receptor agonist, under conditions permitting activation of the Y5 receptor;

15 (b) determining whether the activation of the Y5 receptor is reduced in the presence of the plurality of compounds, relative to the activation of the Y5 receptor in the absence of the plurality of compounds; and if so  
20

(c) separately determining the inhibition of activation of the Y5 receptor for each compound included in the plurality of  
25 compounds, so as to thereby identify the compound which inhibits the activation of the Y5 receptor.

150. A method of claim 148 or claim 149, wherein the Y5  
30 receptor is a human Y5 receptor.

151. A method of any one of claims 143 to 150, wherein the cell is a mammalian cell.

35 152. A method of claim 151, wherein the cell is non-neuronal in origin.

-186-

153. The method of claim 152 wherein the nonneuronal cell is a COS-7 cell, a 293 human embryonic kidney cell, a LM(tk-) cell or an NIH-3T3 cell.
- 5 154. A pharmaceutical composition comprising a drug identified by the method of claim 147 and a pharmaceutically acceptable carrier.
- 10 155. A pharmaceutical composition comprising a drug identified by the method of claim 150 and a pharmaceutically acceptable carrier.
- 15 156. A method of detecting expression of Y5 receptor by detecting the presence of mRNA coding for the Y5 receptor which comprises obtaining total mRNA from the cell and contacting the mRNA so obtained with the nucleic acid probe of claim 93 under hybridizing conditions, detecting the presence of mRNA hybridized to the probe, and thereby  
20 detecting the expression of the Y5 receptor by the cell.
- 25 157. A method of treating an abnormality in a subject, wherein the abnormality is alleviated by the inhibition of a Y5 receptor which comprises administering to a subject an effective amount of the pharmaceutical composition of any of claims 104, 105, 106, 107, 108, 109, 140, 141 or 155 effective to decrease the activity of the Y5  
30 receptor in the subject, thereby treating the abnormality in the subject.
- 35 158. The method of claim 157, wherein the abnormality is obesity or bulimia.
159. A method of treating an abnormality in a subject wherein the abnormality is alleviated by the

-187-

activation of a Y5 receptor which comprises administering to a subject an effective amount of the pharmaceutical composition of any of claims 148, 139, or 154 effective to activate the Y5 receptor in the subject.

5

160. The method of claim 159, wherein the abnormal condition is anorexia.

10

161. A method of detecting the presence of a human Y5 receptor on the surface of a cell which comprises contacting the cell with the antibody of claim 99 under conditions permitting binding of the antibody to the receptor, detecting the presence of the antibody bound to the cell, and thereby detecting the presence of a human Y5 receptor on the surface of the cell.

15

20

162. A method of determining the physiological effects of varying levels of activity of human Y5 receptors which comprises producing a transgenic nonhuman mammal of claim 110 whose levels of human Y5 receptor activity are varied by use of an inducible promoter which regulates human Y5 receptor expression.

25

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163. A method of determining the physiological effects of varying levels of activity of human Y5 receptors which comprises producing a panel of transgenic nonhuman mammals of claim 110 each expressing a different amount of human Y5 receptor.

35

164. A method for identifying an antagonist capable of alleviating an abnormality wherein the abnormality is alleviated by decreasing the activity of a human Y5 receptor comprising administering the

-188-

- antagonist to the transgenic nonhuman mammal of any of claims 110 to 115, and determining whether the substance alleviates the physical and behavioral abnormalities displayed by the transgenic nonhuman mammal as a result of overactivity of a human Y5 receptor, the alleviation of the abnormality indicating the identification of an antagonist.
- 5
- 10      165. An antagonist identified by the method of claim 164.
- 15      166. A pharmaceutical composition comprising an antagonist identified by the method of claim 164 and a pharmaceutically acceptable carrier.
- 20      167. A method of treating an abnormality in a subject wherein the abnormality is alleviated by decreasing the activity of a human Y5 receptor which comprises administering to a subject an effective amount of the pharmaceutical composition of claim 166, thereby treating the abnormality.
- 25      168. A method for identifying an agonist capable of alleviating an abnormality in a subject wherein the abnormality is alleviated by increasing the activity of a human Y5 receptor comprising administering the agonist to the transgenic nonhuman mammal of claims 110 to 115, and determining whether the substance alleviates the physical and behavioral abnormalities displayed by the transgenic nonhuman mammal, the alleviation of the abnormality indicating the identification of an agonist.
- 30
- 35      169. An agonist identified by the method of claim 168.

-189-

170. A pharmaceutical composition comprising an agonist identified by the method of claim 168 and a pharmaceutically acceptable carrier.
- 5 171. A method for treating an abnormality in a subject wherein the abnormality is alleviated by increasing the activity of a human Y5 receptor which comprises administering to a subject an effective amount of the pharmaceutical composition of claim 170, thereby treating the abnormality.
- 10
172. A method for diagnosing a predisposition to a disorder associated with the activity of a specific human Y5 receptor allele which comprises:
- 15
- a. obtaining DNA of subjects suffering from the disorder;
  - b. performing a restriction digest of the DNA with a panel of restriction enzymes;
  - c. electrophoretically separating the resulting DNA fragments on a sizing gel;
  - d. contacting the resulting gel with a nucleic acid probe capable of specifically hybridizing with a unique sequence included within the sequence of a nucleic acid molecule encoding a human Y5 receptor and labelled with a detectable marker;
  - e. detecting labelled bands which have hybridized to the DNA encoding a human Y5 receptor of claim 50 labelled with a detectable marker to create a unique band pattern specific to the DNA of subjects suffering from the disorder;
- 20
- 25
- 30
- 35

-190-

- f. preparing DNA obtained for diagnosis by steps a-e; and
- 5 g. comparing the unique band pattern specific to the DNA of subjects suffering from the disorder from step e and the DNA obtained for diagnosis from step f to determine whether the patterns are the same or different and to diagnose thereby
- 10 predisposition to the disorder if the patterns are the same.
173. The method of claim 172 wherein a disorder associated with the activity of a specific human
- 15 Y5 receptor allele is diagnosed.
174. A method of preparing the purified Y5 receptor of claim 60 which comprises:
- 20 a. inducing cells to express Y5 receptor;
- b. recovering the receptor from the induced cells; and
- 25 c. purifying the receptor so recovered.
175. A method of preparing the purified Y5 receptor of claim 60 which comprises:
- 30 a. inserting nucleic acid encoding Y5 receptor in a suitable vector;
- b. introducing the resulting vector in a
- 35 suitable host cell;
- c. placing the resulting cell in suitable



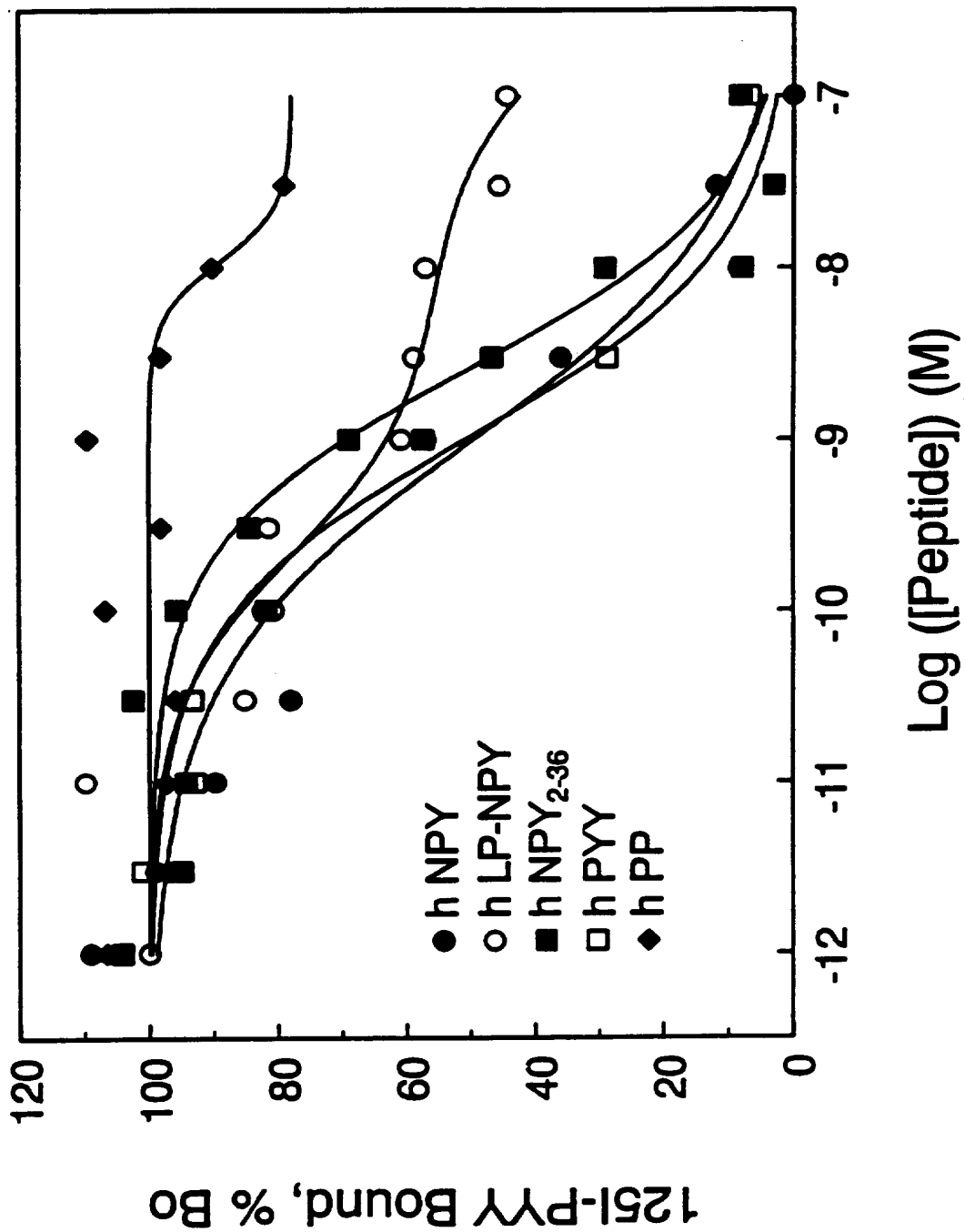
-191-

condition permitting the production of the isolated Y5 receptor;

- 5           d.   recovering the receptor produced by the  
            resulting cell; and
- e.   purifying the receptor so recovered.

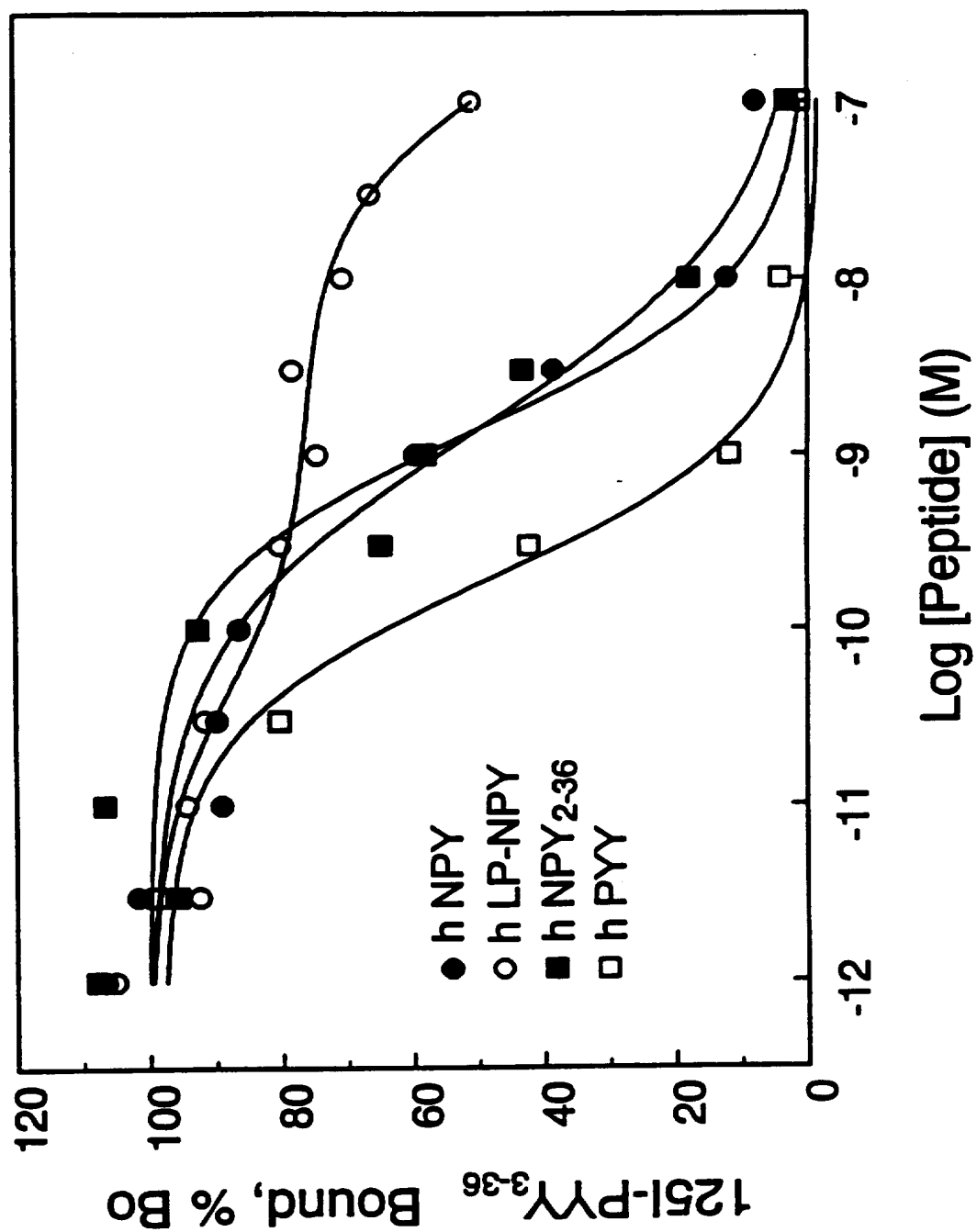
1/38

FIGURE 1



2/38

FIGURE 2



3/38

FIGURE 3

1	TTAGTTTGTCTGTGAGAACGTTAGAGTTATAGTACC GTGCGATCGTTCTTCAAGCTGCTA	60
61	ATGGACGTCCTCTTCTTCCACCAGGATCTAGTATGGAGTTTAA GCTTGAGGAGCATTTT	120
121	AACAAGACATTTGTACAGAGAACATAACAGCTGCTCGGAATGCAGCCTTCCCTGCC	180
181	TGGGAGGACTACAGAGCGTAGACGATTTACAATACTTCTGATTGGGCTCTATACA	240
241	TTCGTAAGTCTTCTTGCGTTTATGGGCAATCTACTTATTTTAA TGGCTGTTATGAAAAAG	300
301	CGCAATCAGAAAGACTACAGTGAACCTTCTCATAGGCAACCTGGCCCTTCTCCGACATCTTG	360
361	GTCGTCCGTGTTTGTCTCCCTTTTCAACCCTGACCTCTGTCTTGTGGATCAGTGGATGTTT	420
421	GGCAAGCCATGTGCCATATCATGCCGTTCCCTTCAATGTGTGTCAGTTCTGGTTTCAACT	480
481	CTGATTTTAATATCAATTGCCATTGTCAAGTATCATATGATAAAGCACCCCTATTTCTAAC	540
541	AATTTAACGGCAAAACCATGGCTACTTCCCTGATAGCTACTGTCTGGACACTGGGCTTTGCC	600
601	ATCTGTTCTCCCTCCCAAGTGTTCACAGTCTTGTGGAACCTTAAAGGAGACCTTTGGCTCA	660
661	GCACTGCTGAGTAGCAAAATATCTCTGTGTGAGTCAATGGCCCTCTGATTCTATACAGAAAT	720
721	GCTTTCACAAATCTTTATTGCTAGTGCAGTATATCCTGCCCTCTAGTATGTTTAAACGGTA	780
781	AGTCATACCAGCGTCTGCCGAAGCATAAGCTGTGGATTGTCTCCACAAGAAACAGACTC	840
841	GAAGAAATGAGATGATCAACTTAACCTACAGCCATCCAAAGAGCAGGAAACCAGGCA	900
901	AAACCCCCAGCACTCAAAAGTGGAGCTACTCATTCATCAGAAAGCACAGAGGAGGTAC	960
961	AGCAAGAAGACGGCCTGTGTCTTACCCGCCCCAGCAGGACCCTTCCAGGGGAAGCACCTA	1020
1021	GCCGTTCCAGAAATCCAGCCTCCGTCCGTAGCCAGCTGTGCGCATCCAGTAAGGTCAAT	1080
1081	CCAGGGTCCCAATCTGCTTTGAGGTGAACCTGAAGAAAGCTCAGATGCTCATGAGATG	1140
1141	AGAGTCAAGCGTTCCATCACTAGAAATAAAAGAGATCTCGAAGTGTTTTCTACAGACTG	1200
1201	ACCATACTGATACTCGTGTTCGCCGTTAGCTGGATGCCACTCCAGCTTCCACGTTGGTG	1260
1261	ACTGACTTCAATGATAACTTGATTTCCAATAGGCATTTCAAGCTGGTATACTGCATCTGT	1320
1321	CAC TTGTTAGGCATGATGTCCTGTGTCTAAATCCGATCC TATATGGTTTCTTAATAAT	1380
1381	GGTATCAAAGCAGACTTGAGAGCCCTTATCCACTGCCCTACACATGTCA TGATTCTCTCTG	1440
1441	TGCACCAAAGAGAGAAACGTTGGTAATTGACACATAATTATATACAGAA GTATTCTGGAT	1501

4/38

FIGURE 4

20	F	A	T	K	L	F	T	N	A	S	I	V	L	A	Y	L	I	M	L	V	C	N
40	H	P	Y	K	I	M	S	S	F	G	R	T	R	Q	R	H	V	E	R	V	I	N
60	E	F	L	M	D	W	V	I	G	F	Y	L	N	N	R	K	K	H	Y	H	C	L
80	E	A	G	V	S	Q	L	P	L	T	S	C	E	R	R	G	S	A	F	F	Y	F
100	L	A	I	A	F	D	V	H	T	E	D	V	K	S	H	Q	S	D	V	V	V	G
120	K	N	L	M	A	L	S	K	W	K	S	L	H	K	K	S	P	S	S	H	L	Y
140	F	R	F	L	L	L	V	I	V	L	P	P	S	K	R	P	S	S	R	L	K	L
160	E	A	Y	I	N	V	C	M	T	E	W	L	L	S	I	G	L	E	S	P	F	I
180	M	A	Q	L	G	S	Q	H	A	V	S	I	G	P	F	A	Q	E	R	M	H	P
200	S	A	L	L	I	T	L	Y	I	L	E	Y	C	Q	S	P	S	P	K	W	R	N
220	S	T	D	N	L	L	F	R	L	S	V	Q	S	L	Y	A	R	K	K	S	N	L
240	D	N	D	G	F	T	P	V	F	H	C	V	I	T	S	P	V	V	I	V	S	C
260	Q	N	V	M	N	F	M	I	Y	F	L	L	S	L	W	L	S	E	R	A	I	C
280	H	E	S	F	V	P	I	A	G	V	Y	L	R	N	K	V	A	F	T	F	L	S
300	F	T	G	G	T	S	H	I	H	P	K	L	C	I	Q	C	P	C	I	V	N	M
320	F	V	R	L	T	C	C	S	N	L	S	S	V	M	T	A	N	I	S	L	D	M
340	L	F	Y	L	K	F	M	I	A	P	S	I	S	E	S	T	E	P	R	I	N	G
360	V	T	D	S	Q	L	A	L	T	S	L	T	T	N	P	K	P	V	K	L	F	L
380	D	K	E	V	N	V	K	I	L	C	L	F	H	E	T	K	V	G	V	I	D	L
400	M	N	W	F	R	V	G	L	N	I	A	A	S	E	K	S	A	P	R	T	T	H
420	1	21	41	61	81	101	121	141	161	181	201	221	241	261	281	301	321	341	361	381	401	
440	1	21	41	61	81	101	121	141	161	181	201	221	241	261	281	301	321	341	361	381	401	
456	1	21	41	61	81	101	121	141	161	181	201	221	241	261	281	301	321	341	361	381	401	

5/38

FIGURE 5

1	GTTTCCCTCTGAATAGATTAAATTAAAGTAGTCATGTAATGTTTTTTTTTGGTTGCTGACAA	60
61	ATGTCCTTTTATTCCAAAGCAGGACTATAATAATGGATTTAGAGCTCGACGAGTATTATAAC	120
121	AAGACACTTGGCCACAGAGAAATAACTGCTGCCACTCGGAATCTGATTTCCAGTCTGG	180
181	GATGACTATAAAAGCAGTGTAGATGACTTACAGTATTTTCTGATTGGGCTCTATACATTT	240
241	GTAAGTCTTCTTGGCTTTATGGGGAATCTACTTATTTTAAATGGCTCTCATGAAAAGCGT	300
301	AATCAGAAAGACTACGGTAAACTTCCCTCATAGGCAATCTGGCCCTTTTCTGTATATCTTGGTT	360
361	GTGCTGTTTTGCTCACCTTTCACACTGACGCTGTCTTGTCTGGATCAGTGGATGTTTGGC	420
421	AAAGTCATGTGCCATAATTATGCCCTTTTCTTCAATGTGTGTCTGTTTGGTTTCAACTTTA	480
481	ATTTTAAATCATATTGCCATTGTTCAGGTATCATATGATAAAACATCCCATACTATAAAT	540
541	TTAACAGCAAAACCATGGCTACTTTCTGTATAGCTACTGTCTGGACACTAGGTTTGGCCATC	600
601	TGTTCTCCCTTCCAGTGTTCACAGTCTTGTGGAACTTCAAGAAACATTTGGTTTCAGCA	660
661	TTGCTGAGCAGCAGGTATTTATGTGTTGAGTCATGGCCATCTGATTCATACAGAAATTGCC	720
721	TTTACTATCTCTTTATTGCTAGTTCAGTATATTCTGCCCTTAGTTTGTCTTACTGTAAGT	780
781	CATACAAGTGCTGCAGAAAGTATAAGCTGTGGATTGTCTCCAAACAAGAAAACAGACTTGAA	840
841	GAAAATGAGATGATCAACTTAACCTTTCATCCATCCAAAAGAGTGGGCTTCAGGTGAAA	900
901	CTCTCTGGCAGCCATAAATGGAGTTATTCAATTCATCAAAAACACAGAAAGATATAGC	960
961	AAGAAGACAGCATGTGTGTACCTGTCTCCAGAAAGACCTTCTCAAGAGAACCACTCCAGA	1020
1021	ATACTTCCAGAAAACCTTTGGCTCTGTAAAGATCAGCTCTCTTCATCCAGTAAAGTTCATA	1080
1081	CCAGGGGTCCCCACTTGCTTTGAGATAAAACCTGAAGAAAATTCAGATGTTCAATGAAATG	1140
1141	AGAGTAAACGTTCTGTACAGAAATAAAAGAGATCTCGAAGTGTTTTCTACAGACTG	1200
1201	ACCATACTGATATTAGTATTTGCTGTTAGTTGGATGCCACTACACCTTTTCCATGTGGTA	1260
1261	ACTGATTTTAAATGACAAATCTTATTCAAAATAGGCATTTTCAAGTTGGTGATTTGCATTTGT	1320
1321	CATTTGTTGGCATGATGTCCTGTTGTCTTAATCCAAATTCATATATGGGTTTCTTAATAAT	1380
1381	GGGATTAAAGCTGATTTAGTGTCCTTATACACTGTCTTTCATATGTAAATAATTCTCACTG	1440
1441	TTTACCAAGGAAAGAAC	1457

6/38

FIGURE 6

20	N	20	N
40	W	40	W
60	F	60	F
80	R	80	R
100	V	100	V
120	G	120	G
140	L	140	L
160	N	160	N
180	I	180	I
200	A	200	A
220	A	220	A
240	S	240	S
260	E	260	E
280	K	280	K
300	S	300	S
320	R	320	R
340	I	340	I
360	L	360	L
380	L	380	L
400	V	400	V
420	C	420	C
440	N	440	N
455		455	

1	Y	21	Y
21	V	41	V
41	T	61	T
61	K	81	K
81	L	101	L
101	F	121	F
121	T	141	T
141	N	161	N
161	A	181	A
181	S	201	S
201	I	221	I
221	V	241	V
241	L	261	L
261	S	281	S
281	E	301	E
301	G	321	G
321	T	341	T
341	P	361	P
361	R	381	R
381	T	401	T
401	H	421	H
421	G	441	G

Y	V	T	K	L	F	T	N	A	S	I	V	L	V	Y	S	F	E	R	V	I	N	
Y	P	Y	K	I	M	S	S	F	G	R	T	R	Q	R	H	K	H	Y	H	C	L	
E	F	L	M	D	W	V	I	G	F	Y	L	N	P	R	N	S	V	F	F	Y	F	
D	D	G	L	S	Q	L	P	L	T	S	C	E	G	R	E	S	D	V	L	V	G	
L	S	I	A	F	D	V	H	T	E	D	V	K	S	H	Q	S	S	S	H	L	Y	M
E	N	L	M	A	L	S	K	W	Q	S	L	N	K	K	S	S	N	R	L	K	L	H
L	R	F	L	L	L	V	I	V	L	P	P	S	K	K	P	L	E	S	P	F	I	L
D	T	Y	I	N	V	C	M	T	E	W	L	L	S	I	R	Q	E	R	M	H	P	C
M	A	Q	L	G	S	Q	H	A	V	S	I	G	P	F	E	S	P	K	W	R	N	H
N	A	L	L	I	T	L	Y	I	L	E	Y	C	H	S	P	R	K	K	S	N	L	I
Y	T	D	N	L	L	F	R	L	S	V	Q	S	L	Y	A	V	I	I	V	S	C	L
D	N	D	G	F	T	P	V	F	H	C	V	I	T	S	P	S	E	R	A	I	C	S
Q	N	V	M	N	F	M	I	Y	F	L	L	S	L	W	L	G	F	T	F	L	S	V
K	E	S	F	V	P	I	A	G	V	Y	L	R	N	K	V	F	C	V	V	N	M	L
S	T	S	G	T	S	H	I	H	P	R	L	C	I	H	C	N	T	S	L	D	M	D
Y	A	K	L	T	C	C	S	N	L	S	S	V	M	S	A	E	P	R	I	N	G	A
F	L	Y	L	K	F	M	I	A	P	S	I	S	E	G	T	P	V	K	L	F	L	K
S	T	D	S	Q	L	V	L	T	S	L	T	T	N	S	K	L	G	V	I	D	L	I
M	K	D	V	N	V	K	I	L	C	L	F	H	E	L	K	I	P	R	T	T	H	G

**FIGURE 7A**

**FIGURE 7A**  
**FIGURE 7B**  
**FIGURE 7C**  
**FIGURE 7D**  
**FIGURE 7E**

1	ATGACGTCCTTCTTCC.ACCAGGATTCTAGTAGTGAGTTTAAGCTTG	50
1	. . . . ATGTCTTTTTATTCCAAGCAGGACTATAATGGATTAGAGCTCG	46
51	AGGAGCATTTTAACAAGACATTTGTCAAGAGAACAAATACAGCTGCTGCT	100
47	ACGAGTATTATAACAAGACACTTGCCACAGAGAAATAACTGCTGCCACT	96
101	CGGAATGCAGCCTTCCCTGCCTGGGAGGACTACAGAGGCAGCGTAGACGA	150
97	CGGAATTCTGATTTCCCAGTCTGGGATGACTATAAAGCAGGTAGATGA	146
151	TTTACAATACTTTTCTGATTGGGCTCTATACATTCTGTAAGTCTTCTTGCT	200
147	CTTACAGTATTTTCTGATTGGGCTCTATACATTTGTAAAGTCTTCTTGCT	196
201	TTATGGGCAAATCTACTTATTTTAAATGGCTGTTATGAAAAAGCGCAATCAG	250
197	TTATGGGGAATCTACTTATTTTAAATGGCTCTCATGAAAAAGCGTAATCAG	246





9/38

FIGURE 7C

551	CCCCTCCAGTGTTTCACAGTCTTGTGGAACTTAAGGAGACCTTTGGCTC	600
547	CCCCCTCCAGTGTTTCACAGTCTTGTGGAACTTCAAGAAACATTTGGTTC	596
601	AGCACTGCTGAGTAGCAAAATATCTGTGTTGAGTCATGGCCCTCTGATT	650
597	AGCATTGCTGAGCAGCAGGTAATTATGTGTTGAGTCATGGCCCATCTGATT	646
651	CATACAGAAATTGCTTTCACAAATCTCTTATTGCTAGTGCAGTATATCCTG	700
647	CATACAGAAATTGCCCTTACTATCTCTTATTGCTAGTTCAGTATATTCTG	696
701	CCTCTAGTATGTTTAAACGGTAAGTCATACCAGCGTCTGCCGAAGCATAAG	750
697	CCCTTAGTTTGTCTTACTGTAAAGTCATACAAGTGTCTGCAGAAGTATAAG	746
751	CTGTGGATTGTCCCAACAAGAAACAGACTCGAAGAAATGAGATGATCA	800
747	CTGTGGATTGTCCCAACAAGAAACAGACTTGAAGAAATGAGATGATCA	796
801	ACTTAACCCCTACAGCCATCCAAAAGAGCAGGAACCAAGGCAAAAACCCCC	850
797	ACTTAACCTTTCATCCATCCAAAAGAGTGGGCCCTCAGGTGAAACTCTCT	846









FIGURE 8A

FIGURE 8A  
FIGURE 8B  
FIGURE 8C

Y5h MSFYSKQDYNMMDLELDEYYNKTTLATENNNTAAATRNMSDFPVWDDYKSSVDDL 50  
Y1h MN-STLFSQVENHSHSVHSNFSEKMAQLLA-FENDDCHLPL-AM 39  
Y2h MGPIGAEAADENQTV EEMKVEQYGPQTTPRGELVPPDPEPELIDSTKLI-EV 49  
Y4h MNTSHLLALLLPKSPQGENRSKPLGTPY-NFSEHCQDSV-DV 40

14/38

Y5h QYFLIGLYTFVSLLGFMGNLLILMALMKKRNQKTTVMFLIGNLAFSDILLV 100  
Y1h IFTLALAYGAVIILGVSGNLAALIILKQKEMRNVTNILLVMSFSDLLV 89  
Y2h QVVLIILAYCSIILLLGVIGNSLVIHVVVKFKSMRTVTNFFIANLAVADLLV 99  
Y4h MVFIVTSYSIETVVGVLGMLCLMCVTVRQKEKANVTNLLIANLAFSDFLM 90

Y5h VLFCSPTLLTSVLLDQWMFQKVMCHIMPFLQCVSVLVSTLLILISIAIVRY 150  
Y1h AIMCLPLTFVYTLMDHVVVFGEAMCKLNPFFVQCVSITV\$IFSLVLI AVERH 139  
Y2h NTLCLPFTLLTYTLMGEWKMGPPVLCCHLVPPYAQQGLAVQVSTITLTVIALDRH 149  
Y4h CLLCQPLTAVYTIMDYUIFGETLCKMSAFIQCMSVTVSILSLVLVALERH 140

Y5h HMIKHPISNNLTIANHG YFLIATVUTLGF AICSPLPVFFHSLVELQETFGSA 200  
Y1h QLIINPRGWRPNNRHAYVGI AVIVVLAVASSLPFLIYQVMTDEPFQNVTL 189  
Y2h RCIYHLESKRISKRISFLIIGLAWGISALLASPLAIFREYSLEIIPDFE 199  
Y4h QLIINPTGWKPSISQAYLGI VLIUVIACVLSLPFLANSILENVFHKMHSK 190





**FIGURE 8C**

H	L	F	H	V	V	T	D	F	N	D	N	L	I	S	M	R	R	H	F	K	L	V	Y	C	I	C	H	L	L	G	M	S	C	C	L	N	P	I	L	Y	G	F	L	N	N	G	I	K	A	444
T	I	F	N	T	V	F	D	W	N	H	Q	I	I	A	T	C	N	H	N	L	L	F	L	L	C	H	L	T	A	M	I	S	T	C	V	N	P	I	F	Y	G	F	L	N	K	N	F	Q	R	329
H	A	F	Q	L	A	V	D	I	D	S	Q	V	L	D	L	K	E	Y	K	L	I	F	T	V	F	H	I	I	A	M	C	S	T	F	A	M	P	L	L	Y	G	W	M	S	N	Y	R	K	334	
H	V	F	N	S	L	E	D	W	H	H	E	A	I	P	I	C	H	G	N	L	I	F	L	V	C	H	L	L	A	M	A	S	T	C	V	N	P	F	I	Y	G	F	L	N	T	N	F	K	K	331

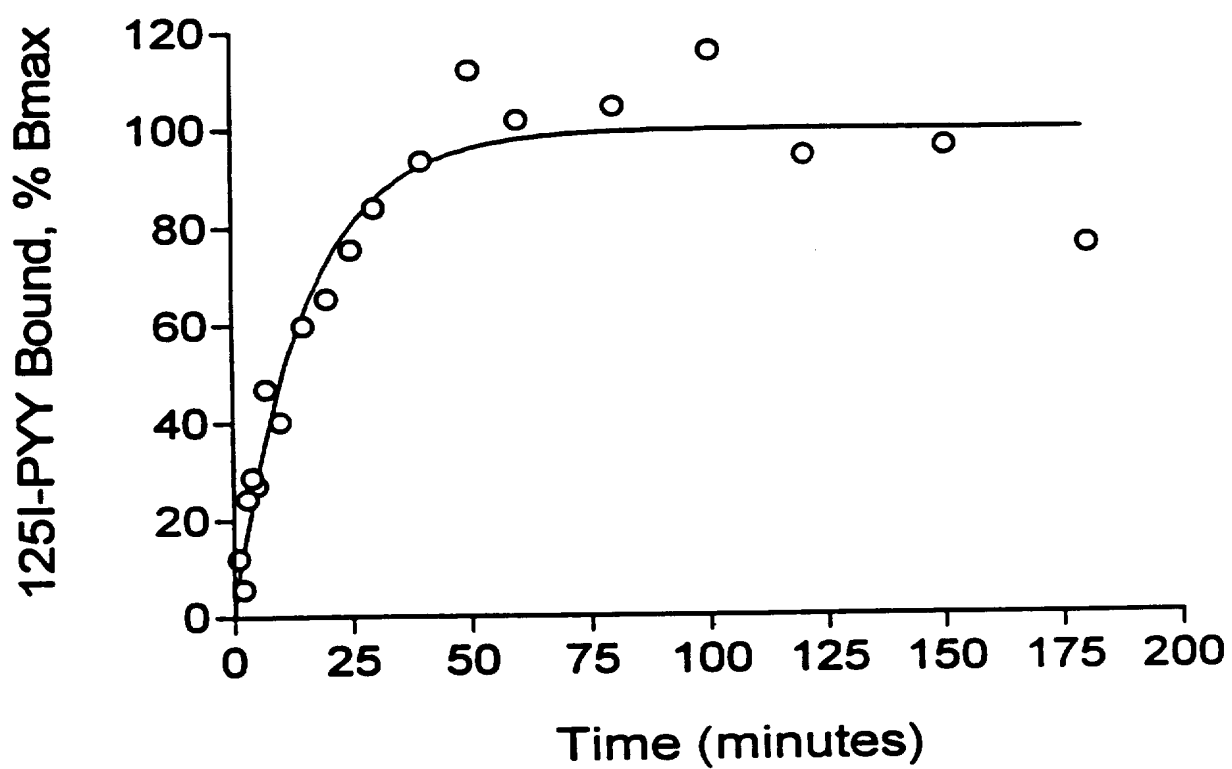
	16/38
455	
379	
381	
375	
Y5h	
Y1h	
Y2h	
Y4h	

Y5h	.	.	.	.	.	455
Y1h	D	N	E	K	I	384
Y2h	.	.	.	.	.	365
Y4h	.	.	.	.	.	375



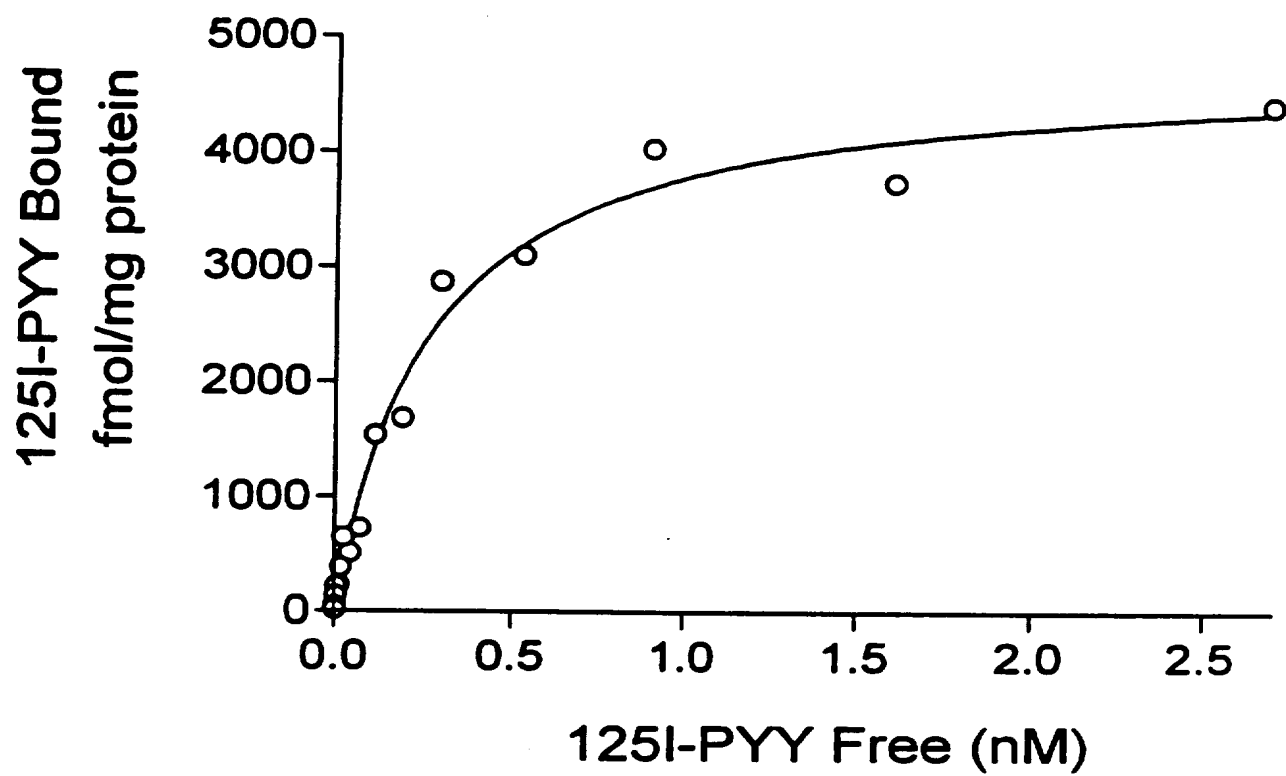
17/38

FIGURE 9



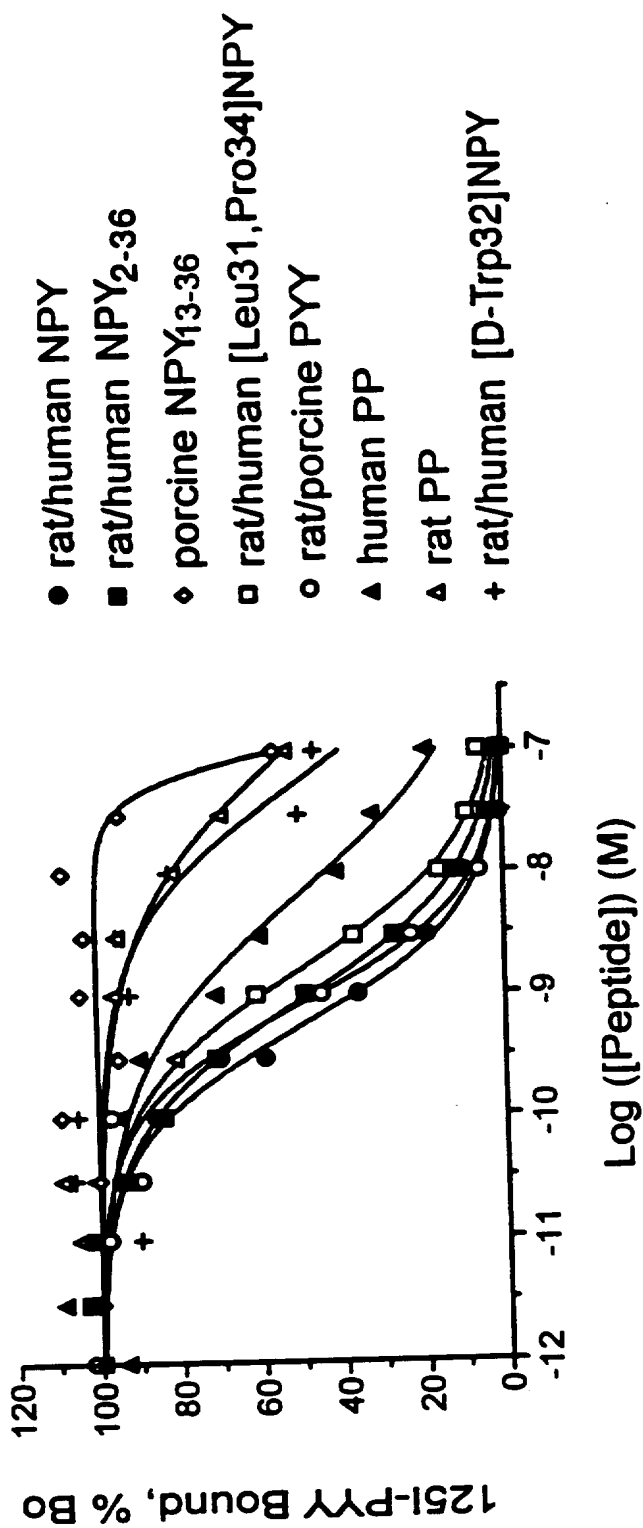
18/38

FIGURE 10



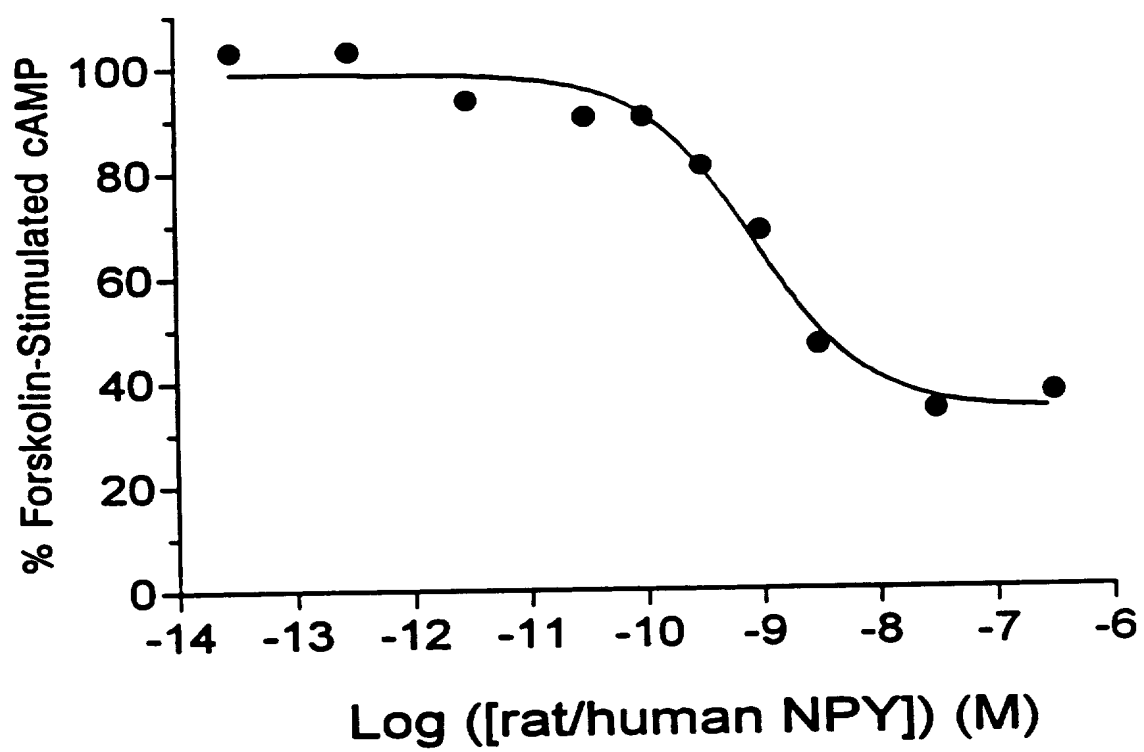
19/38

FIGURE 11

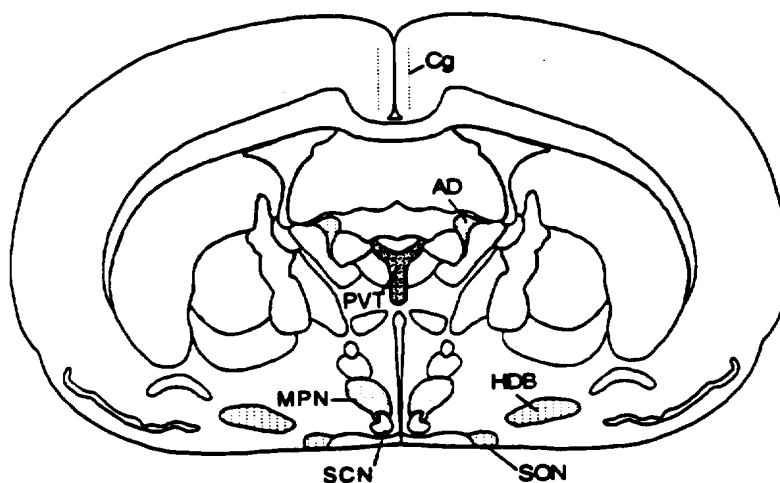
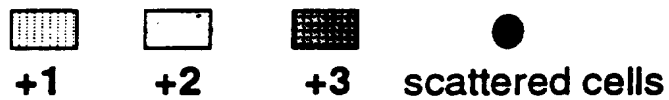
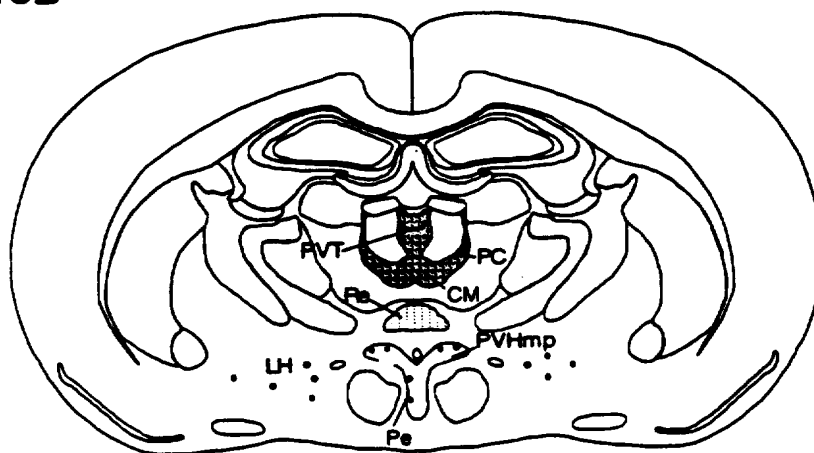
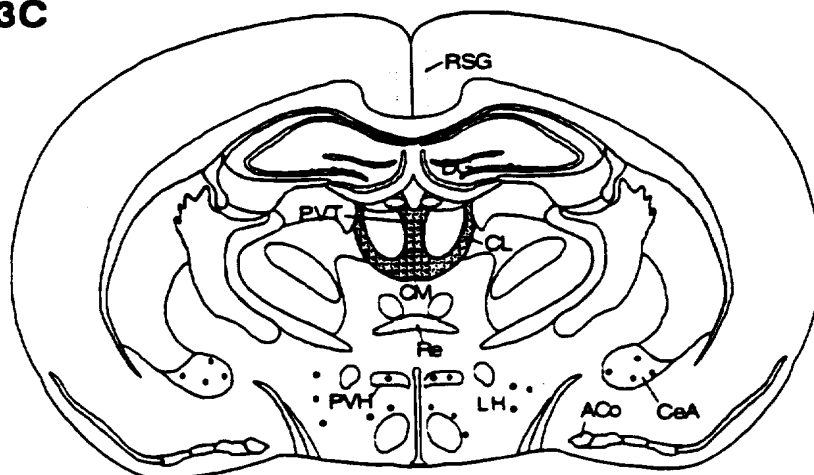


20/38

FIGURE 12



21/38

**FIGURE 13A** Silver grain density:**FIGURE 13B****FIGURE 13C**

22/38

FIGURE 13D

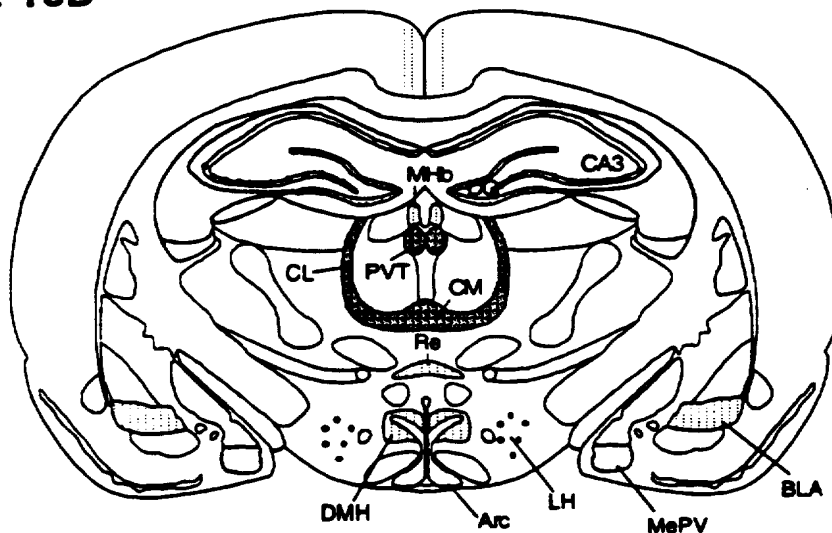


FIGURE 13E

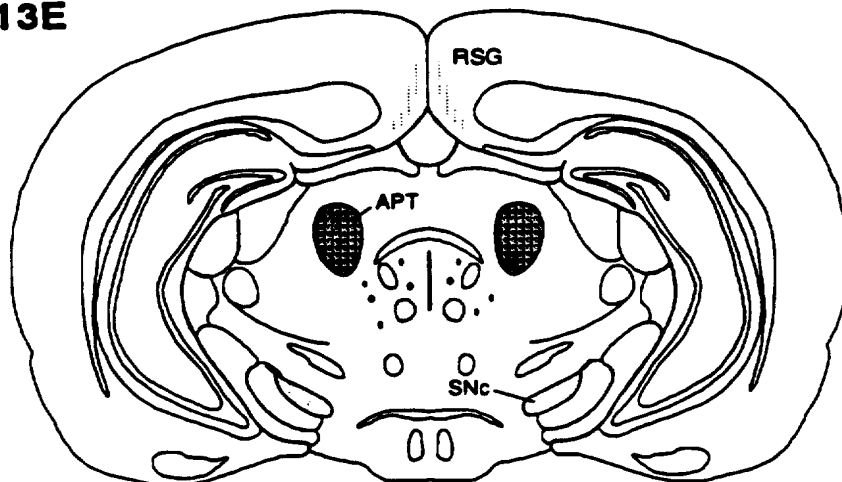
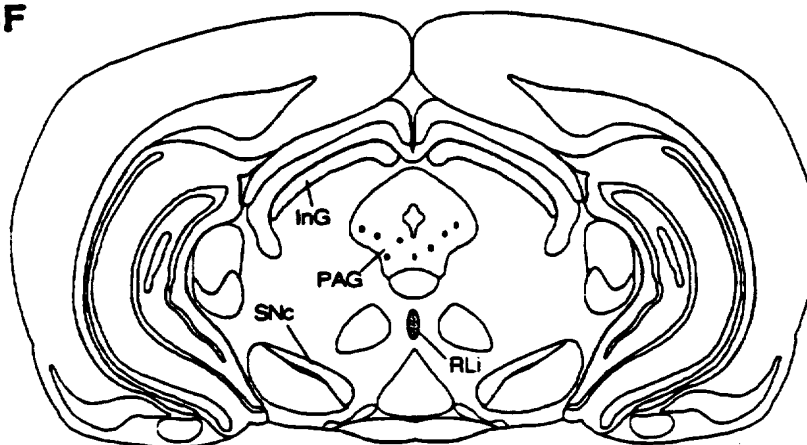


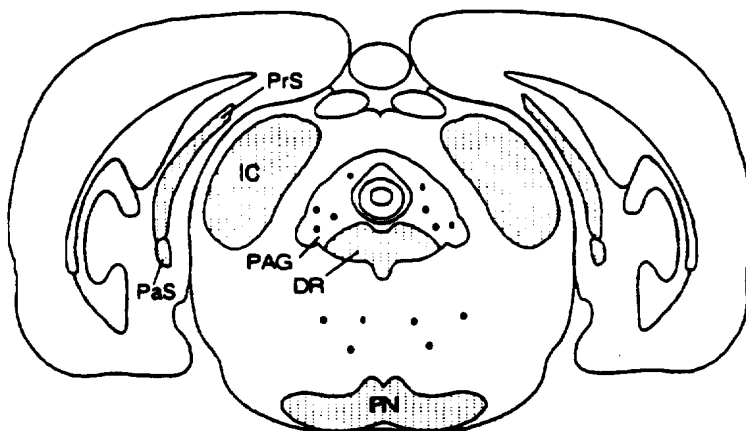
FIGURE 13F



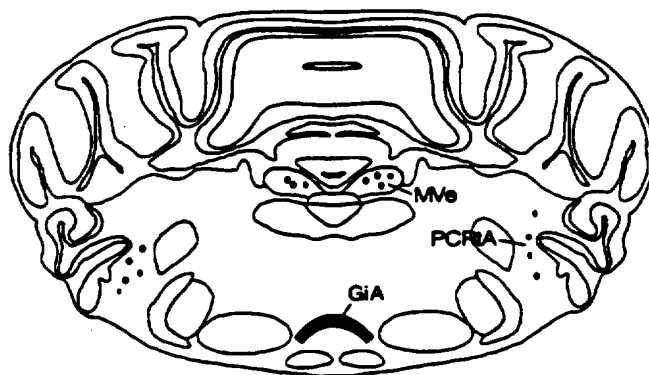


23/38

**FIGURE 13G**



**FIGURE 13H**



24/38

FIGURE 14

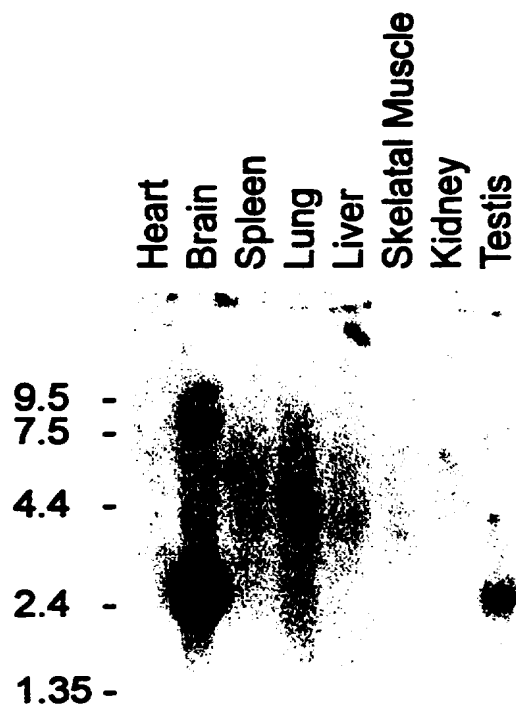
1	TCATGTGTCA	CATTATGCCT	TTTCTTCAAT	GTGTGTCAGT	TCTGGTTTCA	50
51	ACTTTAATTC	TAAATATCAAT	TGCCATTGTC	AGGTATCATA	TGATCAAGCA	51
101	TCCTATATCT	AACAATTAA	CAGCAAACCA	TGGCTACTTC	CTGATTGCTA	150
151	CTGTCTGGAC	ACTAGGTTTT	GCGATTTGTT	CTCCCCCTCC	AGTGTTTCAC	200
201	AGTCTGGTGG	AAC TTCAGGA	AACATTTGAC	TCCGCATTGC	TGAGCAGCAG	250
251	GTATTTATGT	GTTGAGTCGT	GGCCATCTGA	TTCGTACAGA	ATCGCTTTTA	300
301	CTATCTCTTT	ATTGCTAGTC	CAGTATATTC	TTCCCTTGGT	GTGTCTAACT	350
351	GTGAGCCATA	CCAGTGTCTG	CAGGAGTATA	AGCTGCGGGT	TGTCCAACAA	400
401	AGAAAACAAA	CTGGAAGAA	ACGAGATGAT	CAACTTAACT	CTTCAACCAT	450
451	TCAAAAAGAG	TGGGCCTCAG	GTGAAACTTT	CCAGCAGCCA	TAAATGGAGC	500
501	TATTCATTCA	TCAGAAAACA	CAGGAGAAGG	TACAGCAAGA	AGACGGCGTG	550
551	TGTCCTTACCT	GCTCCAGCAA	GACCTCCTCA	AGAGAACCAC	TCAAGAAATG	600
601	TTCCAGAAAA	CTTTGGTTCT	GTAAGAAGTC	AGCATTCCTC	ATCCAGTAAG	650
651	TTCATATACCG	GGTCCCCCAC	CTGCTTTGAG	GTGAAACCTG	AAGAAAACCTC	700
701	GGATGTTTCA	GACATGAGAG	TAAACCGTTC	TATCATGAGA	ATCAAAAAGA	750
751	GATCCCGAAG	TGTTTTCTAT	AGACTAACCA	TACTGATACT	AGTGTTTGCC	800
801	GTTAGCTGGA	TGCCACTACA	CCTTTTCCAT	GTGGTAACCTG	ATTTTAATGA	850
851	CAACCTCAT	TCAAACAGGC	ATTTCAAATT	GGTGATATGC	ATTTGTCAAT	900
901	TGTTAGGCAT	GATGTCCTGT	TGTCCTTAATC	CTATTCTGTA	TGGTTTTCCTC	950
951	AATAATGGGA	TCAAAGCTGA	TTTAATTTC	CTTATACAGT	GTCCTCATAT	1000
1001	GTCATAATTA	TTAATGTTTA	CCAAGGAGAC	AACAAATGTT	GGGATCGTCT	1050
1051	AAAA					

FIGURE 15

1 MCHIMPELQC VSVLVSTLIL ISIAIVRYHM IKHPISNNLT ANHGYFLIAT 50  
51 VWTLGFAICS PLPVFHSIVE LQETFDSALL SSRYLCVESW PSDSYRIAFT 100  
101 ISLLLVQYIL PLVCLTVSHT SVCRSISCGL SNKENKLEEN EMINLTLQPF 150  
151 KKSGPQVKLS SSHKWSYSFI RKHRRRYSKK TACVLPAPAR PPQENHSRML 200  
201 PENFGSVRSQ HSSSKFIPG VPTCFEVKPE ENSDVHDMRV NRSIMRIKKR 250  
251 SRSVFYRLTI LILVFAVSWM PLHLFHVVT D FNDNLISNRH FKLVCICHL 300  
301 LGMMSCCLNP ILYGFLNNGI KADLISLIQC LHMS

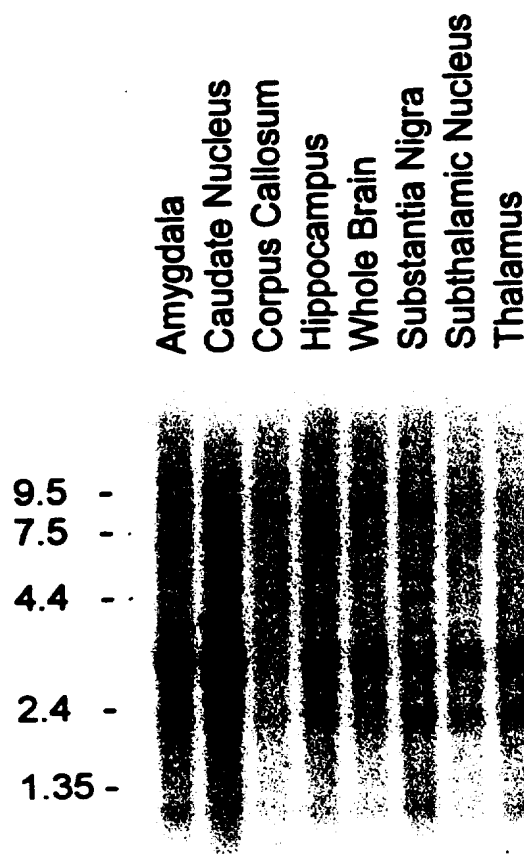
26/38

FIGURE 16A

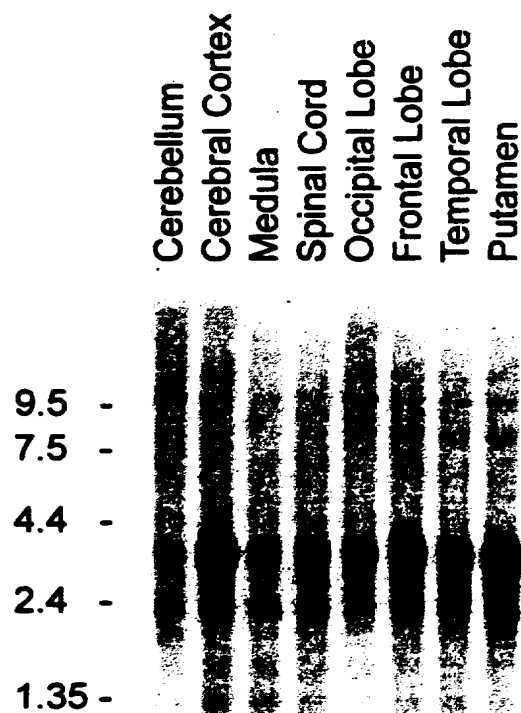


27/38

**FIGURE 16B**

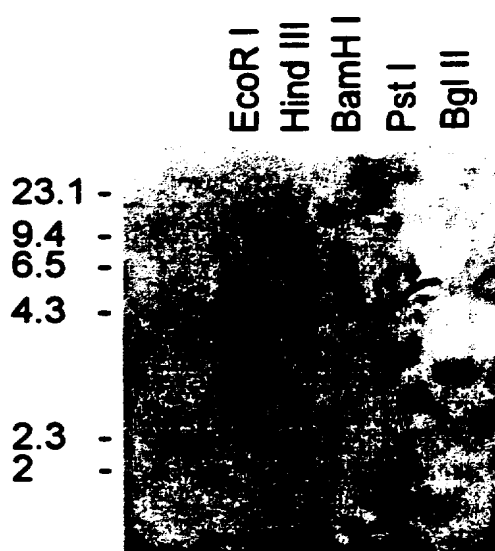


28/38

**FIGURE 16C**

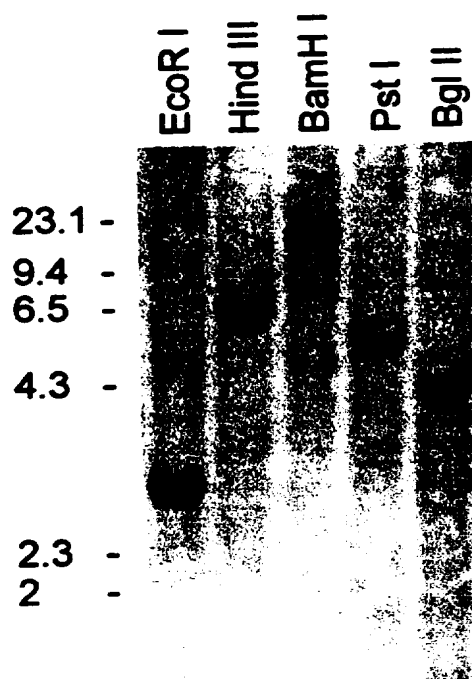
29/38

FIGURE 17A



30/38

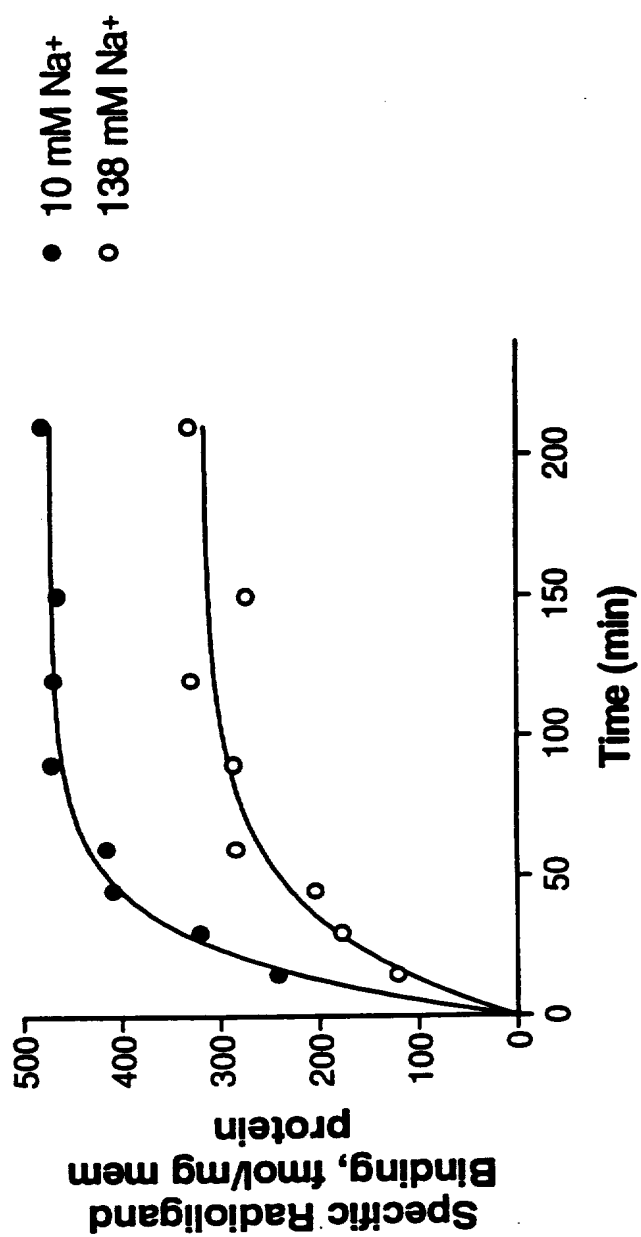
FIGURE 17B





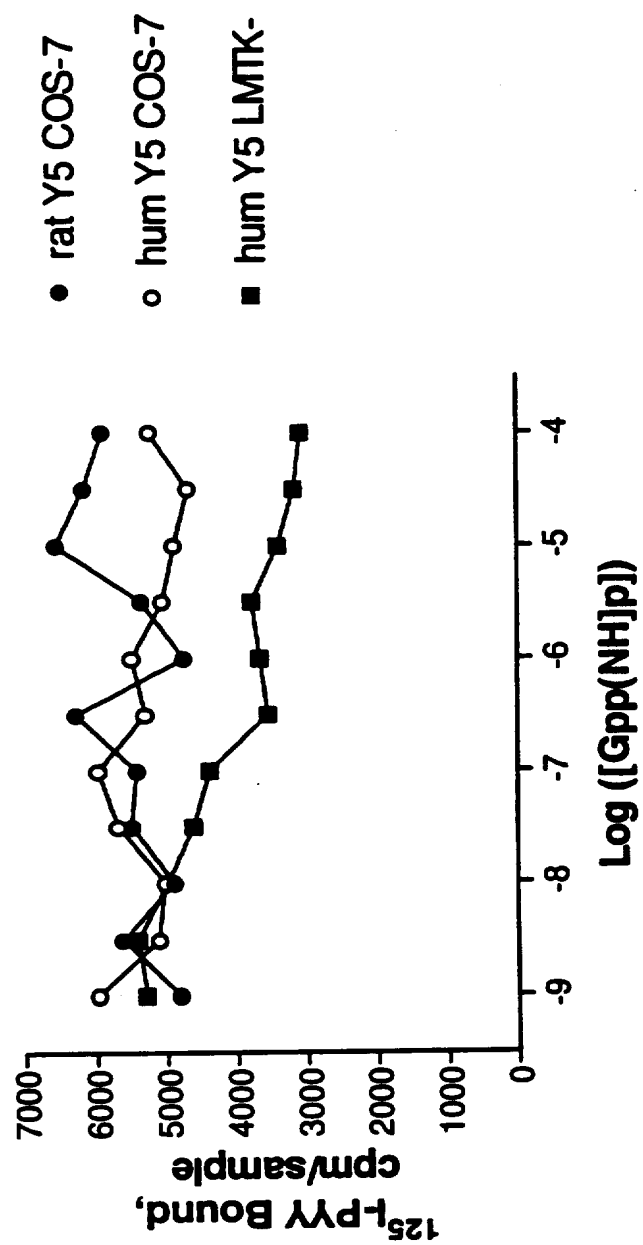
31/38

FIGURE 18



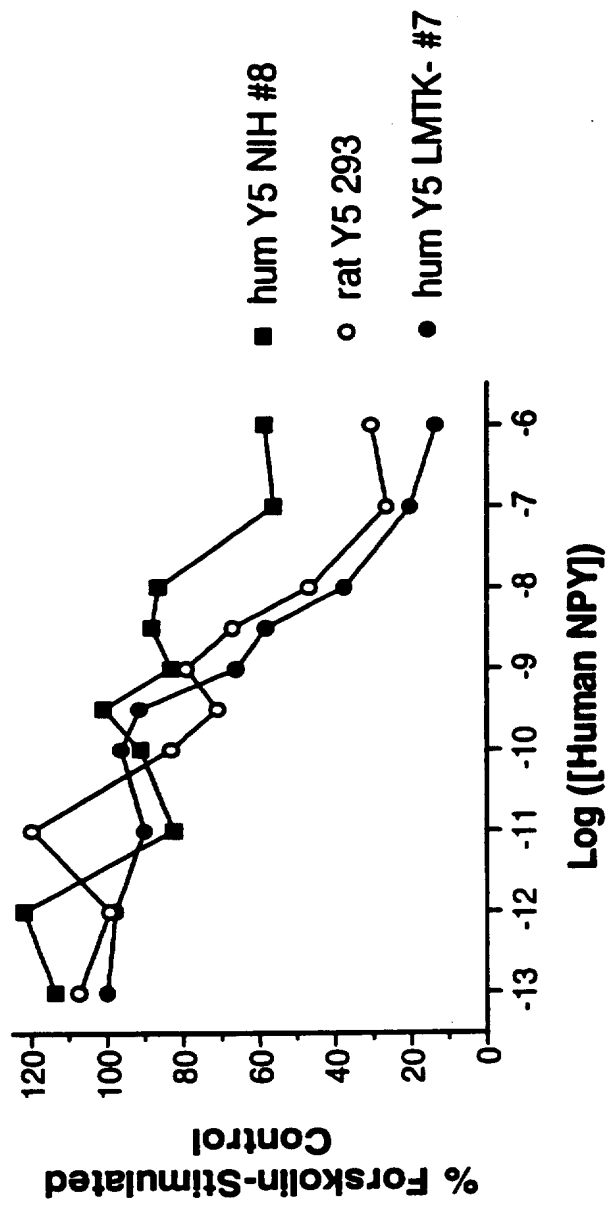
32/38

FIGURE 19



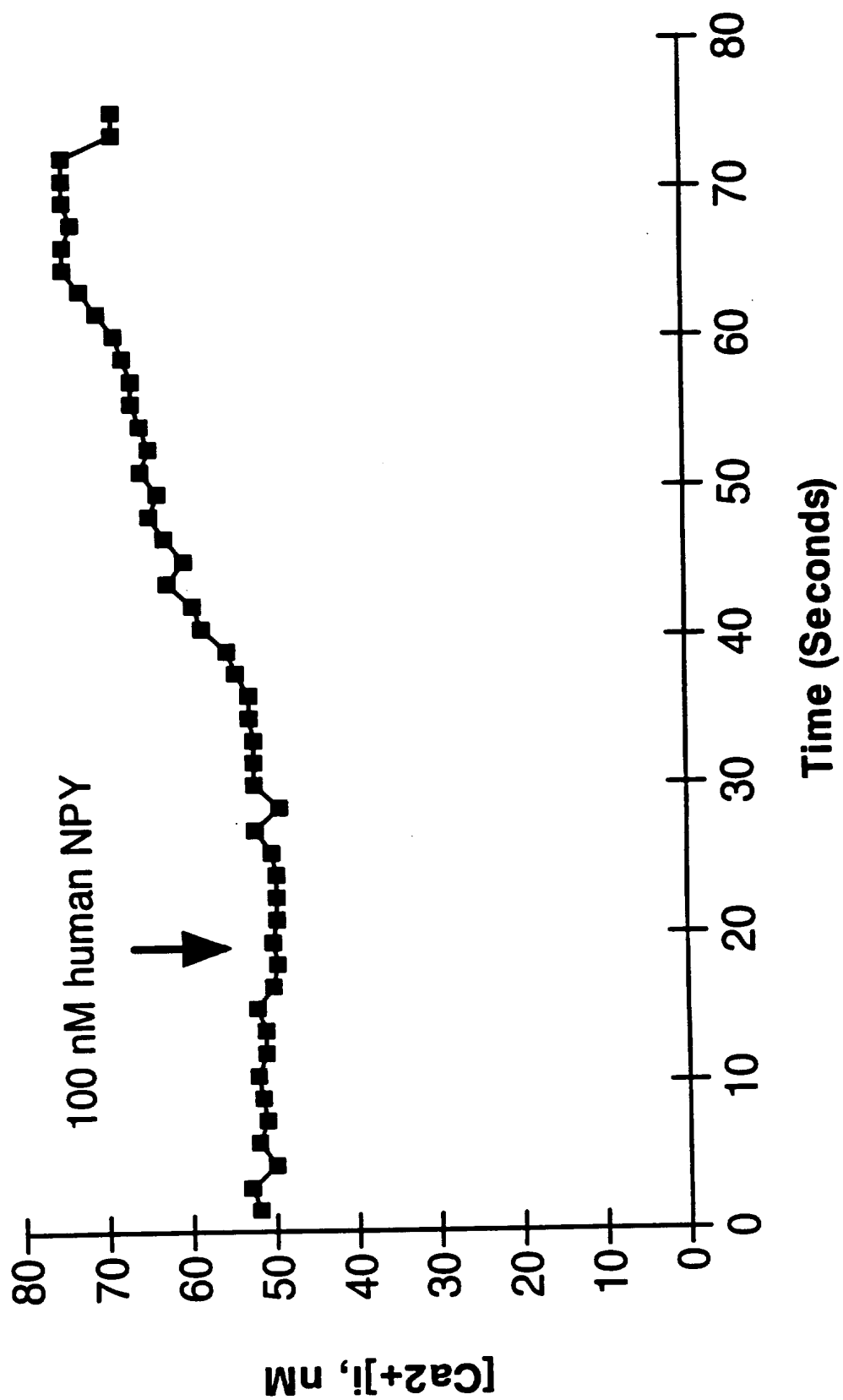
33/38

FIGURE 20



34/38

FIGURE 21A



35/38

FIGURE 21B

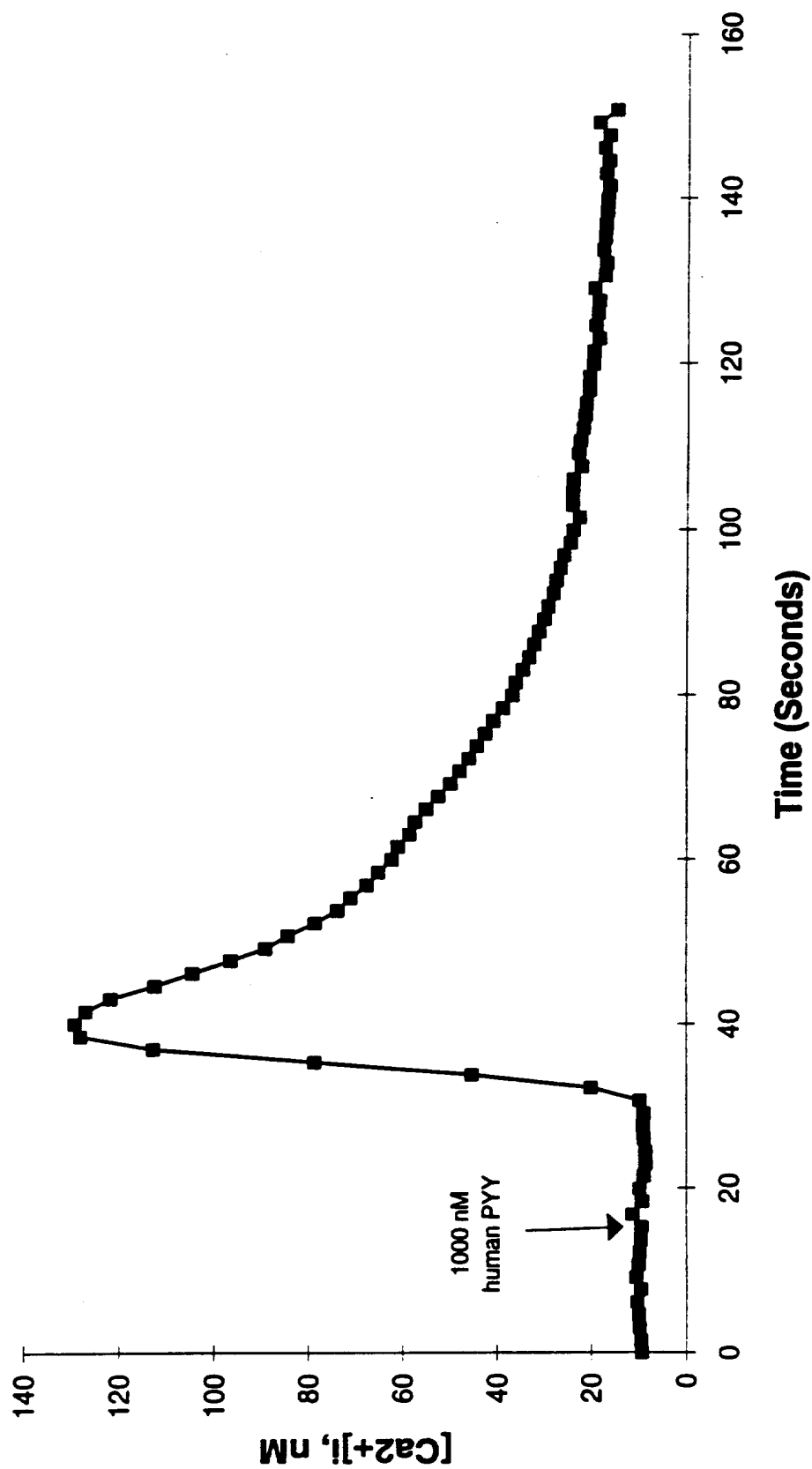
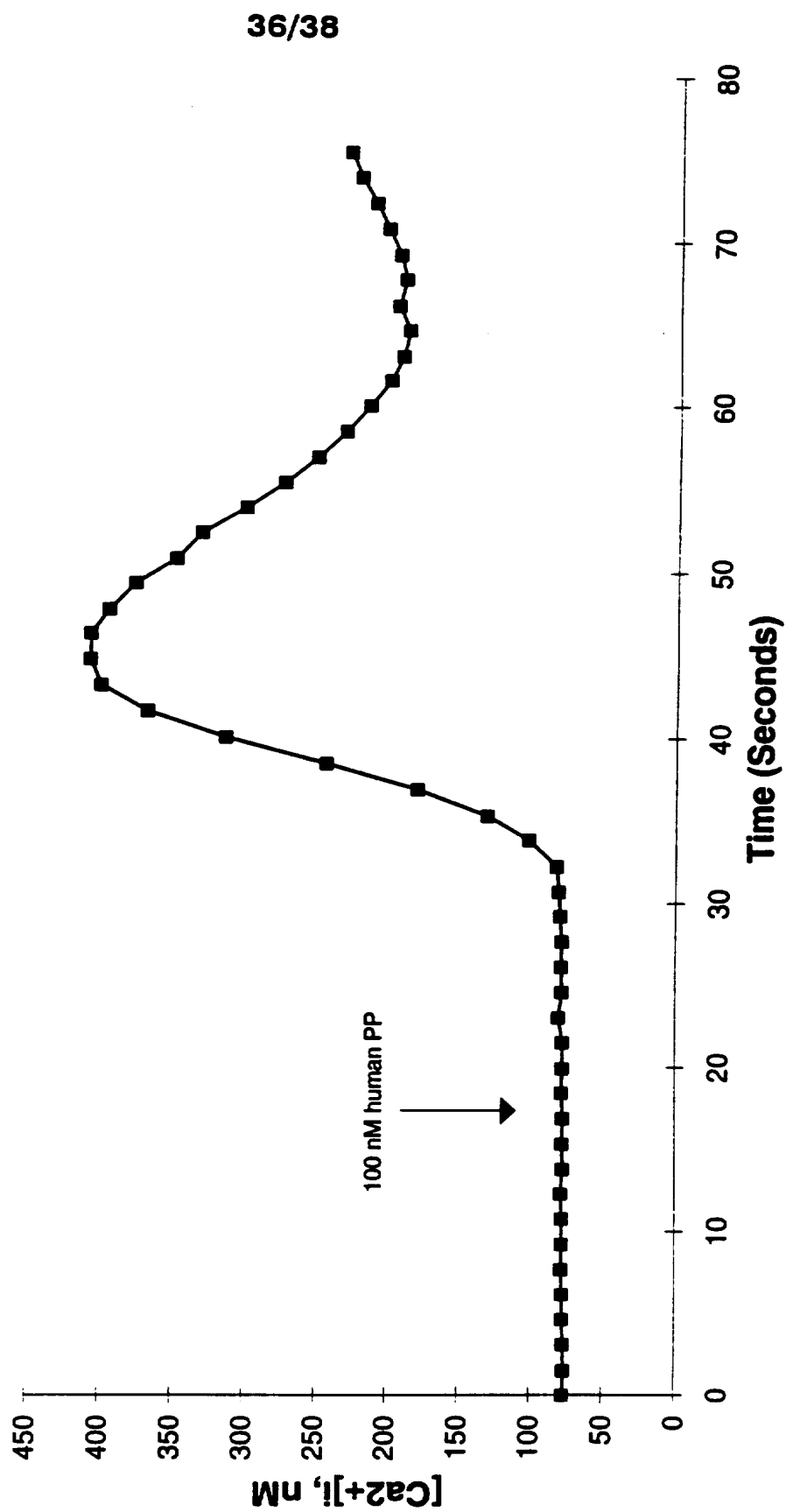
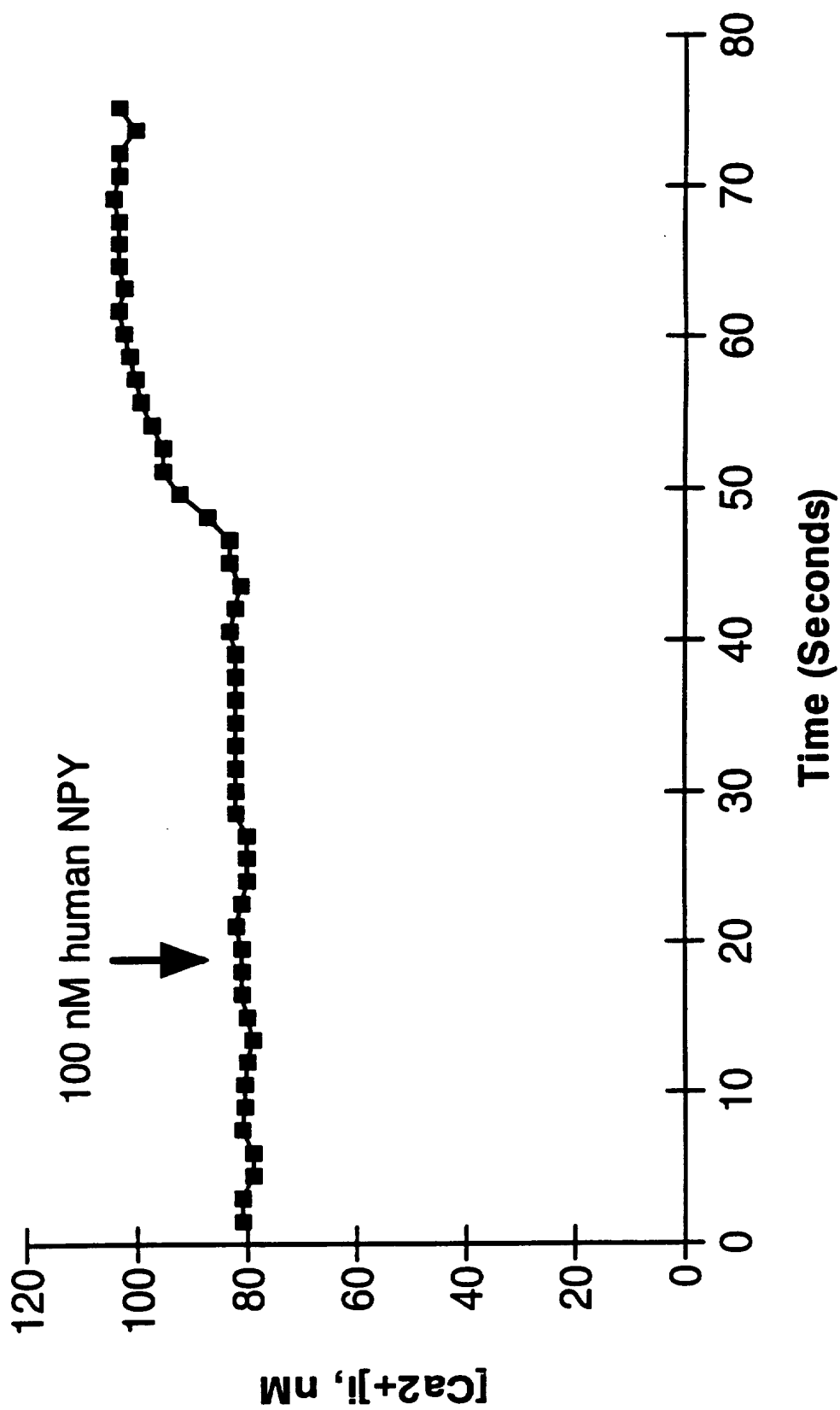


FIGURE 21C



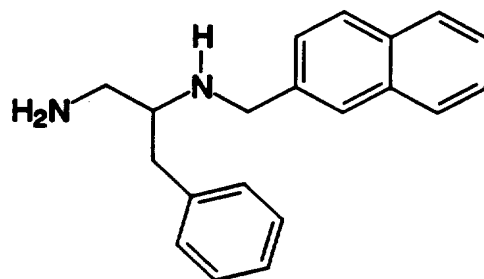
37/38

FIGURE 21D



38/38

FIGURE 22





# INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US95/15646

## A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : Please See Extra Sheet.  
US CL : 514/2; 536/22.1, 23.1, 23.5, 24.3, 24.31, 24.5; 435/91.1, 240.2, 252.3, 320.1  
According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 514/2; 536/22.1, 23.1, 23.5, 24.3, 24.31, 24.5; 435/91.1, 240.2, 252.3, 320.1

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)  
Please See Extra Sheet.

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO, A, 93/09227, (GARVAN INSTITUTE OF MEDICAL RESEARCH) 13 May 1993, see entire document.	1-3, 5-59, and 61-98

☐ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	* T	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
* A		document defining the general state of the art which is not considered to be of particular relevance
* E	* X	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
* L		document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
* O	* Y	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
* P	* A	document published prior to the international filing date but later than the priority date claimed
		document member of the same patent family

Date of the actual completion of the international search  
25 MARCH 1996

Date of mailing of the international search report  
02 APR 1996

Name and mailing address of the ISA/US  
Commissioner of Patents and Trademarks  
Box PCT  
Washington, D.C. 20231  
Facsimile No. (703) 305-3230

Authorized officer  
Patricia A. Duffy  
Telephone No. (703) 308-0196

# INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US95/15646

## Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:  
because they relate to subject matter not required to be searched by this Authority, namely:
2. ☐ Claims Nos.:  
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. ☐ Claims Nos.:  
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

## Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Please See Extra Sheet.

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☒ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:  
1-3, 5-59 and 61-98
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

☐

The additional search fees were accompanied by the applicant's protest.

☒

No protest accompanied the payment of additional search fees.

# INTERNATIONAL SEARCH REPORT

International application No.

PCT/US95/15646

## A. CLASSIFICATION OF SUBJECT MATTER:

IPC (6):

A01N 37/18; A61K 38/00; C07H 19/00, 21/00, 21/02, 21/04; C12P 19/34; C12N 1/20, 5/00, 15/00

## B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

APS, DIALOG, DERWENT WPI, JAPIO, EMBASE, BIOSYS, MEDLINE, CAB ABSTRACTS, PROTEIN AND DNA DATABASES.

search terms: Y5 receptor, neuropeptide Y receptor, disclosed sequences, feeding behavior, bulimia, anorexia, food, consumption, eating, behavior.

## BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be examined, the appropriate additional examination fees must be paid.

Group I, claims 1, 3-6, and 28-42, drawn to a method of modifying feeding behavior and to a method of treating a feeding disorder using a Y5 receptor agonist.

Group II, claims 1-3, 5-17, and 18-27, drawn to a method of modifying feeding behavior and to a method of treating a feeding disorder using a Y5 receptor antagonist.

Group III, claims 43-59, 61-98 and drawn to an isolated nucleic acid molecule encoding a Y5 receptor, a vector comprising said isolated nucleic acid molecule, a probe comprising said isolated nucleic acid molecule, a cell comprising said isolated nucleic acid molecule, and a pharmaceutical composition comprising said isolated nucleic acid molecule.

Group IV, claims 60, 174, and 175, drawn to a purified Y5 receptor protein and method of making.

Group V, claims 99-103 and 109 drawn to an antibody to Y5 receptor protein, a pharmaceutical composition comprising said antibody. Group VI, claims 110-115, drawn to a transgenic animal and first method of use.

Group VII, claims 125, 137-141, 154, 155, 165, 166, 169, and 170, drawn to a Y5 receptor ligand, a pharmaceutical composition, and a drug. Group VIII, claim 156, drawn to a method of detecting expression of a Y5 receptor using the product of Group III.

Group IX, claims 157-160, 166-167 and 171, drawn to a method of treating an abnormality.

Group X, claim 164, drawn to a method of identifying an antagonist using a transgenic animal.

Group XI, claim 168, drawn to a method of identifying an agonist using a transgenic animal.

Group XII, claims 172 and 173, drawn to a method of diagnosing a predisposition to a disorder using a nucleic acid probed for Y5.

Group XIII, claims 116-124, 126-136 and 142-153 are drawn to a method for determining ligand binding to a receptor.

Group XIV, claim 161, drawn to a method of detecting the presence of a receptor on a cell surface. Group XV, claims 162-163, drawn to a method of determining the physiological effects using transgenic animals.

The inventions listed as Groups I-XII do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons. Groups III-VII are products. The products claimed are an isolated nucleic acid molecule encoding a Y5 receptor, a vector comprising said isolated nucleic acid molecule, a probe comprising said isolated nucleic acid molecule, a cell comprising said isolated nucleic acid molecule, a pharmaceutical composition comprising said isolated nucleic acid molecule (Group III), the Y5 protein (Group IV), an antibody to Y5 receptor protein (Group V), a transgenic animal (Group VI), and to a Y5 receptor ligand (Group VII). The products are distinct because they are made by materially

# INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US95/15646

different methods, and have different structures and functional properties. For example, the DNA and vector are comprised of nucleic acids and bind complementary nucleic acids. The protein is comprised of amino acids and binds its ligand. The transgenic animal is an organism and is not a molecule, like the other products. Groups I, II, III, IV, V, VI, VIII, IX, X, XI, XII and XIII-XV are different methods, involving different reagents, steps, and objectives. Note that PCT Rule 13 does not provide for multiple methods within a single application.

This application contains claims directed to more than one species of the generic invention. These species are deemed to lack Unity of Invention because they are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for more than one species to be examined, the appropriate additional examination fees must be paid. The species are as follows:

Group VII, claims 125, 137-141, 154, 155, 165, 166, 169, and 170, drawn to a Y5 receptor ligand, a pharmaceutical composition, and a drug.

The claims are deemed to correspond to the species listed above in the following manner:

Species A, agonist (claims 138, 139, 169, and 170) Species B, antagonist (claims 140, 141, 165, and 166) The following claims are generic: claims 125, 137, 154, and 155.

The species listed above do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, the species lack the same or corresponding special technical features for the following reasons. The species are different reagents and serve different purposes, producing either inhibition (antagonist) or stimulation (agonist) of receptor activity.

This application contains claims directed to more than one species of the generic invention. These species are deemed to lack Unity of Invention because they are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for more than one species to be examined, the appropriate additional examination fees must be paid. The species are as follows:

Group IX, claims 157-160, 166-167 and 171, drawn to a method of treating an abnormality.

The claims are deemed to correspond to the species listed above in the following manner:

Species A, nucleic acid (claim 157)

Species B, antibody (claim 157)

Species C, antagonist (claims 159 and 166)

Species D, agonist (claims 159 and 171)

The following claims are generic: claims 158 and 160.

The species listed above do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, the species lack the same or corresponding special technical features for the following reasons. The species are different classes of reagents made by materially different methods, and have different structures and functional properties, and serve different purposes, producing either inhibition (antagonist) or stimulation (agonist) of receptor activity.